

This item is the archived peer-reviewed author-version of:

Coadministration of a Gloriosa superba extract improves the in vivo antitumoural activity of gemcitabine in a murine pancreatic tumour model

Reference:

Capistrano I. Rica, Vangestel Christel, Vanpachtenbeke Hanne, Fransen Erik, Staelens Steven, Apers Sandra, Pieters Luc.-Coadministration of a Gloriosa superba extract improves the in vivo antitumoural activity of gemcitabine in a murine pancreatic tumour model

Phytomedicine: international journal of phytotherapy and phytopharmacology - ISSN 0944-7113 - 23:12(2016), p. 1434-1440

Full text (Publishers DOI): <http://dx.doi.org/doi:10.1016/j.phymed.2016.07.012>

To cite this reference: <http://hdl.handle.net/10067/1348890151162165141>

4 Rica Capistrano^{I,a}, Christel Vangestel^{b,c}, Hanne Vanpachtenbeke^a, Erik Fransen^d, Steven Staelens^c,
5 Sandra Apers^a, Luc Pieters^a

6

⁷ ^a Natural Products & Food Research and Analysis (NatuRA), Department of Pharmaceutical Sciences,
⁸ University of Antwerp, Antwerp (Wilrijk), Belgium

⁹ ^bDepartment of Nuclear Medicine, Antwerp University Hospital, Edegem, Belgium

10 ^c Molecular Imaging Center Antwerp (MICA), University of Antwerp, Antwerp (Wilrijk), Belgium

¹¹ ^d StatUA Center for Statistics, University of Antwerp, Prinsstraat 13, 2000 Antwerp, Belgium

12

13 **Abstract**

14 *Background:* *Gloriosa superba* L. (glory lily, Colchicaceae) contains colchicine, and related alkaloids
15 such as 3-O-demethylcolchicine and its glycoside colchicoside. Previously the *in vivo* efficacy of a
16 crude extract and a colchicine-poor / colchicoside-rich extract of *G. superba* seeds was shown in a
17 murine model of pancreatic adenocarcinoma.

18 *Hypothesis/Purpose:* The efficacy can be improved without obvious signs of toxicity by increasing the
19 treatment dose; the efficacy of gemcitabine can be improved by coadministration of a *Gloriosa*
20 *superba* extract.

21 *Study Design:* A survival experiment was carried out in a murine model of pancreatic adenocarcinoma
22 and the semi-long-term toxicity of both *G. superba* extracts was determined; a combination therapy
23 with gemcitabine was evaluated.

24 *Methods:* A crude ethanolic extract (GS) and a colchicine-poor / colchicoside-rich extract (GS2B)
25 were prepared, containing 3.22% colchicine, 2.52% colchicoside and 1.52% 3-O-demethylcolchicine
26 (GS), and 0.07%, 2.26% and 0.46% (m/m) (GS2B). They were evaluated in a murine model of
27 pancreatic adenocarcinoma at a dose of 4.5 mg/kg (p.o., daily) total content of colchicine and
28 derivatives during 3 weeks, or at 3.0 mg/kg (p.o., daily) combined with gemcitabine (60 mg/kg, i.p.,
29 3x/week) during 54 days.

30 *Results:* A significant effect in tumour growth over time was observed for gemcitabine and the
31 combination therapy compared to the control group. No significant difference was observed for the
32 groups treated with colchicine and both extracts. However, combination therapy was significantly
33 better than the monotherapy with gemcitabine. Moreover, survival analysis showed a significant
34 prolongation of the survival of the groups treated with gemcitabine and the combination therapy. A
35 slight difference in survival was observed between gemcitabine and the combination therapy, the latter
36 one being slightly better. No significant prolongation of survival was observed for the extracts and
37 colchicine compared to the control group.

38 *Conclusion:* Although a relevant tumour growth inhibition and a difference of relative tumour volume
39 compared to the control group were observed on day 11, and a slightly longer survival was noticed for
40 GS2B, the most important conclusion from this study is that the crude *G. superba* extract (GS) might
41 have an added value combined with gemcitabine in the treatment of pancreatic tumours.

42

43 **Keywords**

44 *Gloriosa superba*; Colchicaceae; pancreatic tumours; gemcitabine; combination therapy

45

46

47 Abbreviations

48	BW	Body weight
49	GS	Crude ethanolic extract of <i>Gloriosa superba</i>
50	GS2B	Colchicine-poor / colchicoside-rich extract of <i>Gloriosa superba</i>
51	RTV	Relative tumour volume
52	TC	Total content of colchicine and derivatives (expressed as colchicine)
53	TGD	Tumour growth delay
54	TGI	Tumour growth inhibition

55

56 **Introduction**

57 *Gloriosa superba* L. (glory lily), belonging to the Colchicaceae, has traditionally been used for various
58 diseases and is widely cultivated as an ornamental plant. It contains colchicine, which is an inhibitor of
59 tubulin polymerisation used against gout, and related alkaloids such as 3-O-demethylcolchicine and its
60 glycoside colchicoside (Jana et al., 2011; Maroyi et al., 2011; Risinger et al., 2009). The use of pure
61 colchicine as an anticancer agent has been excluded for toxicity reasons (Stanton et al., 2011; Yue et
62 al., 2010), although more recently this has been questioned again (Slobodnick et al., 2015; Solak et al.,
63 2015). However, it is known that plant extracts may display a better activity profile than the individual
64 constituents (Wagner et al., 2009). *Gloriosa superba* was selected for the present study because it is
65 rich in colchicoside, the glucosidic derivative of 3-O-demethyl-colchicine, being the active aglycone.
66 In a previous study the *in vivo* efficacy of a crude extract and a colchicine-poor / colchicoside-rich
67 extract of *G. superba* seeds was shown in a murine model of pancreatic adenocarcinoma, supporting
68 the hypothesis that colchicoside indeed can be considered as a prodrug that is activated after oral
69 administration (Capistrano et al., 2016). Among all cancers, pancreatic cancer has an extremely poor
70 prognosis and is one of the most deadly types. Most pancreatic cancer patients die within the first year
71 of diagnosis and only 6% will survive five years. The lack of progress in primary prevention, early
72 diagnosis and treatment, underscores the need for new approaches in pancreatic cancer research
73 (American Cancer Society, 2013; Cascinu et al., 2009; Dureux et al., 2015; Li et al., 2004). Mice
74 having a fully competent immune response were used, since the immune system can play a potentially
75 beneficial role during therapy. This implies the use of mouse-strain specific tumour models (syngeneic
76 tumours), i.e. the application of murine PANC02 cancer cells rather than human pancreatic cancer
77 (PANC-1) cells (Campbell et al., 2014). In the present study a survival experiment was carried out and
78 the semi-long-term toxicity of both the crude extract and a colchicine-poor / colchicoside-rich extract
79 of *G. superba* seeds was determined. A combination therapy with gemcitabine was also evaluated to
80 establish whether this combination had an added value to monotherapy with gemcitabine, a nucleoside
81 analogue that is the first-line treatment in patients with locally advanced pancreatic cancer (Dureux et
82 al., 2015). However, even gemcitabine alone or in combination with other chemotherapeutics such as
83 cisplatin or 5-fluorouracil or radiotherapy only has a low objective response and a low survival benefit
84 (Arslan et al., 2014).

85

86 **Materials and methods**

87 *Plant material and preparation of the extracts*

88 Dried seeds of *Gloriosa superba* L. were kindly provided by Indena® (Milano, Italy) (batch n°
89 C140020, certificate of analysis n° 11/0208/LSP). A crude ethanolic extract (GS) and a colchicine-

90 poor / colchicoside-rich extract (GS2B) was prepared as reported before, and in both extracts the
91 amount of colchicine, colchicoside and 3-*O*-demethylcolchicine was determined using a validated
92 HPLC method (Capistrano et al., 2015). GS contained 3.22% colchicine, 1.30% colchicoside
93 expressed as colchicine and 1.27% 3-*O*-demethylcolchicine (all m/m) expressed as colchicine,
94 corresponding to a total content of colchicine equivalents (TC) of 5.79%. Similarly, GS2B contained
95 0.07%, 1.16% and 0.38% (m/m), respectively, corresponding to a total content of colchicine
96 equivalents (TC) (expressed as colchicine) of 1.61%. The amount of colchicine in GS2B was less than
97 0.1% (m/m).

98

99 *Cell line*

100 Murine pancreatic adenocarcinoma cells (PANC02) were kindly provided by Prof. Dr. C. Gravekamp
101 (Albert Einstein College of Medicine, New York, USA) and cultured as previously described
102 (Capistrano et al., 2016).

103

104 *Animals*

105 Six-to-eight weeks old female C57BL/6 mice (16 - 20 g body weight (BW)) were purchased from
106 Harlan laboratories. The mice were housed in individually ventilated cages, under a 10/14 h dark/light
107 cycle at constant temperature and humidity and had access to tap water and food *ad libitum*. All
108 animals were treated in accordance to the guidelines and regulations for use and care of animals. All
109 mice experiments were approved by the local Ethical Committee of the University of Antwerp,
110 approved study number: 2014-34.

111

112 *Subcutaneous tumour model*

113 Cultures of PANC02 cells were harvested using a 0.05% trypsin-EDTA solution, washed twice in
114 sterile PBS and resuspended in sterile PBS at a concentration of 30 x 10⁶ viable cells per ml. The
115 viable cells were counted by using the Muse® Cell Analyzer. Mice (n = 66) were inoculated with 100
116 µL of the cell suspension in the right hind limb. Tumour growth of the subcutaneous model was
117 evaluated by means of calliper measurements from the moment the tumours became palpable and this
118 three times a week. The tumour volume was calculated as in the previous study (Capistrano et al.,
119 2016). Each mouse was randomly assigned to one of 6 groups (n = 11 mice per group) and assigned to
120 a different treatment (Table 1).

121

122 *Preliminary acute toxicity study*

123 Before the actual survival experiment an acute toxicity study was carried out to evaluate the acute
124 toxic effects of a dose of 4.5 mg/kg BW total colchicine and derivatives (TC). Non-tumour-bearing
125 C57BL/6 mice (n = 3 per treatment group) were given 4.5 mg/kg BW colchicine, or the equivalent

126 dose of GS or the colchicine-poor / colchicoside-rich extract, during 5 consecutive days. The body
127 weight was determined daily, and the mice were also daily inspected for clinical signs of toxicity as
128 reported before during the experiment and 5 additional days of follow-up (Montgomery, 1990;
129 Capistrano et al., 2016).

130

131 *Treatments*

132 Treatment was started 12 days after tumour inoculation when tumours had reached a volume of
133 approximately 100 mm³. The first day of treatment was assigned “day 1” (Fig. 1). Mice of the negative
134 control group received 200 µl water p.o. (gavage) daily (group 1) (Table 1). The positive control group
135 was given 3 times/week gemcitabine (Actavis, 38 mg/ml) (group 2). It was administered at a dose of
136 60 mg/kg BW i.p. Group 3 was treated with colchicine at a dose of 4.5 mg/kg BW (daily, p.o.). GS
137 and GS2B were administered p.o. daily at a dose of 4.5 mg/kg BW TC during 3 weeks, i.e. group 4
138 was given 77.6 mg/kg BW GS and group 5 was given 281.3 mg/kg BW GS2B. Group 6 was treated
139 with a combination of extract GS at a dose of 3.0 mg/kg BW TC (p.o., daily) and gemcitabine (60
140 mg/kg, i.p., 3x/week) during 54 days. Tumour growth was evaluated three times per week and the
141 mice were sacrificed when the tumour had reached a volume of more than 1500 mm³ or a tumour
142 weight of more than 10% of the total body weight, and/or a weight loss of more than 20% was
143 observed.

144 *Statistics*

145 Results were expressed as mean values of parameters ± standard error (SE). The parametric ANOVA
146 test was used to determine statistical significance, followed by a post hoc Tukey analysis to establish
147 the statistical difference between the treatment groups. When only two groups were compared, as on
148 day 51, a Mann-Whitney test was used to determine statistical significance. A piecewise linear
149 regression model, in this case a so called linear mixed model was also fitted. This is a type of
150 regression that accounts for the dependence between observations within the same mouse. “Normal”
151 regression assumes that all observations are independent, which is not the case in this study since
152 multiple measurements were made per mouse. A mixed model accounts for the repeated
153 measurements by including random effect terms into the regression equation. These terms model the
154 effect of each individual mouse on the outcome, but were not the real interest of this study. The
155 interesting terms, time and treatment, are referred to as the fixed effects. The construction of the fixed
156 effects part of a linear mixed model is performed in an analogous way as model building in ordinary
157 linear regression. So a mixed model is a statistical model containing random effects in addition to the
158 usual fixed effects. A Kaplan–Meier survival curve was constructed and the logrank test was used to
159 determine statistical difference between the survivals of the different treatment groups. A p-value ≤
160 0.05 was considered significant and statistical analyses were performed using GraphPad Prism 6
161 (Version 6.01) and SPSS (version 22.0).

162

163 **Results and Discussion**

164 As shown in Fig. 2 the actual treatment doses did not cause any weight loss greater than 10% of the
165 initial body weight. None of the observation criteria were met by any animal during this acute toxicity
166 study. In addition, 5 days after treatment the body weights were still stable and still none of the mice
167 showed any clinical symptoms of toxicity. It could be concluded that the given dose of the different
168 treatments did not cause any acute toxicity.

169

170 All mice developed subcutaneous tumours in the right hind limb (100% successful tumour growth).
171 Eleven days after inoculation, the tumours had reached a mean volume of 100 mm³, the mice were
172 randomised and treatment started. The starting mean tumour volume per group was evaluated to check
173 whether there was no significant difference between the groups (Fig. 3) and the tumour volumes were
174 normalised at this time point. The mean relative tumour volumes were then calculated for each group
175 for each day. The mean relative tumour volumes on day 11, 21 and 51 were graphed (Fig. 4) and the
176 difference between the groups was statistically evaluated determining the significance of the effect of
177 each treatment. The %TGI was calculated for day 11 and 21. The tumour growth delay (TGD), which
178 is the difference or delay in days for treated versus control groups to reach a specified volume (in this
179 case twice the starting volume) was also determined. The %TGI and TGD are summarised in Table
180 **2.Fout! Verwijzingsbron niet gevonden.** All animals were weighed daily during treatment to
181 evaluate toxicity and the mean body weight per group was graphed (Fig. 5). Table 3 summarises the
182 mean body weight of the different treatments and the relative mean body weight at day 21. This table
183 shows a clear gain of body weight for all groups after 21 days of treatment.

184

185 The individual scatter of each mouse was plotted (Fig. 6 left). This graph shows the relative tumour
186 volume of all individual mice. Each dot represents one measurement in one mouse at a given time
187 point, with separate panels for each mouse. The strip text shows the mouse identification and the
188 treatment. The different colours represent the different treatments. The individual scatter was also
189 graphed per treatment group and is shown in Fig. 6 (right). A plot of the mean relative volume (on the
190 log scale) versus time is shown in Fig. 7. Each dot represents the mean of the ln(relative tumour
191 volume) on one time point. The different lines represent the different treatments. These data were then
192 fitted into a piecewise linear regression model (Fig. 8) and in this case a linear mixed model.

193

194 A Kaplan-Meier survival curve was graphed and shown in Fig. 9. The survival fraction is the fraction
195 of mice that was still alive at a certain time point, so for each time point, the graph shows the fraction

196 of mice still in de study per group. A drop of the line of a group indicated an event, which was in this
197 case a removal of a mouse from the study due to one of the defined study endpoints.

198 The mean RTV on day 11 and 21 showed a significant difference between the control group and the
199 gemcitabine group. A difference between the control group and the combination therapy was also
200 observed on both days. On day 11 a significant difference was observed for GS2B, and although no
201 significant difference was observed for colchicine and GS, a slight decrease in RTV was present. A
202 significant lower RTV was observed for the combination therapy at day 51 compared to the
203 gemcitabine group. The tumour growth inhibition was also calculated for the different groups on day
204 11 and 21. A relevant %TGI (> 50%) was observed for gemcitabine and the combination therapy on
205 both days with %TGI of 86% and 82% for gemcitabine and 94% and 92% for the combination
206 therapy. A relevant %TGI (57%) was also observed for GS2B on day 11. A tumour growth delay of 12
207 and 21 days was observed for gemcitabine and the combination group, respectively, as well as a TGD
208 of 5 days for GS2B.

209 Although the treatment dose was 1.5 times higher compared to our previous study (Capistrano et al.,
210 2016), it was still below the median lethal dose (LD_{50}) of colchicine, which is 6 mg/kg p.o. for mice.
211 No important body weight loss was observed throughout the experiment, and no severe side effects
212 were observed. Therefore, it can be concluded that no extreme toxicity, even after daily treatment of
213 more than 23 days at relatively high concentrations, was caused by the extracts or colchicine,
214 indicating the relatively moderate toxicity of colchicine and the extracts.

215 A piecewise linear regression model, in this case the linear mixed model was also fitted. This is a type
216 of regression that accounts for the dependence between observations within the same mouse.
217 “Normal” regression assumes that all observations are independent, which was not the case in this
218 study since multiple measurements were made per mouse. So a mixed model accounts for the repeated
219 measurements by including random effect terms into the regression equation, such as missing value
220 due to the euthanisation of mice when the tumour volume had reach 1500 mm³. The individual scatter
221 plots and the plot of the mean relative tumour volume suggested that in the control, colchicine, GS and
222 GS2B groups, the tumour growth was linear on the log scale from the start on. In the gemcitabine and
223 combination group, there was a phase of slow growth until day 25, which is followed by a phase of
224 increased growth at a pace comparable to the other 4 treatments. Due to this observation, the evolution
225 of tumour growth with time was modelled using piecewise linear regression, accounting for the
226 change in the slope (“knot”) around day 25. This regression model included the change in slope at a
227 given point in time. In all treatments, a knot (slope change) is modelled at day 25. However, only in
228 the combination therapy and gemcitabine group the change in slope was substantial. The hypothesis
229 was then tested whether the change in slope was different between the groups. So, overall comparisons

230 between groups showed that the null hypothesis, which is that the tumour growth over time was the
231 same in all treatment groups, was rejected with $p < 2 \times 10^{-16}$. By rejecting the null hypothesis of an
232 equal slope before the knot, and an equal change in slope beyond day 25, it could be concluded that
233 there was a statistical difference in tumour growth over time between treatment groups.

234 For the pairwise comparisons, no significant differences were observed between the control group and
235 the groups treated with colchicine, GS and GS2B. There was however a significant difference between
236 the control group and the combination therapy and gemcitabine. Moreover, the combination therapy
237 was significantly better than the gemcitabine group.

238 The Kaplan-Meier survival curve shows for each time point the fraction of mice still alive per group.
239 The calculations performed in this study take censored observations into account. If the data of a
240 certain mouse was censored, it was either because the mouse was removed from the study for reasons
241 not related to the endpoints of the study, or because the study has ended and that mouse was still alive
242 and no information beyond the time of censoring was available. While it seems intuitive that the curve
243 ought to end at a survival fraction computed as the total number of subjects who died divided by the
244 total number of mice, this is only correct if there are no censored data. A strong decline in survival
245 fraction at a certain time point indicated that a great number of mice has reached the endpoint of 1500
246 mm³ and were euthanised. So, for the survival analysis, a significant longer survival time compared to
247 the control group was observed for the group treated with gemcitabine ($p \leq 0.001$) and the
248 combination therapy ($p \leq 0.0001$). All other treatment groups did not show a significant longer
249 survival compared to the control group. The survival of the combination therapy compared to the
250 gemcitabine group did not show a significant difference, however, after 58 days of treatment fewer
251 mice treated with the combination group had died compared to the gemcitabine group due to the
252 defined study endpoints. This result was observed on the Kaplan-Meier curve where the survival
253 fraction at the end of the study was lower for gemcitabine compared to the combination therapy.

254 There have been a few studies on the combination of gemcitabine with an herbal extract before, more
255 in particular a *Rauwolfia vomitoria* extract (Yu et al., 2014) and Pao Pereira (*Geissospermum vellosii*)
256 (Yu et al., 2013). Gamma-tocotrienol potentiated the antitumour activity of gemcitabine in an
257 orthotopic pancreatic tumour model in nude mice (Kunnumakkara et al., 2010). However, this is the
258 first report on the coadministration of a *Gloriosa superba* extract with gemcitabine.

259

260 **Conclusion**

261 In summary, statistical analysis of the tumour growth over time between the treatment groups was
262 assessed by using a mixed linear model and showed a significant difference for gemcitabine and the
263 combination therapy compared to the control group. No significant difference in growth over time was
264 observed for the groups treated with colchicine and both extracts. The combination therapy was
265 significantly better than the monotherapy with gemcitabine. Moreover, survival analysis showed a
266 significant prolongation of the survival of the groups treated with gemcitabine and the combination
267 therapy. A slight difference in survival was observed between the gemcitabine and the combination
268 therapy, the latter one being slightly better. No significant prolongation of survival was observed for
269 the extracts and colchicine compared to the control group. Although a relevant tumour growth
270 inhibition for the colchicine-poor extract and a difference of RTV compared to the control group were
271 observed on day 11 and a slightly longer survival was noticed for the colchicine-poor extract, the most
272 important conclusion from this study is that the *G. superba* extract might have an added value
273 combined with gemcitabine in the treatment of pancreatic tumours. **For future research it seems**
274 **worthwhile also to evaluate the coadministration of the colchicoside-enriched extract with**
275 **gemcitabine.**

276

277

278 **Acknowledgements**

279

280 Thanks are due to Prof. Gravekamp (Albert Einstein College of Medicine, New York, USA), for
281 kindly providing the PANC02 cells and Indena (Milano, Italy) for providing the plant materials. We
282 gratefully acknowledge Yanina Dockx and Sven De Bruycker of the Molecular Imaging Center
283 Antwerp (MICA) for their excellent practical assistance. The *Agency for Innovation by Science and*
284 *Technology in Flanders* (IWT) is acknowledged for granting a PhD fellowship to R.C. Operational
285 expenses were in part defrayed by a research project of L.P. with the Anticancer Fund
286 (www.anticancerfund.org, a non-profit organization), which is acknowledged for scientific and
287 financial support. We would like to mention that the work of C.V. was enabled by IMI (Innovative
288 Medicines Initiative: Quic Concept (21NGN03)).

289

290 **Conflicts of interest**

291 The authors wish to confirm that there are no known conflicts of interest associated with this
292 publication and there has been no significant financial support for this work that could have influenced
293 its outcome.

294 **References**

- 295 American Cancer Society, 2013. Cancer Facts & Figures 2013. American Cancer Society.
- 296 Arslan, C., Yalcin, S., 2014. Current and future systemic treatment options in metastatic pancreatic
297 cancer. *J. Gastrointest. Oncol.* 5, 280-295.
- 298 Campbell, E.J., Dachs,G.U., 2014. Current limitations of murine models in oncology for ascorbate
299 research. *Front. Oncol.* 4, 282.
- 300 Capistrano I., R., 2015. Phytochemical, Analytical and Preclinical Investigations on Plant Extracts as
301 Potential Antitumoural Therapy. PhD Thesis, University of Antwerp, Belgium. chapter 4, 128-
302 154.
- 303 Capistrano I., R., Vangestel, C., Wouters, A., Dockx, Y., Pauwels, P., Stroobants, S., Apers, S.,
304 Lardon, F., Pieters, L., Staelens, S., 2016. Efficacy screening of *Gloriosa superba* extracts in a
305 murine pancreatic cancer model using ¹⁸F-FDG PET/CT for monitoring treatment response.
306 *Cancer Biother. Radiopharm.* **31**, 99-109
- 307 Cascinu, S., Jelic, S., Group, E.G.W., 2009. Pancreatic cancer: ESMO clinical recommendations for
308 diagnosis, treatment and follow-up. *Ann. Oncol.* 20, Suppl. 4, 37-40.
- 309 Ducreux, M., Cuhna, A. Sa., Caramella, C., Hollebecque, A., Burtin, P., Goéré, D., Seufferlein, T.,
310 Haustermans, K., Van Laethem, J. L., Conroy, T., Arnold D., 2015. Cancer of the pancreas:
311 ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann. Onc.* 26,
312 Suppl. 5, 56-68.
- 313 Jana, S., Shekhawat, G.S., 2011. Critical review on medicinally potent plant species: *Gloriosa*
314 *superba*. *Fitoterapia* 82, 293-301.
- 315 Kunnumakkara, A.B., Sung, B., Ravindran, J., Diagaradjane, P., Deorukhkar, A., Dey, S., Koca, C.,
316 Yadav, V.R., Tong, Z., Gelovani, J.G., Guha, S., Krishnan, S., Aggarwal, B.B., 2010. Gamma-tocotrienol inhibits pancreatic tumors and sensitizes them to gemcitabine treatment by
317 modulating the inflammatory microenvironment. *Cancer Res.* 70, 8695-8705.
- 318 Li, D., Xie, K., Wolff, R., Abbruzzese, J.L., 2004. Pancreatic cancer. *Lancet* 363 (9414), 1049-1057.
- 319 Maroyi, A., Van der Maesen, J.L.G., 2011. *Gloriosa superba* L. (family Colchicaceae): Remedy or
320 poison? *J. Med. Plants Res.* 5, 6112-6121.
- 321 Montgomery C.A.J., 1990. Oncological and toxicological research: Alleviation and control of pain and
322 distress in laboratory animals. *Cancer Bulletin* 42, 230-237.
- 323 Risinger, A.L., Giles, F.J., Mooberry, S.L., 2009. Microtubule dynamics as a target in oncology.
324 *Cancer Treat. Rev.* 35, 255-261.
- 325 Slobodnick, A., Shah, B., Pillinger, M.H., Krasnokutsky, S., 2015. Colchicine: old and new. *Am J.*
326 *Med.* 128, 461-470.

- 328 Solak, Y., Acikgoz, S.B., Yildirim, M., 2015. Colchicine Toxicity: An Exaggerated Reality? Am J.
329 Med. 128, e11.
- 330 Stanton, R.A., Gernert, K.M., Nettles, J.H., Aneja, R., 2011. Drugs That Target Dynamic
331 Microtubules: A New Molecular Perspective. Med. Res. Rev. 31, 443-481.
- 332 Wagner, H., Ulrich-Merzenich, G., 2009. Synergy research: Approaching a new generation of
333 phytopharmaceuticals. Phytomed.16, 97–110.
- 334 Yu, J., Chen, Q., 2014. Antitumor Activities of *Rauwolfia vomitoria* Extract and Potentiation of
335 Gemcitabine Effects Against Pancreatic Cancer. Integr. Cancer Ther. 13, 217-225.
- 336 Yu, J, Drisko, J, Chen, Q., 2013. Inhibition of pancreatic cancer and potentiation of gemcitabine
337 effects by the extract of Pao Pereira. Oncol. Rep. 30, 149-156.
- 338 Yue, Q.X., Liu, X.A., Guo, D.A., 2010. Microtubule-Binding Natural Products for Cancer Therapy.
339 Planta Med. 76, 1037-1043.
- 340