

This item is the archived peer-reviewed author-version of:

Exercise training to reduce cardiovascular risk in patients with metabolic syndrome and type 2 diabetes mellitus : how does it work?

Reference:

Kränkel Nicolle, Bahls Martin, van Craenenbroeck Emeline, Adams Volker, Serratos Luis, Ekker Solberg Erik, Hansen Dominique, Dörr Marcus, Kemps Harelde.-
Exercise training to reduce cardiovascular risk in patients with metabolic syndrome and type 2 diabetes mellitus : how does it work?
European journal of preventive cardiology - ISSN 2047-4881 - (2018), p. 1-8
Full text (Publisher's DOI): <https://doi.org/10.1177/2047487318805158>
To cite this reference: <https://hdl.handle.net/10067/1571210151162165141>

Exercise training to reduce cardiovascular risk in patients with metabolic syndrome and type 2 diabetes mellitus: How does it work?

Nicolle Kränkel^{1,2,*}, Martin Bahls^{3,4}, Emeline M. Van Craenenbroeck⁵, Volker Adams⁶, Luis Serratos^{7,8}, Erik Ekker Solberg⁹, Dominique Hansen^{10,11}, Marcus Dörr^{3,4}, Hareld Kemps¹²

- 1 Charité – Universitätsmedizin Berlin, Klinik für Kardiologie, Campus Benjamin Steglitz, Berlin, Germany
- 2 DZHK (German Centre for Cardiovascular Research), partner site Berlin, Germany
- 3 University Medicine Greifswald, Department of Internal Medicine B, Greifswald, Germany
- 4 DZHK (German Centre for Cardiovascular Research), partner site Greifswald, Germany
- 5 Department of Cardiology, Antwerp University Hospital, Antwerp, Belgium
- 6 Department of Molecular and Experimental Cardiology, TU Dresden, Heart Center Dresden, Dresden, Germany
- 7 Hospital Universitario Quironsalud, Madrid, Spain
- 8 Ripoll & De Prado Sport Clinic, FIFA Medical Centre of Excellence, Spain
- 9 Diakonhjemmet hospital, Department of Medicine, Oslo, Norway
- 10 Hasselt University, Faculty of Medicine and Life Sciences, Diepenbeek, Belgium
- 11 Heart Centre Hasselt, Jessa Hospital, Hasselt, Belgium
- 12 Department of Cardiology, Máxima Medical Centre, Veldhoven, The Netherlands

*** corresponding author's full address:**

PD Dr. rer. nat. Nicolle Kränkel

Charité – Universitätsmedizin Berlin, Campus Benjamin Franklin

Klinik für Kardiologie

Hindenburgdamm 30

12203 Berlin

phone: +49 (0)30 450 522 246

fax: +49 (0)30 450 513 999

email: nicolle.kraenkel@charite.de

word count: 4558

Sources of funding related to this work:

German Centre for Cardiovascular Research: NK, MB, MD

EvC: Fund for Scientific Research Flanders

none: VA, LS, EES, DH, HK

1. Abstract

Metabolic syndrome (MetS) - a clustering of pathological conditions, including abdominal obesity, hypertension, dyslipidemia and hyperglycaemia - is closely associated with the development of type 2 diabetes mellitus (T2DM) and a high risk of cardiovascular disease.

A combination of multigenetic predisposition and lifestyle choices accounts for the varying inter-individual risk to develop MetS and T2DM, as well as for the individual amount of the increase in cardiovascular risk in those patients. A physically active lifestyle can offset about half of the genetically mediated cardiovascular risk. Yet, the extent to which standardized exercise programmes can reduce cardiovascular risk differs between patients. Exercise parameters, such as frequency, intensity, type and duration or number of repetitions, differentially target metabolic function, vascular health and physical fitness. In addition, exercise-induced molecular mechanisms are modulated by other patient-specific variables, such as age, diet and medication.

This review discusses molecular and cellular mechanisms underlying the effects of exercise training on cardiovascular risk specifically in patients with MetS and T2DM.

key words: exercise training, diabetes, signalling, glycaemic control, inflammation

2. Introduction

Patients with metabolic syndrome (MetS) suffer from a number of pathologic conditions, including abdominal obesity, hypertension, dyslipidemia and hyperglycaemia. Those factors strongly predispose MetS patients to type 2 diabetes (T2DM) and cardiovascular disease. A chronically positive caloric balance leads to overloading of adipose tissue lipid storage capacity. As a result, free fatty acid (FFA) - induced cellular stress responses feed, among other, hepatic and systemic inflammatory processes and a reduction in skeletal muscle insulin sensitivity.¹ Products of fatty acid metabolism and advanced glycated end-products, accumulating due to hyperglycaemia, upregulate and activate various pattern recognition receptors, thereby aggravating cytokine synthesis in innate immune cells and inflammatory aspects of vascular disease.²⁻⁷

Activation of innate immune cells feeds back into their metabolism: all cellular processes, including mitochondrial adenosine triphosphate (ATP) generation, take second place to the perceived need to defend against a pathogen. Hence, a “broken” Krebs cycle was observed in activated, pro-inflammatory “M1” type macrophages.^{8, 9} Krebs cycle intermediates then feed synthesis of pro-inflammatory molecules instead of ATP.¹⁰

Metabo-inflammatory processes are strongly influenced by the patient’s genetic and epigenetic background as well as their lifestyle choices (**Figure 1**). Importantly, even in patients with unfavourable genetic background a healthy lifestyle can significantly reduce cardiovascular risk to levels below that of patients with a beneficial genetic background but unhealthy lifestyles.¹¹ Exercise training (ET) is an integral component of a lifestyle benefitting cardiovascular health with the ability to improve distinct target parameters (glucose control, lipid status, physical fitness).^{1, 12, 13} Interestingly, exercise parameters – frequency, intensity, type and duration - appear to target different physiological responses. Endurance training usually leads to improved oxygen uptake and vascular function, while resistance training with increasing workloads causes skeletal muscle hypertrophy. Hence, combined endurance/resistance training programmes have been reported superior for targeting glycaemic control, anti-inflammatory effects or body composition.^{12, 14-19} In the accompanying paper by Kemps et al. (pp XXX of this issue), target parameters of ET in T2DM patients and suitable exercise parameters are discussed from a clinical perspective, while here we discuss the underlying molecular principles.

3. Contraction-induced molecular mechanisms

During the contraction cycle, intracellular cytoplasmic calcium levels fluctuate and ATP is dephosphorylated by myosin (**Figure 2**). The relative kinetics of those two signals can be modulated

by type, intensity and duration of exercise as well as by the cellular content of fuels such as glycogen, and impacts on exercise effects. Other factors, such as sympathetic activation take a modulating role (**Figure 2**).

Within the contracting myocyte, the increase in adenosine monophosphate (AMP) in relation to ATP is sensed by the AMP-activated kinase (AMPK).²⁰ AMPK activation upon rise of the AMP-to-ATP ratio generally results in an increase of processes providing ATP, such as mitochondrial beta-oxidation and glucose uptake into the cell, and in downregulation of competing ATP-consuming processes not immediately needed for survival, including fatty acid biosynthesis.²¹⁻²⁵ Exercise intensity affects the choice of substrates with fatty acid oxidation increasing until about 60% of the individual's maximal oxygen uptake capacity (VO₂max), then reaching a plateau. Glucose utilization instead increases further with increasing exercise intensity. Thus, at high exercise intensity, glucose outweighs FFAs as substrate for mitochondrial oxidation in healthy individuals.^{26, 27}

One important downstream mediator of AMPK signalling is the transcriptional coactivator peroxisome proliferator-activated receptor gamma coactivator 1-alpha (PGC-1α). PGC-1α target genes mostly serve mitochondrial biogenesis and FFA oxidation, but extend to anti-inflammatory processes, muscle growth and angiogenesis.²⁸⁻³⁰ Differential effects of exercise type and intensity on skeletal muscle growth versus energy metabolism are mediated by differential PGC-1α splicing.³¹ The two prominent PGC-1α isoforms - PGC-1α1 and PGC-1α4 - activate a large number of genes in common, but each isoform also governs their own specific gene set.³¹ PGC-1α1 regulates genes involved in mitochondrial oxidation and fuel supply, while PGC-1α4 also regulates genes mediating skeletal muscle hypertrophy, including *IGF1* and myostatin.³¹ Consistently, resistance exercise or a combination of resistance and endurance exercise increase expression of PGC-1α4 more than endurance training alone.³¹

Contraction-related calcium fluxes modulate AMPK signalling at several levels. Calcium-dependent kinases phosphorylate AMPK, thereby modulating its activity.³² Calcium also increases the activity of the protein kinase B (PKB), which relieves the inhibition of the mammalian target of Rapamycin (mTOR) by AMPK.³³ Resistance training also activates mTOR leading to the induction of protein synthesis serving skeletal muscle hypertrophy.³⁴ Intriguingly, glucose tolerance remains normal in PKB overexpressing mice on a high-fat diet, but not in the overfed wild type controls.³⁵ Therefore, the interplay between AMPK signalling (which favours glucose and FFA utilization for ATP synthesis) and PKB/mTOR signalling (which prefers protein synthesis) represents another mechanism explaining why combining improved glucose handling in patients with T2DM through combined resistance training and endurance training.³³⁻⁴¹

Systemic inflammation is a hallmark of MetS, T2DM and coronary disease. ET programs have repeatedly been shown to reduce systemic inflammation, as well as the activation state of innate immune cells.^{42, 43} One mechanism, by which ET can relieve the inflammatory load in T2DM could be an increased FFA uptake into the skeletal muscle. This reduces plasma FFA levels and their activation of toll-like receptors (**Figure 1**).⁴⁻⁷ In line with this, recent research has pointed to the mitochondria linking metabolic and anti-inflammatory effects of ET.^{10, 44} AMPK signalling is impaired in inflammatory activation of macrophages, caused by saturated fatty acids.^{6, 7} Improved FFA removal by the active skeletal muscle might therefore also act to relieve AMPK activity and improve mitochondrial oxidative function.^{13, 45} In line with this, skeletal muscle mitochondrial oxidative capacity - more so than mitochondrial density - appears to mediate the beneficial effects of exercise on T2DM-associated metabolic and cardiovascular structural and functional impairments.^{46, 47}

Skeletal muscle cells release a number of mediators during or upon exercise, which link local contractile activity to systemic inflammatory state. Examples include not only cytokines and growth factors, but also metabolites, long and short non-coding RNAs and cell-derived vesicles.⁴⁸ Brain-derived neurotrophic factor (BDNF) and interleukin-6 (IL-6), support AMPK signalling and result in improved FFA oxidation.⁴⁵ The resulting reduction of pattern-recognition receptor ligands in long-term ET programmes may contribute to the shift in the cytokine profile.^{2, 4, 13} Long and short non-coding RNAs (e.g. microRNA) are released into the circulation as a means of inter-organ communication and differ with exercise parameters and the patient's training status and underlying disease.⁴⁹⁻⁵⁵ Associations between microRNA (miRs), transcriptional regulators (HDAC4, NRF1) and levels of circulating effectors (IGF-1, IL-10, testosterone) support the notion of distinct regulatory networks activated by acute resistance versus endurance exercise.^{49, 53} Of note, alterations in circulating miR levels do not simply mirror local tissue miR levels, but resemble an integration between release and uptake by several different tissues.^{51, 55, 56} Hence, better integrated OMICs studies - accounting for the patient's genetic makeup, pathology, medication, training status and exercise parameters, combined with assessment of nucleic acids, (lipo-)proteins and multi-molecular complexes, such as extracellular vesicles, in various tissues will be more informative on systemic regulatory mechanisms than assessment of only individual players.^{45, 48}

4. Mechanical forces mediating exercise-induced signalling mechanisms

Exercise acutely applies mechanical strains and stresses to the muscle itself, but also to the heart and vasculature.⁵⁷ Within the muscle, not only myocytes, but also fibroblasts, macrophages and vascular cells respond to mechanical forces.^{58, 59} In the contracting muscle, tension changes are sensed by focal adhesions and ion channels and initiate signalling cascades resulting in altered deposition and remodelling of extracellular matrix (**Figure 1**).^{58, 60, 61} Force intensity appears to

modulate individual signalling mechanisms, as has been shown for kinases downstream of MAPK and for the matrix-metalloprotease-2, which can impact on both, matrix stiffness and angiogenesis.^{59, 61-64} Low-intensity and endurance training, especially in trained individuals, stimulates MAPK-ERK to induce cell proliferation and survival, while high-power resistance training activates MAPK-JNK which is associated with pro-apoptotic functions.⁶⁵⁻⁶⁷ Differential effects in skeletal muscle fibre types might exist and other cues such as hormones are likely to interact.⁵⁹

The heart releases – among other factors - natriuretic peptides in response to strain. Natriuretic peptides can induce lipolysis in human adipocytes and increase plasma non-esterified fatty acids, thus potentially linking mechanical forces in ET to metabolism.^{68, 69} In support of this claim, natriuretic peptide plasma levels are lower in persons with obesity, MetS and T2DM.⁷⁰⁻⁷³ An improvement of exercise capacity was associated with increased levels of B-type natriuretic peptides in heart failure patients.⁷⁴ Whether natriuretic peptides are causally involved in ET metabolic effects in T2DM patients, or whether the above-mentioned studies rather point to non-causal associations in responsive individuals remains unanswered. A recent murine study could not observe exercise-mediated increases in natriuretic peptides.⁷⁵ Neprilysin inhibitors might provide a pharmacological support strategy here and can help to unravel mechanistic links.⁷⁶

In the local and systemic vasculature, shear stress increase during exercise mediates endothelial survival and function, including regulation of vascular tone and angiogenesis (**Figure 2**).^{77, 78} Flow patterns and velocity distinctly affect cellular signalling mechanisms via inter-cellular junctions and luminal membrane proteins, including syndecans and glypicans which connect to the glycocalyx.⁷⁹⁻⁸⁴ The probably best-studied response of endothelial cells to physiological shear stress is the synthesis of nitric oxide (NO). The transcription factor KLF2 links the mechanosensors to expression of the endothelial NO synthase, while kinases, such as PKB, and Ca²⁺ entry through ion channels enhance its activity.^{79, 82, 83, 85-87} In addition, lower superoxide generation via NADPH oxidases and mitochondria together with enhanced superoxide dismutation protect NO levels.⁸⁸⁻⁹⁰ Hence, vascular NO bioavailability is increased under atheroprotective flow. NO not only induces vasorelaxation and anti-thrombotic endothelial mechanisms, but also activates protective enzymes such as SIRT1.⁹¹⁻⁹³ Apart from eNOS, KLF2 also influences the release of inflammatory cytokines and adhesion molecules, which are involved in vascular inflammation.^{94, 95} Hence, shear stress crucially mediates anti-atherosclerotic effects of regular ET on the vasculature in T2DM and MetS.⁹⁶⁻⁹⁸

5. Exercise-induced adrenergic signalling mechanisms

Sufficiently intense exercise increases adrenergic signalling, which governs not only adaptations in cardiac and skeletal muscle contraction force and perfusion but also metabolic adjustments.⁹⁹⁻¹⁰⁴ These include an initial reduction in insulin release (required for glucose release from the liver),

increases in adipocyte lipolysis (resulting in the release of FFAs) and glycogenolysis within skeletal muscle (providing an endogenous fuel). Epinephrine and norepinephrine levels increase in T2DM patients during exercise to a similar or even greater extent than in healthy controls, albeit slower and more prolonged.¹⁰⁵ This might explain slower FFA release into the plasma in response to moderate-intensity endurance exercise in T2DM patients observed in one study.¹⁰⁶ Others reported a normal increase of norepinephrine levels during acute exercise in T2DM patients with normal heart rate response, but only a small increase in epinephrine.¹⁰⁷

A bout of acute endurance exercise initially decreases plasma glucose in T2DM patients but not in non-diabetic controls. Further, the subsequent increase in plasma glucose occurred earlier and was greater compared to non-diabetic controls.¹⁰⁸⁻¹¹¹ Whether this was due to likely through reduced insulin sensitivity remains to be discussed, as a large part of exercise-mediated glucose uptake into skeletal muscle cells is insulin-independent.^{25, 112}

While catecholamines serve to adapt the cardiovascular system to an acute challenge, chronic employment of catecholamines to compensate a reduction of cardiac output cause systemic dysregulations in signalling mechanisms, as seen in heart failure. Regular ET can normalize catecholamine levels at rest, also in T2DM.^{113, 114}

The exact mechanisms by which acute and regular exercise modulate the release and uptake of glucose and FFA in T2DM patients still remain to be elucidated. Heterogeneous observations are likely through individual combinations of patient characteristics (state of insulin insensitivity and chronotropic competence) and exercise parameters (type, intensity).^{104, 105, 115} Considering the wide range of functions regulated by the various adrenergic receptors beyond exerting chronotropic and inotropic effects – including lipolysis, thermoregulation and vasorelaxation – further studies also need to look at the role of genetic variations in genes encoding components of the adrenergic system when assessing the effect of exercise programmes.¹¹⁶⁻¹¹⁸

6. Interaction of exercise with other determinants

A number of modifiable and non-modifiable variables interact with the individuals' response to exercise, including diet, pharmacotherapy, age, sex and the genetic background (**Figure 2**). There is a definitive lack of knowledge regarding most of these interactions. Hence, further research into those matters is warranted to provide better guidance in combining medication and diet with personalized exercise programmes to optimize the benefit for the individual patient.

6.1. Modifiable determinants

Interventional lifestyle change programmes in patients with MetS and T2DM recommend to simultaneously target diet and exercise to achieve a larger effect on body weight reduction and glucose handling.¹¹⁹ It is currently under debate, however, how exactly an optimal diet for patients with T2DM should be composed.¹²⁰ Recent evidence has led to a change in recommendations, now avoiding simple fat reduction, but focusing more on the quality of each nutrient component - i.e. complex versus refined carbohydrates and sugars, saturated versus mono- and polyunsaturated fats and trans fats.¹²¹ While large studies, such as the PURE study, can help provide associative evidence on diet and cardiovascular mortality¹²¹, there is still too little information on the interaction between diets of different micro- and macronutrient composition and exercise. Diet and exercise can interact on several mechanistic levels: Diet and frequency of exercise influence intracellular glycogen content, which in turn modulates the activation of AMPK.¹²²⁻¹²⁶ Further, low-to-moderate and high intensity interval training have distinct effects on insulin sensitivity. Dietary supplements, such as L-carnitine, affect lipid oxidation during exercise in healthy athletes, but it also conveys cardiovascular risk by yet unknown mechanisms.^{127, 128} Further attention in this respect needs therefore also been given to the gut microbiome, which is affected by both, diet and exercise and via its metabolites modulates cardiovascular risk.^{129, 130}

Interaction of pharmacologic treatment of MeTS and T2DM, or their cardiovascular co-morbidities with exercise effects have only recently gained attention. Hence, knowledge is still severely limited and mostly regards specific interactions.¹³¹ Exercise-induced hyperglycaemia is normally counteracted by hepatic glucose release. Sulfonylureas, and potentially also other insulin secretagogues, might therefore increase hyperglycaemic risk during exercise, albeit less so for metformin.¹³²⁻¹³⁴ Sodium-glucose co-transporter type 2 inhibitors may provide an alternative treatment to accompany exercise for T2DM patients.¹³⁵ Beta-blockers reduce sensing of hypoglycaemia in diabetic patients, but might - counter-intuitively - increase liver glucose production.^{136, 137} In patients requiring insulin, long- and short-acting formulations can be used to avoid exercise hyper- and hypo-glycaemia.¹³⁸⁻¹⁴⁰ Nevertheless, blood glucose levels need to be well monitored when combining these pharmacologic treatments with exercise in T2DM patients. An impairment of mitochondrial function by metformin has been observed, but it is currently unknown whether exercise can still improve mitochondrial function and mass despite metformin treatment in T2DM patients and which exercise conditions might be best suited in combination with this important pharmacological tool, also acknowledging sex-specific metformin excretion.¹⁴¹⁻¹⁴⁴ Statins, while lowering plasma cholesterol levels, are also associated with an increased risk for T2DM.^{145, 146} Addition of exercise to statin treatment is more effective in improving insulin sensitivity, reducing inflammation, and increasing exercise capacity than statin treatment alone.¹⁴⁷ Individual adaptation

of statin type and dose, as well as vitamin D or Q10 replacement might potentially help to limit statin-induced muscle pain potentially help maintain exercise adherence.¹⁴⁸

In contrast to genetic features, epigenetic markers can be modified (e.g. by diet and exercise), thus modulating metabolic risk in the offspring.¹⁴⁹ Vice versa, epigenetic mechanisms might determine the response to exercise, and targeting these might provide a further means to optimize ET response.^{150, 151}

6.2. Non-modifiable determinants of exercise effects

Sex-specific effects in substrate utilization and muscle anabolism at different exercise intensities are known well from healthy athletes and have recently also been observed in patients with MetS and T2DM.^{99-103, 152-161} On the other hand, fasting glucose and triglyceride levels improved significantly in response to a 12-week endurance training programme, regardless of sex in middle-aged T2DM patients.¹⁶¹ Sex hormones, with estrogen the most intensely studied, affect vascular function, fibrosis and immunity, thus conferring cardiovascular protection.¹⁶²⁻¹⁶⁹ This effect deteriorates in post-menopausal women, but can be preserved to some degree by regular ET.^{166, 170}

Exercise-mediated improvements in glucose handling and insulin sensitivity is less efficient in older than in younger mice and humans.¹⁷¹ Moreover, age modulated the hypertrophic effects of resistance training.¹⁷² Adrenergic β 2-receptor sensitivity, which falls sharply with advancing age, has been postulated as a responsible factor. However, treatment of older diabetic mice with a β 2-agonist improved insulin sensitivity, indicating that age-associated loss of β 2 sensitivity can be overcome pharmacologically.¹⁷¹

Response to exercise in patients with MetS and T2DM is also affected by the patient's genetic makeup. Relevant mechanisms include the actual contraction process and metabolic adaptations to acute exercise^{173, 174}, the regenerative phase where exercise-induced muscle damage is repaired¹⁷⁵ and metabolic processes take place^{118, 176, 177}, as well as structural and metabolic adaptations to regularly repeated exercise.^{178, 179} A recent study in over 2,800 individuals identified genes determining the success of lifestyle-mediated as well as pharmacologic interventions to improve insulin sensitivity, including the genes for hepatocyte nuclear factor 4 α (*HNF4*) and neuronal differentiation 1 (*NEUROD1*).¹⁸⁰ Genetic predisposition might also modulate effects and side-effects of pharmacological treatments, as shown for statins, metformin and sulfonylureas.^{141, 143, 181-184}

Murine studies point to the bi-directional interaction between voluntary wheel running and genetic mechanisms in determining VO_2 max, thus linking genetic background with adherence.^{185, 186} Since findings in mice cannot be directly translated to humans with their high diversity of genetic background, pathologies and life styles, big-data based methodologies are warranted in future

studies to better delineate the interaction between individual modifiable and non-modifiable factors influencing the patients' response and adherence to exercise.

7. Conclusions

In conclusion, regular ET exerts pleiotropic effects, beneficial not only to metabolic, but also to cardiovascular endpoints in patients with MetS and T2DM. Exercise parameters, such as type, intensity and duration modulate effects on individual clinical target parameters. In addition, the patient's individual genetic background, as well as sex, age, diet and medication interact with exercise and confer risks as well as effectiveness to meet treatment goals.

Future studies need to delineate these multi-dimensional interactions and enable the delivery of personalized exercise programmes. Novel data exploitation methods in combination with telemedicine approaches can help to deal with the high degree of inter-individual variation in a human population as well as the difficulties to standardize lifestyle interventions. In addition, target parameters other than HbA_{1c} need to be taken into consideration when aiming to tackle cardiovascular risk, among them inflammation and blood pressure control.

8. Acknowledgements

Sources of Funding

NK, MB and MD are supported by the German Centre for Cardiovascular Research (DZHK, partner sites Berlin (NK) and Greifswald (MB, MD)). EvC is supported by the Fund for Scientific Research Flanders.

Conflicting Interests

The authors declare no conflicting interests.

9. Figure Legends

Figure 1: Effects of an exercise programme on target treatment parameters in T2DM and MetS are affected by modifiable and non-modifiable, patient-related parameters, in addition to exercise-related parameters.

Figure 2: Exercise-induced stimuli, such as calcium fluxes and ATP-to-AMP conversion initiate signalling events favouring substrate uptake and oxidation, and the synthesis of mitochondrial and contractile proteins. Systemic inflammatory status is improved by reduced circulating levels of fatty acid metabolites and glycated end-products and altered paracrine spectra of myocytes and immune cells. Mechanical forces initiate muscle and vascular remodelling and vascular nitric oxide availability. Jointly, those mechanisms lead to reduced plasma levels of LDL, glucose and inflammatory mediators and to improved vascular function. Depending on the individual's age, sex, the parameters of the exercise programme, genetic background and medication, each of the listed mechanisms may be modulated, affecting the individual effect of ET.

10. References

1. Bacchi E, Negri C, Targher G, et al. Both resistance training and aerobic training reduce hepatic fat content in type 2 diabetic subjects with nonalcoholic fatty liver disease (the RAED2 Randomized Trial). *Hepatology*. 2013; 58: 1287-95.
2. Jimenez-Dalmaroni MJ, Xiao N, Corper AL, et al. Soluble CD36 ectodomain binds negatively charged diacylglycerol ligands and acts as a co-receptor for TLR2. *PLoS one*. 2009; 4: e7411.
3. Hanssen NM, Beulens JW, van Dieren S, et al. Plasma advanced glycation end products are associated with incident cardiovascular events in individuals with type 2 diabetes: a case-cohort study with a median follow-up of 10 years (EPIC-NL). *Diabetes*. 2015; 64: 257-65.
4. Hussey SE, Lum H, Alvarez A, et al. A sustained increase in plasma NEFA upregulates the Toll-like receptor network in human muscle. *Diabetologia*. 2014; 57: 582-91.
5. Lee JY, Zhao L, Youn HS, et al. Saturated fatty acid activates but polyunsaturated fatty acid inhibits Toll-like receptor 2 dimerized with Toll-like receptor 6 or 1. *The Journal of biological chemistry*. 2004; 279: 16971-9.
6. Schaeffler A, Gross P, Buettner R, et al. Fatty acid-induced induction of Toll-like receptor-4/nuclear factor-kappaB pathway in adipocytes links nutritional signalling with innate immunity. *Immunology*. 2009; 126: 233-45.
7. Suganami T, Tanimoto-Koyama K, Nishida J, et al. Role of the Toll-like receptor 4/NF-kappaB pathway in saturated fatty acid-induced inflammatory changes in the interaction between adipocytes and macrophages. *Arterioscler Thromb Vasc Biol*. 2007; 27: 84-91.
8. Mills EL and O'Neill LA. Reprogramming mitochondrial metabolism in macrophages as an anti-inflammatory signal. *Eur J Immunol*. 2016; 46: 13-21.
9. Jha AK, Huang SC, Sergushichev A, et al. Network integration of parallel metabolic and transcriptional data reveals metabolic modules that regulate macrophage polarization. *Immunity*. 2015; 42: 419-30.
10. Mills EL, Kelly B and O'Neill LAJ. Mitochondria are the powerhouses of immunity. *Nat Immunol*. 2017; 18: 488-98.
11. Khera AV, Emdin CA, Drake I, et al. Genetic Risk, Adherence to a Healthy Lifestyle, and Coronary Disease. *The New England journal of medicine*. 2016; 375: 2349-58.
12. Thomas DE, Elliott EJ and Naughton GA. Exercise for type 2 diabetes mellitus. *The Cochrane database of systematic reviews*. 2006: CD002968.
13. Liang H, Tantiwong P, Sriwijitkamol A, et al. Effect of a sustained reduction in plasma free fatty acid concentration on insulin signalling and inflammation in skeletal muscle from human subjects. *The Journal of physiology*. 2013; 591: 2897-909.
14. Hopps E, Canino B and Caimi G. Effects of exercise on inflammation markers in type 2 diabetic subjects. *Acta diabetologica*. 2011; 48: 183-9.
15. Oliveira C, Simoes M, Carvalho J and Ribeiro J. Combined exercise for people with type 2 diabetes mellitus: a systematic review. *Diabetes research and clinical practice*. 2012; 98: 187-98.
16. Balducci S, Zanuso S, Cardelli P, et al. Changes in physical fitness predict improvements in modifiable cardiovascular risk factors independently of body weight loss in subjects with type 2 diabetes participating in the Italian Diabetes and Exercise Study (IDES). *Diabetes care*. 2012; 35: 1347-54.
17. Schwingshackl L, Missbach B, Dias S, Konig J and Hoffmann G. Impact of different training modalities on glycaemic control and blood lipids in patients with type 2 diabetes: a systematic review and network meta-analysis. *Diabetologia*. 2014; 57: 1789-97.
18. Liu Y, Liu SX, Cai Y, Xie KL, Zhang WL and Zheng F. Effects of combined aerobic and resistance training on the glycolipid metabolism and inflammation levels in type 2 diabetes mellitus. *Journal of physical therapy science*. 2015; 27: 2365-71.
19. Codella R, Ialacqua M, Terruzzi I and Luzi L. May the force be with you: why resistance training is essential for subjects with type 2 diabetes mellitus without complications. *Endocrine*. 2018.

20. Hardie DG, Salt IP and Davies SP. Analysis of the role of the AMP-activated protein kinase in the response to cellular stress. *Methods Mol Biol.* 2000; 99: 63-74.
21. Hardie DG. AMP-activated/SNF1 protein kinases: conserved guardians of cellular energy. *Nat Rev Mol Cell Biol.* 2007; 8: 774-85.
22. Winder WW and Hardie DG. Inactivation of acetyl-CoA carboxylase and activation of AMP-activated protein kinase in muscle during exercise. *Am J Physiol.* 1996; 270: E299-304.
23. Park H, Kaushik VK, Constant S, et al. Coordinate regulation of malonyl-CoA decarboxylase, sn-glycerol-3-phosphate acyltransferase, and acetyl-CoA carboxylase by AMP-activated protein kinase in rat tissues in response to exercise. *The Journal of biological chemistry.* 2002; 277: 32571-7.
24. Coderre L, Kandror KV, Vallega G and Pilch PF. Identification and characterization of an exercise-sensitive pool of glucose transporters in skeletal muscle. *The Journal of biological chemistry.* 1995; 270: 27584-8.
25. Sherman LA, Hirshman MF, Cormont M, Le Marchand-Brustel Y and Goodyear LJ. Differential effects of insulin and exercise on Rab4 distribution in rat skeletal muscle. *Endocrinology.* 1996; 137: 266-73.
26. Rose AJ and Richter EA. Skeletal muscle glucose uptake during exercise: how is it regulated? *Physiology (Bethesda).* 2005; 20: 260-70.
27. van Loon LJ, Greenhaff PL, Constantin-Teodosiu D, Saris WH and Wagenmakers AJ. The effects of increasing exercise intensity on muscle fuel utilisation in humans. *The Journal of physiology.* 2001; 536: 295-304.
28. Chinsomboon J, Ruas J, Gupta RK, et al. The transcriptional coactivator PGC-1 α mediates exercise-induced angiogenesis in skeletal muscle. *Proc Natl Acad Sci U S A.* 2009; 106: 21401-6.
29. Eisele PS, Furrer R, Beer M and Handschin C. The PGC-1 coactivators promote an anti-inflammatory environment in skeletal muscle in vivo. *Biochem Biophys Res Commun.* 2015; 464: 692-7.
30. Safdar A, Little JP, Stokl AJ, Hettinga BP, Akhtar M and Tarnopolsky MA. Exercise increases mitochondrial PGC-1 α content and promotes nuclear-mitochondrial cross-talk to coordinate mitochondrial biogenesis. *The Journal of biological chemistry.* 2011; 286: 10605-17.
31. Ruas JL, White JP, Rao RR, et al. A PGC-1 α isoform induced by resistance training regulates skeletal muscle hypertrophy. *Cell.* 2012; 151: 1319-31.
32. Hawley SA, Pan DA, Mustard KJ, et al. Calmodulin-dependent protein kinase kinase-beta is an alternative upstream kinase for AMP-activated protein kinase. *Cell Metab.* 2005; 2: 9-19.
33. Atherton PJ, Babraj J, Smith K, Singh J, Rennie MJ and Wackerhage H. Selective activation of AMPK-PGC-1 α or PKB-TSC2-mTOR signaling can explain specific adaptive responses to endurance or resistance training-like electrical muscle stimulation. *FASEB journal : official publication of the Federation of American Societies for Experimental Biology.* 2005; 19: 786-8.
34. Parkington JD, Siebert AP, LeBrasseur NK and Fielding RA. Differential activation of mTOR signaling by contractile activity in skeletal muscle. *Am J Physiol Regul Integr Comp Physiol.* 2003; 285: R1086-90.
35. Izumiya Y, Hopkins T, Morris C, et al. Fast/Glycolytic muscle fiber growth reduces fat mass and improves metabolic parameters in obese mice. *Cell Metab.* 2008; 7: 159-72.
36. Trebak JT, Birk JB, Rose AJ, Kiens B, Richter EA and Wojtaszewski JF. AS160 phosphorylation is associated with activation of α 2 β 2 γ 1- but not α 2 β 2 γ 3-AMPK trimeric complex in skeletal muscle during exercise in humans. *American journal of physiology Endocrinology and metabolism.* 2007; 292: E715-22.
37. Egan B, Carson BP, Garcia-Roves PM, et al. Exercise intensity-dependent regulation of peroxisome proliferator-activated receptor coactivator-1 mRNA abundance is associated with differential activation of upstream signalling kinases in human skeletal muscle. *The Journal of physiology.* 2010; 588: 1779-90.
38. Chen ZP, Stephens TJ, Murthy S, et al. Effect of exercise intensity on skeletal muscle AMPK signaling in humans. *Diabetes.* 2003; 52: 2205-12.

39. Sriwijitkamol A, Coletta DK, Wajcberg E, et al. Effect of acute exercise on AMPK signaling in skeletal muscle of subjects with type 2 diabetes: a time-course and dose-response study. *Diabetes*. 2007; 56: 836-48.
40. Dreyer HC, Fujita S, Cadenas JG, Chinkes DL, Volpi E and Rasmussen BB. Resistance exercise increases AMPK activity and reduces 4E-BP1 phosphorylation and protein synthesis in human skeletal muscle. *The Journal of physiology*. 2006; 576: 613-24.
41. Vissing K, McGee S, Farup J, Kjolhede T, Vendelbo M and Jessen N. Differentiated mTOR but not AMPK signaling after strength vs endurance exercise in training-accustomed individuals. *Scandinavian journal of medicine & science in sports*. 2013; 23: 355-66.
42. Peters PG, Alessio HM, Hagerman AE, Ashton T, Nagy S and Wiley RL. Short-term isometric exercise reduces systolic blood pressure in hypertensive adults: possible role of reactive oxygen species. *International journal of cardiology*. 2006; 110: 199-205.
43. Diment BC, Fortes MB, Edwards JP, et al. Exercise Intensity and Duration Effects on In Vivo Immunity. *Medicine and science in sports and exercise*. 2015; 47: 1390-8.
44. Weinberg SE, Sena LA and Chandel NS. Mitochondria in the regulation of innate and adaptive immunity. *Immunity*. 2015; 42: 406-17.
45. Pedersen BK and Febbraio MA. Muscles, exercise and obesity: skeletal muscle as a secretory organ. *Nat Rev Endocrinol*. 2012; 8: 457-65.
46. Lumini JA, Magalhaes J, Oliveira PJ and Ascensao A. Beneficial effects of exercise on muscle mitochondrial function in diabetes mellitus. *Sports medicine*. 2008; 38: 735-50.
47. Sparks LM, Johannsen NM, Church TS, et al. Nine months of combined training improves ex vivo skeletal muscle metabolism in individuals with type 2 diabetes. *J Clin Endocrinol Metab*. 2013; 98: 1694-702.
48. Safdar A, Saleem A and Tarnopolsky MA. The potential of endurance exercise-derived exosomes to treat metabolic diseases. *Nat Rev Endocrinol*. 2016; 12: 504-17.
49. Cui S, Sun B, Yin X, et al. Time-course responses of circulating microRNAs to three resistance training protocols in healthy young men. *Scientific reports*. 2017; 7: 2203.
50. Wardle SL, Bailey ME, Kilikevicius A, et al. Plasma microRNA levels differ between endurance and strength athletes. *PloS one*. 2015; 10: e0122107.
51. Cui SF, Wang C, Yin X, et al. Similar Responses of Circulating MicroRNAs to Acute High-Intensity Interval Exercise and Vigorous-Intensity Continuous Exercise. *Front Physiol*. 2016; 7: 102.
52. Banzet S, Chennaoui M, Girard O, et al. Changes in circulating microRNAs levels with exercise modality. *Journal of applied physiology*. 2013; 115: 1237-44.
53. Russell AP, Lamon S, Boon H, et al. Regulation of miRNAs in human skeletal muscle following acute endurance exercise and short-term endurance training. *The Journal of physiology*. 2013; 591: 4637-53.
54. Morais Junior GS, Souza VC, Machado-Silva W, et al. Acute strength training promotes responses in whole blood circulating levels of miR-146a among older adults with type 2 diabetes mellitus. *Clin Interv Aging*. 2017; 12: 1443-50.
55. Pietrangelo T, Di Filippo ES, Mancinelli R, et al. Low Intensity Exercise Training Improves Skeletal Muscle Regeneration Potential. *Front Physiol*. 2015; 6: 399.
56. D'Souza RF, Markworth JF, Aasen KMM, Zeng N, Cameron-Smith D and Mitchell CJ. Acute resistance exercise modulates microRNA expression profiles: Combined tissue and circulatory targeted analyses. *PloS one*. 2017; 12: e0181594.
57. Kjaer M. Role of extracellular matrix in adaptation of tendon and skeletal muscle to mechanical loading. *Physiol Rev*. 2004; 84: 649-98.
58. Timmons JA, Jansson E, Fischer H, et al. Modulation of extracellular matrix genes reflects the magnitude of physiological adaptation to aerobic exercise training in humans. *BMC Biol*. 2005; 3: 19.
59. Wretman C, Lionikas A, Widegren U, Lannergren J, Westerblad H and Henriksson J. Effects of concentric and eccentric contractions on phosphorylation of MAPK(erk1/2) and MAPK(p38) in isolated rat skeletal muscle. *The Journal of physiology*. 2001; 535: 155-64.

60. Ingber DE. Mechanical signaling and the cellular response to extracellular matrix in angiogenesis and cardiovascular physiology. *Circ Res.* 2002; 91: 877-87.
61. Carmeli E, Moas M, Lennon S and Powers SK. High intensity exercise increases expression of matrix metalloproteinases in fast skeletal muscle fibres. *Experimental physiology.* 2005; 90: 613-9.
62. Parker L, Trewin A, Levinger I, Shaw CS and Stepto NK. The effect of exercise-intensity on skeletal muscle stress kinase and insulin protein signaling. *PLoS one.* 2017; 12: e0171613.
63. Williamson D, Gallagher P, Harber M, Hollon C and Trappe S. Mitogen-activated protein kinase (MAPK) pathway activation: effects of age and acute exercise on human skeletal muscle. *The Journal of physiology.* 2003; 547: 977-87.
64. Mackey AL, Donnelly AE, Turpeenniemi-Hujanen T and Roper HP. Skeletal muscle collagen content in humans after high-force eccentric contractions. *Journal of applied physiology.* 2004; 97: 197-203.
65. Galpin AJ, Fry AC, Chiu LZ, Thomason DB and Schilling BK. High-power resistance exercise induces MAPK phosphorylation in weightlifting trained men. *Appl Physiol Nutr Metab.* 2012; 37: 80-7.
66. Gonzalez AM, Hoffman JR, Townsend JR, et al. Intramuscular MAPK signaling following high volume and high intensity resistance exercise protocols in trained men. *European journal of applied physiology.* 2016; 116: 1663-70.
67. Kramer HF and Goodyear LJ. Exercise, MAPK, and NF-kappaB signaling in skeletal muscle. *Journal of applied physiology.* 2007; 103: 388-95.
68. Sengenès C, Berlan M, De Glisezinski I, Lafontan M and Galitzky J. Natriuretic peptides: a new lipolytic pathway in human adipocytes. *FASEB journal : official publication of the Federation of American Societies for Experimental Biology.* 2000; 14: 1345-51.
69. Galitzky J, Sengenès C, Thalamas C, et al. The lipid-mobilizing effect of atrial natriuretic peptide is unrelated to sympathetic nervous system activation or obesity in young men. *Journal of lipid research.* 2001; 42: 536-44.
70. Das SR, Drazner MH, Dries DL, et al. Impact of body mass and body composition on circulating levels of natriuretic peptides: results from the Dallas Heart Study. *Circulation.* 2005; 112: 2163-8.
71. Wang TJ, Larson MG, Keyes MJ, Levy D, Benjamin EJ and Vasán RS. Association of plasma natriuretic peptide levels with metabolic risk factors in ambulatory individuals. *Circulation.* 2007; 115: 1345-53.
72. Salomaa V, Havulinna A, Saarela O, et al. Thirty-one novel biomarkers as predictors for clinically incident diabetes. *PLoS one.* 2010; 5: e10100.
73. Magnusson M, Jujic A, Hedblad B, et al. Low plasma level of atrial natriuretic peptide predicts development of diabetes: the prospective Malmo Diet and Cancer study. *J Clin Endocrinol Metab.* 2012; 97: 638-45.
74. Nakanishi M, Nakao K, Kumasaka L, et al. Improvement in Exercise Capacity by Exercise Training Associated With Favorable Clinical Outcomes in Advanced Heart Failure With High B-Type Natriuretic Peptide Level. *Circulation journal : official journal of the Japanese Circulation Society.* 2017; 81: 1307-14.
75. Broderick TL, Jankowski M and Gutkowska J. The effects of exercise training and caloric restriction on the cardiac oxytocin natriuretic peptide system in the diabetic mouse. *Diabetes, metabolic syndrome and obesity : targets and therapy.* 2017; 10: 27-36.
76. Malek V and Gaikwad AB. Nephilysin inhibitors: A new hope to halt the diabetic cardiovascular and renal complications? *Biomed Pharmacother.* 2017; 90: 752-9.
77. Green DJ, Hopman MT, Padilla J, Laughlin MH and Thijssen DH. Vascular Adaptation to Exercise in Humans: Role of Hemodynamic Stimuli. *Physiol Rev.* 2017; 97: 495-528.
78. Baeyens N, Bandyopadhyay C, Coon BG, Yun S and Schwartz MA. Endothelial fluid shear stress sensing in vascular health and disease. *J Clin Invest.* 2016; 126: 821-8.
79. Wang N, Miao H, Li YS, et al. Shear stress regulation of Kruppel-like factor 2 expression is flow pattern-specific. *Biochem Biophys Res Commun.* 2006; 341: 1244-51.

80. Chien S. Mechanotransduction and endothelial cell homeostasis: the wisdom of the cell. *Am J Physiol Heart Circ Physiol*. 2007; 292: H1209-24.
81. Jalali S, del Pozo MA, Chen K, et al. Integrin-mediated mechanotransduction requires its dynamic interaction with specific extracellular matrix (ECM) ligands. *Proc Natl Acad Sci U S A*. 2001; 98: 1042-6.
82. Voyvodic PL, Min D, Liu R, et al. Loss of syndecan-1 induces a pro-inflammatory phenotype in endothelial cells with a dysregulated response to atheroprotective flow. *The Journal of biological chemistry*. 2014; 289: 9547-59.
83. Tzima E, Irani-Tehrani M, Kiosses WB, et al. A mechanosensory complex that mediates the endothelial cell response to fluid shear stress. *Nature*. 2005; 437: 426-31.
84. Kuchan MJ, Jo H and Frangos JA. Role of G proteins in shear stress-mediated nitric oxide production by endothelial cells. *Am J Physiol*. 1994; 267: C753-8.
85. Zhang DX, Mendoza SA, Bubolz AH, et al. Transient receptor potential vanilloid type 4-deficient mice exhibit impaired endothelium-dependent relaxation induced by acetylcholine in vitro and in vivo. *Hypertension*. 2009; 53: 532-8.
86. Davis ME, Cai H, Drummond GR and Harrison DG. Shear stress regulates endothelial nitric oxide synthase expression through c-Src by divergent signaling pathways. *Circ Res*. 2001; 89: 1073-80.
87. Yamamoto K, Sokabe T, Matsumoto T, et al. Impaired flow-dependent control of vascular tone and remodeling in P2X4-deficient mice. *Nature medicine*. 2006; 12: 133-7.
88. Inoue N, Ramasamy S, Fukai T, Nerem RM and Harrison DG. Shear stress modulates expression of Cu/Zn superoxide dismutase in human aortic endothelial cells. *Circ Res*. 1996; 79: 32-7.
89. Godbole AS, Lu X, Guo X and Kassab GS. NADPH oxidase has a directional response to shear stress. *American journal of physiology Heart and circulatory physiology*. 2009; 296: H152-8.
90. Takabe W, Jen N, Ai L, et al. Oscillatory shear stress induces mitochondrial superoxide production: implication of NADPH oxidase and c-Jun NH2-terminal kinase signaling. *Antioxid Redox Signal*. 2011; 15: 1379-88.
91. Meares GP, Hughes KJ, Naatz A, et al. IRE1-dependent activation of AMPK in response to nitric oxide. *Mol Cell Biol*. 2011; 31: 4286-97.
92. Chen Z, Peng IC, Cui X, Li YS, Chien S and Shyy JY. Shear stress, SIRT1, and vascular homeostasis. *Proc Natl Acad Sci U S A*. 2010; 107: 10268-73.
93. Zhang W, Huang Q, Zeng Z, Wu J, Zhang Y and Chen Z. Sirt1 Inhibits Oxidative Stress in Vascular Endothelial Cells. *Oxid Med Cell Longev*. 2017; 2017: 7543973.
94. Parmar KM, Larman HB, Dai G, et al. Integration of flow-dependent endothelial phenotypes by Kruppel-like factor 2. *J Clin Invest*. 2006; 116: 49-58.
95. SenBanerjee S, Lin Z, Atkins GB, et al. KLF2 Is a novel transcriptional regulator of endothelial proinflammatory activation. *J Exp Med*. 2004; 199: 1305-15.
96. Greyling A, Schreuder TH, Landman T, et al. Elevation in blood flow and shear rate prevents hyperglycemia-induced endothelial dysfunction in healthy subjects and those with type 2 diabetes. *Journal of applied physiology*. 2015; 118: 579-85.
97. Limberg JK, Johansson RE, McBride PE and Schrage WG. Increased leg blood flow and improved femoral artery shear patterns in metabolic syndrome after a diet and exercise programme. *Clin Physiol Funct Imaging*. 2014; 34: 282-9.
98. Madsen SM, Thorup AC, Overgaard K, Bjerre M and Jeppesen PB. Functional and structural vascular adaptations following 8 weeks of low volume high intensity interval training in lower leg of type 2 diabetes patients and individuals at high risk of metabolic syndrome. *Arch Physiol Biochem*. 2015; 121: 178-86.
99. Peake JM, Tan SJ, Markworth JF, Broadbent JA, Skinner TL and Cameron-Smith D. Metabolic and hormonal responses to isoenergetic high-intensity interval exercise and continuous moderate-intensity exercise. *American journal of physiology Endocrinology and metabolism*. 2014; 307: E539-52.

100. Kraemer WJ, Gordon SE, Fragala MS, et al. The effects of exercise training programs on plasma concentrations of proenkephalin Peptide F and catecholamines. *Peptides*. 2015; 64: 74-81.
101. Kolnes AJ, Birk JB, Eilertsen E, Stuenkel JT, Wojtaszewski JF and Jensen J. Epinephrine-stimulated glycogen breakdown activates glycogen synthase and increases insulin-stimulated glucose uptake in epitrochlearis muscles. *American journal of physiology Endocrinology and metabolism*. 2015; 308: E231-40.
102. Sutherland LN, Bomhof MR, Capozzi LC, Basaraba SA and Wright DC. Exercise and adrenaline increase PGC-1 α mRNA expression in rat adipose tissue. *The Journal of physiology*. 2009; 587: 1607-17.
103. Schmidt SL, Bessesen DH, Stotz S, Peelor FF, 3rd, Miller BF and Horton TJ. Adrenergic control of lipolysis in women compared with men. *Journal of applied physiology*. 2014; 117: 1008-19.
104. Marliss EB and Vranic M. Intense exercise has unique effects on both insulin release and its roles in gluco-regulation: implications for diabetes. *Diabetes*. 2002; 51 Suppl 1: S271-83.
105. Turgan N, Coker C, Hamulu F, et al. Glucose metabolism and catecholamine responses during physical exercise in non-insulin-dependent diabetes. *Eur J Clin Chem Clin Biochem*. 1996; 34: 683-9.
106. Blaak EE, van Aggel-Leijssen DP, Wagenmakers AJ, Saris WH and van Baak MA. Impaired oxidation of plasma-derived fatty acids in type 2 diabetic subjects during moderate-intensity exercise. *Diabetes*. 2000; 49: 2102-7.
107. Wilson GA, Wilson LC, Lamberts RR, et al. beta-Adrenergic Responsiveness in the Type 2 Diabetic Heart: Effects on Cardiac Reserve. *Medicine and science in sports and exercise*. 2017; 49: 907-14.
108. Kjaer M, Hollenbeck CB, Frey-Hewitt B, Galbo H, Haskell W and Reaven GM. Gluco-regulation and hormonal responses to maximal exercise in non-insulin-dependent diabetes. *Journal of applied physiology*. 1990; 68: 2067-74.
109. Price TB, Perseghin G, Duleba A, et al. NMR studies of muscle glycogen synthesis in insulin-resistant offspring of parents with non-insulin-dependent diabetes mellitus immediately after glycogen-depleting exercise. *Proc Natl Acad Sci U S A*. 1996; 93: 5329-34.
110. Schneider SH, Khachaturian AK, Amorosa LF, Gavras H, Fineberg SE and Ruderman NB. Abnormal gluco-regulation during exercise in type II (non-insulin-dependent) diabetes. *Metabolism: clinical and experimental*. 1987; 36: 1161-6.
111. Giacca A, Groenewoud Y, Tsui E, McClean P and Zinman B. Glucose production, utilization, and cycling in response to moderate exercise in obese subjects with type 2 diabetes and mild hyperglycemia. *Diabetes*. 1998; 47: 1763-70.
112. Douen AG, Ramlal T, Klip A, Young DA, Cartee GD and Holloszy JO. Exercise-induced increase in glucose transporters in plasma membranes of rat skeletal muscle. *Endocrinology*. 1989; 124: 449-54.
113. Madsen SM, Thorup AC, Bjerre M and Jeppesen PB. Does 8 weeks of strenuous bicycle exercise improve diabetes-related inflammatory cytokines and free fatty acids in type 2 diabetes patients and individuals at high-risk of metabolic syndrome? *Arch Physiol Biochem*. 2015; 121: 129-38.
114. Hamasaki H, Kawashima Y, Tamada Y, et al. Associations of Low-Intensity Resistance Training with Body Composition and Lipid Profile in Obese Patients with Type 2 Diabetes. *PloS one*. 2015; 10: e0132959.
115. Schumann U, Jenkinson CP, Alt A, Zugel M, Steinacker JM and Flechtner-Mors M. Sympathetic nervous system activity and anti-lipolytic response to iv-glucose load in subcutaneous adipose tissue of obese and obese type 2 diabetic subjects. *PloS one*. 2017; 12: e0173803.
116. Lowell BB and Bachman ES. Beta-Adrenergic receptors, diet-induced thermogenesis, and obesity. *The Journal of biological chemistry*. 2003; 278: 29385-8.
117. Dessy C, Moniotte S, Ghisdal P, Havaux X, Noirhomme P and Balligand JL. Endothelial beta3-adrenoceptors mediate vasorelaxation of human coronary microarteries through nitric oxide and endothelium-dependent hyperpolarization. *Circulation*. 2004; 110: 948-54.

118. Sakane N, Sato J, Tsushita K, et al. Effects of lifestyle intervention on weight and metabolic parameters in patients with impaired glucose tolerance related to beta-3 adrenergic receptor gene polymorphism Trp64Arg(C/T): Results from the Japan Diabetes Prevention Program. *Journal of diabetes investigation*. 2016; 7: 338-42.
119. Zhang X, Devlin HM, Smith B, et al. Effect of lifestyle interventions on cardiovascular risk factors among adults without impaired glucose tolerance or diabetes: A systematic review and meta-analysis. *PLoS one*. 2017; 12: e0176436.
120. Dyson PA, Twenefour D, Breen C, et al. Diabetes UK evidence-based nutrition guidelines for the prevention and management of diabetes. *Diabetic medicine : a journal of the British Diabetic Association*. 2018; 35: 541-7.
121. Dehghan M, Mente A, Zhang X, et al. Associations of fats and carbohydrate intake with cardiovascular disease and mortality in 18 countries from five continents (PURE): a prospective cohort study. *Lancet*. 2017; 390: 2050-62.
122. McBride A, Ghilagaber S, Nikolaev A and Hardie DG. The glycogen-binding domain on the AMPK beta subunit allows the kinase to act as a glycogen sensor. *Cell Metab*. 2009; 9: 23-34.
123. Cochran AJ, Little JP, Tarnopolsky MA and Gibala MJ. Carbohydrate feeding during recovery alters the skeletal muscle metabolic response to repeated sessions of high-intensity interval exercise in humans. *Journal of applied physiology*. 2010; 108: 628-36.
124. Creer A, Gallagher P, Slivka D, Jemiolo B, Fink W and Trappe S. Influence of muscle glycogen availability on ERK1/2 and Akt signaling after resistance exercise in human skeletal muscle. *Journal of applied physiology*. 2005; 99: 950-6.
125. Febbraio MA, Steensberg A, Walsh R, et al. Reduced glycogen availability is associated with an elevation in HSP72 in contracting human skeletal muscle. *The Journal of physiology*. 2002; 538: 911-7.
126. Psilander N, Frank P, Flockhart M and Sahlin K. Exercise with low glycogen increases PGC-1alpha gene expression in human skeletal muscle. *European journal of applied physiology*. 2013; 113: 951-63.
127. Broad EM, Maughan RJ and Galloway SD. Effects of exercise intensity and altered substrate availability on cardiovascular and metabolic responses to exercise after oral carnitine supplementation in athletes. *International journal of sport nutrition and exercise metabolism*. 2011; 21: 385-97.
128. Koeth RA, Wang Z, Levison BS, et al. Intestinal microbiota metabolism of L-carnitine, a nutrient in red meat, promotes atherosclerosis. *Nature medicine*. 2013; 19: 576-85.
129. Clarke SF, Murphy EF, O'Sullivan O, et al. Exercise and associated dietary extremes impact on gut microbial diversity. *Gut*. 2014; 63: 1913-20.
130. Le Chatelier E, Nielsen T, Qin J, et al. Richness of human gut microbiome correlates with metabolic markers. *Nature*. 2013; 500: 541-6.
131. Hemmingsen B, Sonne DP, Metzendorf MI and Richter B. Insulin secretagogues for prevention or delay of type 2 diabetes mellitus and its associated complications in persons at increased risk for the development of type 2 diabetes mellitus. *The Cochrane database of systematic reviews*. 2016; 10: CD012151.
132. Marwick TH, Hordern MD, Miller T, et al. Exercise training for type 2 diabetes mellitus: impact on cardiovascular risk: a scientific statement from the American Heart Association. *Circulation*. 2009; 119: 3244-62.
133. Larsen JJ, Dela F, Madsbad S, Vibe-Petersen J and Galbo H. Interaction of sulfonylureas and exercise on glucose homeostasis in type 2 diabetic patients. *Diabetes care*. 1999; 22: 1647-54.
134. Shahar J and Hamdy O. Medication and exercise interactions: considering and managing hypoglycemia risk. *Diabetes Spectr*. 2015; 28: 64-7.
135. Lam KS, Chow CC, Tan KC, et al. Practical considerations for the use of sodium-glucose co-transporter type 2 inhibitors in treating hyperglycemia in type 2 diabetes. *Curr Med Res Opin*. 2016; 32: 1097-108.

136. Sigal RJ, Purdon C, Bilinski D, Vranic M, Halter JB and Marliss EB. Glucoregulation during and after intense exercise: effects of beta-blockade. *J Clin Endocrinol Metab.* 1994; 78: 359-66.
137. Best JD and Halter JB. Blood pressure and norepinephrine spillover during propranolol infusion in humans. *Am J Physiol.* 1985; 248: R400-6.
138. Tanaka N and Hiura Y. Effects of rapid-acting insulin analogues insulin glulisine and insulin aspart on postprandial glycemic excursion with single bout of exercise in patients with type 2 diabetes. *Endocr J.* 2015; 62: 411-6.
139. Plockinger U, Topuz M, Riese B and Reuter T. Risk of exercise-induced hypoglycaemia in patients with type 2 diabetes on intensive insulin therapy: comparison of insulin glargine with NPH insulin as basal insulin supplement. *Diabetes research and clinical practice.* 2008; 81: 290-5.
140. Praet SF, Manders RJ, Lieveise AG, et al. Influence of acute exercise on hyperglycemia in insulin-treated type 2 diabetes. *Medicine and science in sports and exercise.* 2006; 38: 2037-44.
141. Boule NG, Robert C, Bell GJ, et al. Metformin and exercise in type 2 diabetes: examining treatment modality interactions. *Diabetes care.* 2011; 34: 1469-74.
142. Wessels B, Ciapaite J, van den Broek NM, Nicolay K and Prompers JJ. Metformin impairs mitochondrial function in skeletal muscle of both lean and diabetic rats in a dose-dependent manner. *PLoS one.* 2014; 9: e100525.
143. Hou W, Zhang D, Lu W, et al. Polymorphism of organic cation transporter 2 improves glucose-lowering effect of metformin via influencing its pharmacokinetics in Chinese type 2 diabetic patients. *Mol Diagn Ther.* 2015; 19: 25-33.
144. Esteghamati A, Mousavizadeh M, Noshad S, Zandieh A, Zarei H and Nakhjavani M. Gender-dependent effects of metformin on vaspin and adiponectin in type 2 diabetes patients: a randomized clinical trial. *Horm Metab Res.* 2013; 45: 319-25.
145. Abd TT and Jacobson TA. Statin-induced myopathy: a review and update. *Expert Opin Drug Saf.* 2011; 10: 373-87.
146. Sattar N, Preiss D, Murray HM, et al. Statins and risk of incident diabetes: a collaborative meta-analysis of randomised statin trials. *Lancet.* 2010; 375: 735-42.
147. Gui YJ, Liao CX, Liu Q, et al. Efficacy and safety of statins and exercise combination therapy compared to statin monotherapy in patients with dyslipidaemia: A systematic review and meta-analysis. *European journal of preventive cardiology.* 2017; 24: 907-16.
148. Deichmann RE, Lavie CJ, Asher T, DiNicolantonio JJ, O'Keefe JH and Thompson PD. The Interaction Between Statins and Exercise: Mechanisms and Strategies to Counter the Musculoskeletal Side Effects of This Combination Therapy. *Ochsner J.* 2015; 15: 429-37.
149. Stanford KI, Takahashi H, So K, et al. Maternal Exercise Improves Glucose Tolerance in Female Offspring. *Diabetes.* 2017; 66: 2124-36.
150. Parr EB, Camera DM, Burke LM, Phillips SM, Coffey VG and Hawley JA. Circulating MicroRNA Responses between 'High' and 'Low' Responders to a 16-Wk Diet and Exercise Weight Loss Intervention. *PLoS one.* 2016; 11: e0152545.
151. Rowlands DS, Page RA, Sukala WR, et al. Multi-omic integrated networks connect DNA methylation and miRNA with skeletal muscle plasticity to chronic exercise in Type 2 diabetic obesity. *Physiol Genomics.* 2014; 46: 747-65.
152. Steffensen CH, Roepstorff C, Madsen M and Kiens B. Myocellular triacylglycerol breakdown in females but not in males during exercise. *American journal of physiology Endocrinology and metabolism.* 2002; 282: E634-42.
153. Roepstorff C, Steffensen CH, Madsen M, et al. Gender differences in substrate utilization during submaximal exercise in endurance-trained subjects. *American journal of physiology Endocrinology and metabolism.* 2002; 282: E435-47.
154. Tate CA and Holtz RW. Gender and fat metabolism during exercise: a review. *Can J Appl Physiol.* 1998; 23: 570-82.
155. Numao S, Hayashi Y, Katayama Y, Matsuo T and Tanaka K. Sex differences in substrate oxidation during aerobic exercise in obese men and postmenopausal obese women. *Metabolism: clinical and experimental.* 2009; 58: 1312-9.

156. Horton TJ, Dow S, Armstrong M and Donahoo WT. Greater systemic lipolysis in women compared with men during moderate-dose infusion of epinephrine and/or norepinephrine. *Journal of applied physiology*. 2009; 107: 200-10.
157. Roepstorff C, Donsmark M, Thiele M, et al. Sex differences in hormone-sensitive lipase expression, activity, and phosphorylation in skeletal muscle at rest and during exercise. *American journal of physiology Endocrinology and metabolism*. 2006; 291: E1106-14.
158. Justice TD, Hammer GL, Davey RJ, et al. Effect of antecedent moderate-intensity exercise on the glycemia-increasing effect of a 30-sec maximal sprint: a sex comparison. *Physiol Rep*. 2015; 3.
159. Lebeck J, Ostergard T, Rojek A, et al. Gender-specific effect of physical training on AQP7 protein expression in human adipose tissue. *Acta diabetologica*. 2012; 49 Suppl 1: S215-26.
160. Miyamoto T, Fukuda K, Watanabe K, Hidaka M and Moritani T. Gender difference in metabolic responses to surface electrical muscle stimulation in type 2 diabetes. *J Electromyogr Kinesiol*. 2015; 25: 136-42.
161. Adeniyi AF, Uloko AE, Ogwumike OO, Sanya AO and Fasanmade AA. Time course of improvement of metabolic parameters after a 12 week physical exercise programme in patients with type 2 diabetes: the influence of gender in a Nigerian population. *BioMed research international*. 2013; 2013: 310574.
162. Hoeg LD, Sjoberg KA, Jeppesen J, et al. Lipid-induced insulin resistance affects women less than men and is not accompanied by inflammation or impaired proximal insulin signaling. *Diabetes*. 2011; 60: 64-73.
163. Kautzky-Willer A, Harreiter J and Pacini G. Sex and Gender Differences in Risk, Pathophysiology and Complications of Type 2 Diabetes Mellitus. *Endocr Rev*. 2016; 37: 278-316.
164. Straub RH. The complex role of estrogens in inflammation. *Endocr Rev*. 2007; 28: 521-74.
165. Liu H, Liu K and Bodenner DL. Estrogen receptor inhibits interleukin-6 gene expression by disruption of nuclear factor kappaB transactivation. *Cytokine*. 2005; 31: 251-7.
166. Casey DP, Shepherd JR and Joyner MJ. Sex and vasodilator responses to hypoxia at rest and during exercise. *Journal of applied physiology*. 2014; 116: 927-36.
167. Jones AW, Rubin LJ and Magliola L. Endothelin-1 sensitivity of porcine coronary arteries is reduced by exercise training and is gender dependent. *Journal of applied physiology*. 1999; 87: 1172-7.
168. Clerico A, Fontana M, Vittorini S and Emdin M. The search for a pathophysiological link between gender, cardiac endocrine function, body mass regulation and cardiac mortality: proposal for a working hypothesis. *Clin Chim Acta*. 2009; 405: 1-7.
169. Lam CS, Cheng S, Choong K, et al. Influence of sex and hormone status on circulating natriuretic peptides. *Journal of the American College of Cardiology*. 2011; 58: 618-26.
170. Nyberg M, Seidelin K, Andersen TR, Overby NN, Hellsten Y and Bangsbo J. Biomarkers of vascular function in premenopausal and recent postmenopausal women of similar age: effect of exercise training. *Am J Physiol Regul Integr Comp Physiol*. 2014; 306: R510-7.
171. Ziegler MG, Elayan H, Milic M, Sun P and Gharaibeh M. Epinephrine and the metabolic syndrome. *Current hypertension reports*. 2012; 14: 1-7.
172. Phillips BE, Williams JP, Gustafsson T, et al. Molecular networks of human muscle adaptation to exercise and age. *PLoS Genet*. 2013; 9: e1003389.
173. Meirhaeghe A, Sandhu MS, McCarthy MI, et al. Association between the T-381C polymorphism of the brain natriuretic peptide gene and risk of type 2 diabetes in human populations. *Hum Mol Genet*. 2007; 16: 1343-50.
174. Jayewardene AF, Mavros Y, Gwinn T, Hancock DP and Rooney KB. Associations between CD36 gene polymorphisms and metabolic response to a short-term endurance-training program in a young-adult population. *Appl Physiol Nutr Metab*. 2016; 41: 157-67.
175. Baumert P, Lake MJ, Stewart CE, Drust B and Erskine RM. Genetic variation and exercise-induced muscle damage: implications for athletic performance, injury and ageing. *European journal of applied physiology*. 2016; 116: 1595-625.

176. Grunnet LG, Brons C, Jacobsen S, et al. Increased recovery rates of phosphocreatine and inorganic phosphate after isometric contraction in oxidative muscle fibers and elevated hepatic insulin resistance in homozygous carriers of the A-allele of FTO rs9939609. *J Clin Endocrinol Metab.* 2009; 94: 596-602.
177. Tobina T, Mori Y, Doi Y, Nakayama F, Kiyonaga A and Tanaka H. Peroxisome proliferator-activated receptor gamma co-activator 1 gene Gly482Ser polymorphism is associated with the response of low-density lipoprotein cholesterol concentrations to exercise training in elderly Japanese. *J Physiol Sci.* 2017; 67: 595-602.
178. Stephens NA and Sparks LM. Resistance to the beneficial effects of exercise in type 2 diabetes: are some individuals programmed to fail? *J Clin Endocrinol Metab.* 2015; 100: 43-52.
179. Peter I, Papandonatos GD, Belalcazar LM, et al. Genetic modifiers of cardiorespiratory fitness response to lifestyle intervention. *Medicine and science in sports and exercise.* 2014; 46: 302-11.
180. Billings LK, Jablonski KA, Warner AS, et al. Variation in Maturity-Onset Diabetes of the Young Genes Influence Response to Interventions for Diabetes Prevention. *J Clin Endocrinol Metab.* 2017; 102: 2678-89.
181. Group SC, Link E, Parish S, et al. SLCO1B1 variants and statin-induced myopathy--a genomewide study. *The New England journal of medicine.* 2008; 359: 789-99.
182. Dujic T, Zhou K, Yee SW, et al. Variants in Pharmacokinetic Transporters and Glycemic Response to Metformin: A Metgen Meta-Analysis. *Clin Pharmacol Ther.* 2017; 101: 763-72.
183. Song J, Yang Y, Mauvais-Jarvis F, Wang YP and Niu T. KCNJ11, ABCC8 and TCF7L2 polymorphisms and the response to sulfonylurea treatment in patients with type 2 diabetes: a bioinformatics assessment. *BMC Med Genet.* 2017; 18: 64.
184. Erickson ML, Little JP, Gay JL, McCully KK and Jenkins NT. Effects of postmeal exercise on postprandial glucose excursions in people with type 2 diabetes treated with add-on hypoglycemic agents. *Diabetes research and clinical practice.* 2017; 126: 240-7.
185. Rezende EL, Garland T, Jr., Chappell MA, Malisch JL and Gomes FR. Maximum aerobic performance in lines of Mus selected for high wheel-running activity: effects of selection, oxygen availability and the mini-muscle phenotype. *J Exp Biol.* 2006; 209: 115-27.
186. Rezende EL, Gomes FR, Malisch JL, Chappell MA and Garland T, Jr. Maximal oxygen consumption in relation to subordinate traits in lines of house mice selectively bred for high voluntary wheel running. *Journal of applied physiology.* 2006; 101: 477-85.