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**PRESENCE OF TAU ASTROGLIOPATHY IN FRONTOTEMPORAL DEMENTIA CAUSED
BY A NOVEL *GRN* NONSENSE (Trp2*) MUTATION**

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Abbreviations:

ARTAG: aging-related tau astrogliopathy

TSA: thorn-shaped astrocytes

GFA: granular or fuzzy tau immunoreactivity in astrocytic processes

Abstract

Frontotemporal lobar degeneration (FTLD) caused by *GRN* mutations is mainly associated with a TDP-43 type A proteinopathy. We present a family with autosomal dominant FTLD caused by a novel *GRN* nonsense mutation (c.5G>A: p.Trp2*) in which the proband's brain also showed prominent glial tauopathy consistent with an aging-related tau astrogliopathy (ARTAG).

Astrocytic tauopathy, 4R(+) and 3R(-) immunoreactive, was characterized by thorn-shaped astrocytes (TSA) present in subpial, subependymal and perivascular areas, and in gray matter; plus granular or fuzzy tau immunoreactivity in astrocytic processes (GFA) in gray matter, either solitary or clustered in different regions. Some neurofibrillary tangles and pretangles, both 3R and 4R (+), were present in the medial temporal lobe but did not exhibit the characteristic distribution of Alzheimer's type pathology.

This 4R-tau ARTAG is likely a co-occurring pathology, although an interaction between progranulin and tau proteins within the neurodegenerative process should not be ruled out.

Key words: *GRN* mutations, neuropathology, tauopathy, ARTAG, astrogliopathy

1. INTRODUCTION

Frontotemporal lobar degeneration (FTLD) is currently subcategorized into three independent pathological phenotypes —tauopathies, TDP-43 proteinopathies, and FUS proteinopathies—, each with consistent genetic correlations. Some cases with FTLD-tauopathy have been found to carry *MAPT* mutations, while FTLD-TDP-43 proteinopathies have been associated with mutations in the *GRN* gene (subtype A), hexanucleotide expansion in *C9ORF72* (subtype B, and A), or mutations in the *VCP* gene (subtype D) (Bang et al., 2015). So far, no known genetic correlations exist for either subtype C of TDP-43 proteinopathy or for FUS-related FTLD.

However, recent reports indicate that *GRN* mutations, which exert their effect through haploinsufficiency and reduced progranulin protein levels, can also be associated with certain tau pathology. Alzheimer's disease tangles and corticobasal degeneration features have been reported in *GRN* mutation carriers (Redaelli et al., 2016). Additionally, Hosokawa et al. (2017) have recently described a significant burden of tauopathy in the form of neuronal pretangles and glial inclusions in the brains of FTLD patients with *GRN* mutations.

We describe the clinical-pathological phenotype of a family with FTLD caused by a novel *GRN* nonsense mutation (c.5G>A: p.Trp2*) predicted to result in complete allele insufficiency. The proband showed neuropathology characterized by TDP-43 type A inclusions, but also extensive glial tauopathy consistent with an aging-related tau astroglipathy (ARTAG) phenotype (Kovacs et al., 2016).

2. METHODS

2.1. Family: This family belongs to a cohort of FTLD cases recruited from the memory clinic of Fundación Jiménez Díaz (Madrid, Spain). The pattern of inheritance was autosomal dominant and two affected relatives, uncle and niece (II.3 and III.1, Figure 1), had been examined. This study was approved by the Research Ethics Committee at Fundación Jiménez Díaz, and informed consent for genetic studies was provided by patients or their surrogates. The spouse

of Case II.3 consented to brain necropsy. Two asymptomatic, at-risk relatives have also been evaluated in the context of genetic counseling and pre-symptomatic testing.

2.2. Genetic study: Whole exome sequencing (WES) of the two affected plus one healthy relative (case II.2) was conducted by the EO EUD Consortium at the VIB-UAntwerp Center for Molecular Neurology (Antwerp, Belgium). Potential pathogenic variants were confirmed by Sanger sequencing (ABI 3730, Applied Biosystems). The segregation pattern was analyzed in the family.

2.3. Plasma progranulin levels: Plasma samples from five family members (two affected, a healthy relative, and two at-risk asymptomatic individuals) had been stored at -80°C . Progranulin levels were analyzed in the Laboratory of Neurology (Fundación Jiménez Díaz) by means of ELISA assay using a commercial kit (AdipoGen Inc, Seoul, South Korea) and following a standardized protocol. Our usual cut-off point to predict *GRN* mutations is <70 ng/ml.

2.4 Brain study: Neuropathological work-up of case II.3 was performed at the Tissue Bank of the Fundación CIEN (Instituto de Salud Carlos III, Madrid, Spain). Five-micrometer-thick sections were cut from formalin-fixed and paraffin-embedded tissue obtained from multiple brain regions. Sections were stained with haematoxylin-eosin (H&E), Gallyas silver staining, and immunostained using monoclonal (mc) and polyclonal (pc) primary antibodies for beta-amyloid (mc, clone 6F/3D, DAKO), tau (including: phospho-tau Ser202/Thr205, mc, clone AT8, and phospho-tau Thr212/Ser214, mc, clone AT100, both from Thermo Fisher Scientific; anti-tau 3-repeat isoform RD3, mc, clone 8E6/C11, and anti-tau 4-repeat isoform RD4, mc, clone 1E1/A6, both from Millipore), TDP-43 (pc, Proteintech; and mc, clone 2E2-D3, Abnova), alpha-synuclein (mc, clone KMS1, Leica Biosystems), ubiquitin (pc, Z0458, DAKO), and p62 (pc, Thermo Fisher Scientific). Presence of neuronal tau deposits (neurofibrillary tangles and pretangles) and different tau-immunoreactive astrocytic inclusions was recorded, along with their location (subpial, subependymal, perivascular, white matter, and gray matter). The

pathologic profile of the glial tauopathy was defined in terms of the morphological type, location, and intensity of tau (+) cell inclusions, according to the criteria of Kovacs et al. (2016).

3. RESULTS

3.1. Clinical phenotypes: The two affected cases examined had manifested at a presenile age as behavioral (II.3) and nonfluent aphasia (III.1) variants of FTL, respectively (Figure 1). The first symptoms presented by case II.3 consisted of compulsive behaviors including hyperphagia, disinhibition, irritability, decreased verbal fluency, and ideomotor apraxia. CT scan and SPECT with HMPAO showed selective left temporal atrophy/hypoperfusion. Four years after onset the patient had developed severe mixed aphasia and parkinsonism in right extremities. A CT scan obtained one year before his death showed severe frontotemporal (left predominant) atrophy (Figure 1). Case III.1 began with apathy and decreased verbal fluency. After two years, she was aphasic and exhibited prominent dysexecutive syndrome. Currently —55 years of age— she is mute with right parkinsonian/corticobasal features, though no behavioral problems have developed. Her cerebral MRI scan and SPECT with HMPAO also reveal asymmetric frontotemporal (left and frontal predominant) neurodegeneration.

3.2. Whole exome sequencing: Analysis of the WES data was directed towards known neurodegenerative disease genes. This identified a *GRN* nonsense mutation, c.5G>A: p.Trp2* (confirmed by Sanger sequencing), carried by both patients and absent from the healthy elderly relative. This variant was absent from public control databases (dbSNP, ExAc, and the AD&FTD Mutation Database), and its predicted pathogenicity according to CADD score (Kircher et al., 2014) was 36 (> 20 suggests pathogenicity). There were no *MAPT* mutations in the affected relatives, and screening for *C9ORF72* hexanucleotide expansion was also negative. All three relatives were heterozygous H1/H2 for the *MAPT* haplotype. One of the two at-risk relatives (III.3) turned out to be a carrier.

3.3. Plasma progranulin levels: Pathogenicity of the *GRN* mutation was further supported by reduced plasma progranulin levels in the two affected patients and the asymptomatic carrier (all below 40 ng/ml, see values in Figure 1), compared to levels in the two non-carrier relatives (≥ 130 ng/ml).

3.4. Neuropathology (case II.3): Macroscopic evaluation showed severe bilateral frontotemporal and inferior parietal atrophy (left predominant) with an unfixed brain weight of 1053 gr. On H&E near complete neuronal loss with neuropil spongiosis was observed in the orbitofrontal, temporal, and insular cortices. Entorhinal cortex and amygdala showed severe neuronal loss and gliosis. Hippocampal sclerosis was observed including subiculum and CA1-2, with relative sparing of dentate gyrus.

Beta-amyloid was found in very scattered diffuse plaques present in CERAD cortical areas. There was no amyloid angiopathy. α -synuclein immunostaining was negative all throughout the brainstem and cortex.

TDP-43 immunoreactivity (Figure 2) was present in cortex and characterized by short dystrophic neurites and neuronal cytoplasmic inclusions concentrated primarily in layer 2, consistent with type A TDP-43 proteinopathy (Mackenzie et al., 2011). There were also TDP-43 immunoreactive neurons in dentate gyrus, in the hippocampal sclerosis segment, and a few in the striatum and nucleus basalis of Meynert. No intranuclear TDP-43 immunoreactive inclusions were observed. No tau-negative, p62-immunoreactive inclusions were observed in any cortical or subcortical regions.

Tau accumulation was characterized by abundant phospho-tau (+) inclusions; these were predominantly astrocytic and, less frequently, neuronal tau deposits. Thorn-shaped astrocytes (TSA) and granular/fuzzy tau immunoreactivity in processes of astrocytes (GFA) —the two major cytomorphologies of the recently characterized ARTAG phenotype (Kovacs et al., 2016)— were detected by AT8 and AT100 and a 4-repeat (4R) tau antibody, and were negative

for the 3R isoform antibody. TSA were clearly Gallyas positive, whereas GFA showed inconsistent slightly stained granular deposits in the perikaryon. The neuropil contained a moderate number of slender positive threads. Different ARTAG types, according to the brain region involved, were found (Figure 3): subpial (A), subependymal (B), perivascular (C), gray matter (D, E), and white matter (F) locations. No inclusions characteristic of other primary tauopathies were found. The Table summarizes the regional distribution and intensity of involvement by astroglial and neuronal tau pathology following the Kovacs et al. (2016) evaluation strategy for ARTAG. TSA and GFA coexisted in multiple regions, with a high presence found in the anterior medial temporal lobe (MTL). TSA were observed mainly in subpial localization, the deep segments of gyri, and in subependymal and perivascular areas, both in the gray and white matter, whereas GFA were present either in the white matter or clustered in different gray matter regions of the brain. Neuronal tau pathology was detected as pretangles and NFTs in different brain regions, with a higher presence in the MTL, not exhibiting a characteristic distribution for Alzheimer's type pathology. These neuronal inclusions were immunoreactive for both 3R and 4R tau antibodies.

DISCUSSION

The presence of this ARTAG phenotype in the brain of a patient with TDP-43 type A FTLD pathology associated with a *GRN* nonsense mutation suggests two possible scenarios: the co-occurrence of two independent processes, or a relationship underlying both pathologies.

The first scenario, concomitance or co-occurrence of two neuropathologies, appears the most likely in light of recent reports showing the high prevalence of concomitant proteinopathies in brains of cases with neurodegenerative diseases, which increases with the age at death (Robinson et al., 2018). In particular, accumulation of neuronal tau as neurofibrillary tangles seems to be almost universal in brains with all other primary neurodegenerative pathologies

(Robinson et al., 2018). Recent studies suggest that in cases with pathologically confirmed FTLD, multiple neuronal pathologies appear with equal prevalence in young and elderly cases (Tan and Halliday, 2017). Furthermore, previous case reports describe concomitant tau pathology in *GRN* mutation carriers (Leverenz et al., 2007; Hosokawa et al., 2017), and suggest that *GRN* insufficiency could be associated with multiple proteinopathies, including TDP-43, tau, and alpha-synuclein deposition.

Most of these studies concentrate on the co-occurrence of neuronal inclusions and have paid less attention to the presence of glial pathology. However, the report by Hosokawa et al. (2017) is highly relevant to our description because the authors examine free-floating sections of four cases with three different *GRN* mutations and find tau immunoreactivity in astrocytes in all of them (massive in one). In particular, the massive astroglipathy described in that case resembles our studied brain in terms of regional distribution and staining characteristics, although the astrocyte morphology is reported as “bush-like” rather than “thorn-shaped”. Hosokawa et al. (2017) further studied paraffin-embedded sections of another nine cases with *GRN* mutations and find some AT8 immunoreactivity in the majority, but to a lesser extent. The authors emphasize that the sensitivity to detect tau deposition is higher in free-floating tissue compared to paraffin-embedded sections.

A second scenario could be that the presence of ARTAG would be related to the *GRN*-mutation condition and progranulin insufficiency. Some research studies support that progranulin reduction may contribute to abnormal tau deposition. A SNP *GRN* variant (rs5848) has been associated with the risk of AD (Lee et al., 2011) and correlates with increased CSF tau levels (Takahashi et al., 2017). In animal models, a reduction in progranulin in tau transgenic mice (P301L tau/*GRN*-deficient mouse model) is associated with increased tau accumulation (Hosokawa et al., 2015).

The mechanisms by which *GRN* insufficiency may be able to contribute to tauopathy could be

speculated in terms of the complex roles of progranulin in the brain. Progranulin is a pleiotropic glycoprotein with growth factor-like properties and neuronal effects including regulation of neurite outgrowth, synapse biology, stress response and lysosomal functions (Petkau and Leavitt, 2014). *GRN*-deficient mouse models show an aging pathological phenotype characterized by increased astrocytic and microglial proliferation and highly accelerated deposition of lipofuscin (Petkau et al., 2016, Yin et al., 2010). Progranulin contributes to long-term cell survival and decreasing *GRN* expression likely impairs cell resistance to aging and other degenerative diseases. Particularly, *GRN*-deficient hippocampal cells *in vitro* are more vulnerable to direct cytotoxicity of macrophages and microglia as well as to deprivation of oxygen and glucose (Yin et al., 2010). It is possible that a tau astroglial pathology develops because astrocytes are more vulnerable than neurons, as has been shown in preclinical corticobasal degeneration cases, where a tau astroglial pathology seems to be a prodromal stage before the advance of extensive neuronal tauopathy (Ling et al., 2016). Furthermore, it has been suggested that some ARTAG features (GFA) can be premature stages of the astrocytic inclusions characterizing the primary aging related tauopathies (PART) (Kovacs et al., 2017; Kovacs et al., 2018).

Finally, the contribution of the glial tauopathy to the clinical phenotype has yet to be defined. Some clinicopathological studies suggest that circumscribed ARTAG may present clinically with focal symptoms like aphasia (Munoz et al., 2007), whereas in cases with widespread pathology, dementia with or without parkinsonism might be the clinical presentation (Kovacs et al., 2013).

In conclusion, this family adds support to the notion that genetic variations in *GRN* are associated with a variety of neurodegenerative proteinopathies including astroglial tauopathy.

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Figure 1. Family tree including *GRN* Trp2* mutation status (M+ black: affected; M+ gray: asymptomatic carrier; WT: wild type), clinical phenotypes, and progranulin plasma levels (PgrL in ng/ml). CT scan images of the proband (II.3) one year before his death.

Figure 2. TDP-43 immunoreactive inclusions in the cerebral cortex, mainly in layer 2, consistent with TDP-43 type A proteinopathy **(A)**. TDP-43 immunoreactive inclusions were also present in dentate gyrus **(B)**.

Figure 3. Tau immunoreactive inclusions: Different ARTAG types were found in different locations: subpial (A), subependymal (B), perivascular (C), gray matter (D, E), and white matter (F) locations. TSA and GFA were detected by AT8 (G), AT100 (I), and 4R-tau (H) antibodies, and were negative for the 3R isoform antibody. Neuronal inclusions (G, H) were identified in the medial temporal lobe, showing immunoreactivity for both 3R and 4R tau antibodies. Scale bar: 150 μ m. Bottom left: antibody used.

Table. Regional distribution and intensity of involvement (color gradient) by astroglial and neuronal tau pathology, according to the Kovacs' *et al.* evaluation strategy for ARTAG. TSA and GFA coexisted in multiple regions, with a higher presence in the anterior medial temporal lobe (MTL). Neuronal tau pathology was detected as pretangles and NFTs in different brain regions, with higher presence in the MTL.

	Step 1	step 2	Step 3					
	Type	Major anatomical	Severity in detailed regional distribution and extent of ARTAG					
ASTROCYTES	Subpial	MTL	Inf. Temporal Gy	Hippocampus	Entorhinal cortex	Amygdala		
		Lobar	Frontal	Cingular cortex	Insular cortex	Parietal	Occipital	Temporal
		Subcortical				basal forebrain	Hypothalamus	
		Brainstem	Mesencephalon			Pons	medulla Obl.	
	Subependymal	MTL	Temporal horn			Amygdala		
		Lobar	LV: Frontal horn				LV: Occipital horn	
		Subcortical	LV: Caudate	3V: Thalamus				Hypothalamus
		Brainstem	Aq/Mesencephalon	Aq/Pons	Aq/Medulla obl.			
	perivascular	MTL	Inf. Temporal Gy	Hippocampus	Entorhinal cortex	Amygdala		
		Lobar	Frontal	Cingular cortex	Insular cortex	Parietal	Occipital	Temporal
		Subcortical	Accumbens	Caud/Put	Globus pallidus	basal forebrain	Hypothalamus	
		Brainstem	Mesencephalon			Pons	Medulla obl.	
	White matter	MTL	Inf. Temporal Gy	Hippocampus	Entorhinal cortex	Amygdala		
		Lobar	Frontal	Cingular cortex	Insular cortex	Parietal	Occipital	Temporal
		Subcortical	Internal capsule			Subinsular	Pencil fibers	
		Brainstem	Cerebral pedunculi			Pyramids	Midline	
Gray matter	MTL	Inf. Temporal Gy	Hippocampus	Entorhinal cortex	Amygdala			
	Lobar	Frontal	Cingular cortex	Insular cortex	Parietal	Occipital	Temporal	
	Subcortical	Accumbens	Caud/Put	Globus pallidus	basal forebrain	Hypothalamus		
	Brainstem	Substantia nigra			Pons	Medulla obl.		
NEURONS	Gray matter	MTL	Inf. Temporal Gy	Hippocampus	Entorhinal cortex	Amygdala		
		Lobar	Frontal	Cingular cortex	Insular cortex	Parietal	Occipital	Temporal
		Subcortical	Accumbens	Caud/Put	Globus pallidus	basal forebrain	Hypothalamus	
		Brainstem	Substantia nigra			Pons	Medulla obl.	