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***Hevea* latex-associated allergies: piecing together the puzzle of the latex IgE reactivity profile.**

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Running title: Molecular diagnosis of latex allergy.

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Running title: IgE-reactivity profile of latex sensitization

1 Abstract

2 Introduction: IgE-mediated *Hevea* latex allergy and associated food-allergies constitute a significant
3 health issue with serious consequences of diagnostic error. Hence, there is need for more reliable
4 confirmatory diagnostics.

5 Areas covered: Here, we summarize the major limitations of conventional tests using native extracts and
6 describe how piecing together the IgE reactivity profile can benefit correct diagnosis in difficult cases
7 in whom conventional tests yield equivocal or negative results. A diagnostic algorithm integrating
8 traditional sIgE and component resolved diagnosis (CRD) is presented.

9 Expert opinion: Moreover, it is clear that the discoveries in the field of the *Hevea* latex proteome will
10 contribute to our understandings and accurate approach of sometimes complex cross-reactivity
11 phenomena that extend beyond the “latex-fruit syndrome”.

12

13 1. Introduction

14 The term “latex” is often used for the elastic product employed in the composition of rubber articles.
15 Generically, “latex” refers to an aqueous elastomer emulsion, and in the case of natural rubber, the
16 natural latex is drawn from *Hevea brasiliensis* (order Euphorbiales, family Euphorbiaceae) as a milky
17 sap. Natural latex is the cytoplasm of specialised plant cells called laticifers and form a tube-like network
18 through the plant, and functions to seal and protect damaged sites. The approximate composition of the
19 liquid natural latex is water (55-65%), *cis*-1,4-polyisoprene rubber (34%), sugars (1.0-2.0%), sterol
20 glycosides (0.1-0.5%), resins (1.5-3.5%), ash (0.5-1.0%) and finally proteins (2-3%) [1]. The latter
21 causing immunoglobulin (Ig)E-mediated latex allergy and associated cross-allergies, particularly to
22 fruits and vegetables but also nuts and cereals (reviewed in [2-5]). Three different fractions can be
23 obtained by high-speed centrifugation of natural latex. There is a white creamy layer of rubber particles
24 at the top. This layer is also called the “rubber phase” and contains approximately 27% of total protein
25 in *Hevea* latex. These proteins are called rubber particle-associated proteins, i.e. the large particle-
26 associated rubber elongation factor (REF) and the small rubber particle protein (SRPP). The bottom
27 fraction (B-serum), containing specialised cell organelles are collectively called “lutoids” and have an
28 approximate total protein percentage of 25%. As shown in table 1, lutoids contains several hydrolases
29 and some pathogenesis-related proteins (i.e., defence proteins). Finally, the yellowish C-serum in
30 between, corresponds to the cytosol from the laticifer cells, and contains about 48% of the total protein
31 [6].

32 To date, IgE-mediated latex allergy to the constituent proteins of *Hevea* latex is recognized as an
33 international health problem of major importance. Although the exact prevalence of latex sensitization
34 and allergy among the general population is estimated less than 1%, several risk groups such as spina
35 bifida patients and health care workers who are regularly exposed to latex-containing devices have been
36 identified [2, 7, 8]. Besides, several epidemiological surveys have identified IgE-mediated latex allergy
37 as significant cause of anaesthesia-related allergy and anaphylaxis [9, 10] . In these patients, correct

38 diagnosis of IgE-mediated latex allergy is a prerequisite to avert future potentially life-threatening
39 reactions to latex and potential cross-reactive allergens implicated in latex-associated food allergies. In
40 this review we focus on the potential and limitations of traditional latex-specific IgE (latex-sIgE)
41 quantification and component resolved diagnosis to document IgE-mediated latex allergy.

42

43 2. In vitro diagnosis of IgE-mediated latex allergy

44 In general clinical practice, many physicians rely upon quantification of latex-sIgE antibodies as a
45 primary measure to confirm or discard their clinical suspicion of an IgE-mediated latex allergy.
46 However, correct diagnosis of latex allergy via quantification of latex-sIgE can pose significant
47 difficulties. On several occasions, it has been demonstrated that latex-sIgE results are not absolutely
48 predictive for the clinical outcome. Results of latex-sIgE can be false-negative [11-13] or, much more
49 frequently, false-positive, that is, clinically irrelevant [11-16]. The consequences of false-negative
50 results are obvious, as these entail a risk for life-threatening anaphylaxis upon subsequent exposure.
51 However, over-diagnosis by false-positive results can also have dramatic consequences. For example,
52 during diagnostic work-up of perioperative anaphylaxis, clinical irrelevant results could erroneously
53 lead to the diagnosis of IgE-mediated latex allergy and premature stopping of further testing for the true
54 culprit. Besides, identification of clinically irrelevant latex-sensitization should prevent unnecessary and
55 generally expensive latex avoidance measures. Hence, there is need for additional reliable confirmatory
56 tests.

57

58 3. Principles of Component Resolved Diagnosis

59 Traditional latex-sIgE assays, are based upon the quantification of serum sIgE directed against crude
60 natural allergen extracts (figure 1). The complexity, variability and instability of natural allergens and
61 the variation between individual sensitization patterns complicate the correct interpretation of sIgE
62 results to crude allergen extracts. Consequently, a positive sIgE against crude extracts should always be
63 interpreted with care as it might merely reflect (cross)sensitization rather than a genuine allergy. For
64 example, for latex, it has been shown that ubiquitous structures such as α -1,3-fucose and β -1,2-xylose
65 bearing cross-reactive carbohydrate determinants (CCD) present on glycoproteins of plants and α -1,3-
66 fucose bearing CCD of hymenoptera venom glycoproteins [14-19] and plant profilins [15-17, 19-23]
67 can elicit a false-positive latex-sIgE results. Therefore, latex-sIgE should not be used in isolation to
68 diagnose IgE-mediated latex allergy. Strategies that can be adopted to detect and circumvent the CCD
69 and profilin issues, are the use of glycan and profilin biomarkers or inhibitors, latex basophil activation
70 tests (BATs) [14], CRD applying non-glycosylated latex-specific components, or BATs with
71 recombinants [24].

72 The principles as well as major applications of CRD in children and adults are reviewed extensively
73 elsewhere [6, 25-30]. . In contrast to conventional sIgE assays, CRD does not rely upon crude extract
74 preparations obtained from native allergens but on sIgE antibodies directed towards single components

75 purified from natural sources or produced by recombinant techniques. In other words, CRD involves
76 specific marker components and substructures to study the genuine allergic sensitization of patients to
77 a particular allergen source and sensitization to cross-reactive determinants or components that point to
78 cross-sensitization. These so-called “gatekeeper” tests allow an improved discrimination between
79 genuine allergy and merely clinically irrelevant sensitization and allow the establishment of personalized
80 sensitization profiles. Determining such personalized sensitization profiles creates the opportunity to
81 assess the individual risk of severity of an allergic reaction and to predict the natural course. For
82 example, for latex, CRD has unveiled that health care workers and spina bifida patients display distinct
83 sensitization profiles that are not equally associated with a latex-food syndrome (see below). However,
84 CRD also demonstrates limitations. Not all relevant allergen components are available and it has been
85 demonstrated the technique to be of limited use in determining the clinical relevance of sensitization to
86 homologues of the major birch pollen allergen Bet v 1 [31-33] . Besides, when using these individual
87 components or epitopes for the diagnosis of allergy, the number of tests required to enable a correct
88 diagnosis increases significantly since more than one component needs to be included to allow
89 identification of the entire repertoire of disease relevant peptides and epitopes. The microarray technique
90 for CRD elegantly enables sIgE antibody testing in a multiplex format and allows the simultaneous
91 quantification of many sIgE antibodies. The major advantage of this multiplex technique lies in its
92 potential to study significant numbers of components in parallel, detecting sIgE antibody abundance,
93 functionality, and interaction concerning numerous allergenic determinants using only minute amounts
94 of patients' serum which is particularly important in infants and children.. It is anticipated that CRD by
95 flexible allergen-coated microbead assays, as shown in [figure 2](#), should allow a personalized selection
96 of the components of interest can benefit correct diagnosis in the individual patient [34]. Note that the
97 availability of allergenic components of *Hevea* latex can also benefit sensitivity of the conventional
98 latex-sIgE as has been demonstrated by comparison between a latex-sIgE with and without spiking for
99 the acidic protein of *Hevea brasiliensis* latex (Hev b 5) [35, 36].

100

101 [4. Component Resolved Diagnosis for IgE-mediated latex allergy](#)

102 As addressed in the section above, correct serologic diagnosis of IgE-mediated latex allergy by
103 conventional latex-sIgE testing can be seriously be impeded mainly because of the interference of anti-
104 plant/invertebrate CCD and anti-profilin antibodies that can easily be observed in up to one-quarter of
105 patients with a pollen and/or hymenoptera venom allergy [17]. As shown in the table, today 15
106 components of latex from *Hevea brasiliensis* (Hev b) have been identified and successfully cloned.
107 Some of them have become available for single or multiplex molecular diagnosis of IgE-mediated latex
108 allergy. As a matter of fact, the commercially available component-specific IgE assays for natural latex
109 are non-glycosylated recombinant (r) Hev b 1, 3, 5, 6.01, 6.02, 8, 9 and 11. Particularly, rHev b 5 and 6
110 and in a lesser extent also rHev b 1 and 3 (both rubber particle-associated proteins) have been shown to
111 be the most important biomarkers to diagnose genuine IgE-mediated latex allergy [37-41]. Sensitization

112 to Hev b 5 and 6 is primarily found in adult health care workers (HCW) and to a lesser extent also in
113 children suffering from spina bifida (SB) and meningomyelocele. In contrast, sensitization to *Hevea*
114 profilin Hev b 8 (latex profilin) generally, but certainly not always, points to a clinically irrelevant cross-
115 reactivity [16-19, 21, 22]. For example, in an own series, in all patients diagnosis of IgE-mediated latex
116 allergy could be established by the combination rHev b 1, 3, 5 and 6.02. Over three-quarters of our
117 patients were sensitized to rHev b 5 and/or 6.02. Some also displayed sIgE reactivity against rHev b 1
118 and/or rHev b 3. In contrast, none of the individuals showing a clinically irrelevant sensitization to
119 natural rubber latex demonstrated IgE reactivity to one of these components but three-quarters of them
120 displayed a positive microarray result for rHev b 8 [15]. However, recently we identified some patients
121 with an overt IgE-mediated latex allergy apparently related to monosensitization to Hev b 12, the non-
122 specific lipid transfer protein of latex [42]. The main reason(s) for Hev b 12 monosensitization remain(s)
123 elusive but could to some extent relate to an underlying *Cannabis sativa* allergy [43]. As all available
124 latex components are non-glycosylated proteins, they constitute a helpful instrument to depict clinically
125 irrelevant positive sIgE latex results resulting from a sensitization to plant-derived and invertebrate
126 CCD.

127

128 5. Component Resolved Diagnosis for latex-associated food allergy (initially designated as latex-fruit 129 syndrome)

130 As described above, patients suffering from an IgE-mediated latex allergy can display distinct
131 sensitization profiles and clinical phenotypes, that is, with or without cross-allergies. A large majority
132 of latex allergic patients is sensitized to so-called defence and/or structural proteins. These proteins are
133 quite ubiquitously distributed in plant kingdom and might explain the occurrence of a variety of latex-
134 associated plant food allergies historically designated as “latex-fruit syndrome” mainly involving
135 banana, avocado and chestnut [44]. However, today it appears that the list of cross-reactive plant-derived
136 foods extends far beyond these tropical foods and involves many fruits, vegetables, nuts and cereals [2-
137 5]. Such latex-associated food allergies have been described in about 21-58% of patients with an IgE-
138 mediated latex allergy. In contrast, other patients, mainly children with spina bifida, are sensitized to the
139 rubber particle-associated proteins Hev b 1 (REF) and Hev b 3 (SRPP), which by definition are confined
140 to rubber synthesising plants and will not display a secondary latex-associated food syndrome. Like for
141 all IgE-mediated diseases, the diagnostic approach of latex-associated food allergy starts with a thorough
142 clinical history with a main focus on the latex allergy and potential related cross-reactivities and should
143 further be pieced together using different *in vitro* and *in vivo* tests. As addressed in the section “*in vitro*
144 diagnosis of IgE-mediated latex allergy”, in general clinical practice, most physicians will use
145 quantification of latex-sIgE antibodies as a primary confirmatory diagnostic. However, interpretation of
146 positive latex-sIgE results is not always straightforward and correct diagnosis might require additional
147 testing, mainly because of significant interference of clinically irrelevant anti-plant/invertebrate CCD
148 and anti-profilin sIgE antibodies. Moreover, it has repeatedly been shown these antibodies, together with

149 sIgE antibodies to the homologues of Bet v 1 (the major allergen from birch (*Betula verucosa*) and non-
150 specific lipid proteins (ns-LTP), to severely hamper correct diagnosis of IgE-mediated plant-derived
151 food allergies. Therefore, the introduction of CRD to establish the individual sensitization profile should
152 not only benefit correct diagnosis of IgE-mediated latex allergy but might, to some extent, help accurate
153 diagnostic and therapeutic management of patients with a clinical suspicion of a latex-associated food
154 syndrome [2-6]. For example, we found sIgE antibodies to the natural extracts of various fruits,
155 vegetables and ficus to be clinically irrelevant in a majority of latex allergic patients [45]. Briefly, as
156 indicated in the diagnostic algorithm (figure 3), allergens with potential importance for IgE-mediated
157 plant-derived food allergies secondary to an IgE-mediated latex allergy are Hev b 5, 6, 7, 8, 11 and 12
158 [2-6]. Mono-sensitization to Hev b 1, 3 and 4 is unlikely to be associated with cross-reactivity,
159 sensitization to Hev b 9 and 10 can be accompanied by cross-reactivity to moulds [46, 47] and
160 sensitization to Hev b 6 and Hev b 11 with cross-reactivity to ficus species [48]. Sensitization to Hev b
161 2 and Hev b 13 seems less significant [49].

162

163 6. Expert opinion

164 IgE-mediated latex allergy constitutes a significant medical health problem that requires correct
165 diagnosis for adequate and potentially lifesaving management, that is, avoidance measures. On the other
166 hand, erroneous overdiagnosis of IgE-mediated latex allergy should be avoided, mainly because of the
167 cost of alternative elastomers. Unfortunately, correct diagnosis is not always straight forward and can
168 pose significant difficulties. Today, the most important limitation of the conventional latex-sIgE assays
169 remains the high number of false-positives due to interference by clinically irrelevant anti-CCD and
170 anti-profilin sIgE antibodies. To some extent, these difficulties have been solved by basophil activation
171 experiments [14, 15] but mainly by the characterization and production of an increasing number of
172 native and mainly recombinant allergenic *Hevea* components, with some being available for sIgE assays
173 and more laborious basophil activation experiments. The performance of these component-based
174 diagnostics has been thoroughly explored and quintessence of these studies is clear. The continuous
175 efforts in unravelling the *Hevea latex* proteome with characterization of relevant allergens and
176 availability of recombinant components (free of profilin and plant/invertebrate glycans) has enabled the
177 development for a more precise and approach of the individual patients. As a matter of fact, CRD offers
178 a more reliable diagnostic and has paved the way to broaden our knowledge in sometimes complex
179 cross-reactivity syndromes.

180 It is likely the further exploration of the latex proteome to disclose novel (less abundant) allergenic
181 components to benefit precise diagnosis and to shift paradigms about the mechanisms of cross-reactivity
182 syndromes. Whether profiling epitope-specific antibody repertoires will deepen our understandings in
183 the mechanisms behind latex-associated allergies and benefit prediction of severity and phenotypes
184 remains to be established.

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191 **Legends of figures**

192

193 **Figure 1: Source, application and composition of *Hevea* latex.**

194 Generically, “latex” refers to an aqueous elastomer emulsion, and in the case of natural rubber, the
195 natural latex is drawn from *Hevea brasiliensis* (order Euphorbiales, family Euphorbiaceae). *Hevea* latex
196 has many applications as the production of dipped thin-film materials such as balloons and gloves. The
197 approximate composition of the natural latex is water (55-65%), *cis*-1,4-polyisoprene rubber (34%),
198 sugars (1.0-2.0%), sterol glycosides (0.1-0.5%), resins (1.5-3.5%), ash (0.5-1.0%) and finally proteins
199 (2-3%) of which 15 have currently been identified and successfully cloned (designated as Hev b 1-15).
200 Sensitization to some components is associated with severe clinical phenotypes whereas sensitization to
201 other components generally results to milder symptoms or is asymptomatic (e.g. Hev b 8 the profilin of
202 *Hevea* latex). Next to the allergenic components *Hevea* latex contains many other proteins of unknown
203 significance.

204

205

206 **Figure 2: Principle of measurement of specific IgE (sIgE) antibodies by cytometric bead technique.**

207 In the cytometric bead assay, allergens or components thereof are covalently coupled on beads of the
208 same size and color but with different color intensity. These coated-beads are incubated with patient’s
209 serum that contain sIgE antibodies. Subsequently, a fluorochrome-conjugated antihuman-IgE antibody
210 is added. This secondary antibody will bind the antigen-antibody immune complex and can be measured
211 in a flow cytometer. The intensity of the bead determines the antigen/allergen (y-axis), the intensity of
212 the fluorochrome conjugated antibody defines the sIgE concentration.

213 **Figure 3: diagnostic algorithm of *Hevea* latex-associated allergies**

214 Confirmatory testing generally starts with quantification of conventional latex-specific IgE (sIgE)
215 antibodies and/or latex skin prick test (SPT) using native extracts. If these tests yield negative results
216 latex allergy is unlikely. A basophil activation test (BAT) is recommended when history is compelling.
217 If conventional latex-sIgE and/or SPT and/or BAT is/are positive component resolved diagnosis starts
218 with quantification of sIgE to rHev b 5, 6, as well as 1 and 3 to determine clinical significance of
219 conventional latex-sIgE/SPT or BAT and to estimate the risk for a latex-food syndrome. If sIgE to rHev
220 b 1, 3, 5, 6, are negative it is advised to quantify sIgE to *Hevea* latex profilin (rHev b 8) and the glycan
221 biomarker MUXF3. If one of these, or both, is/are positive, a latex allergy is unlikely and sIgE to rHev
222 b 9 and 11 can be quantified, mainly to identify patients at risk for a latex-food syndrome because of
223 sensitization to the class I endochitinase Hev b 11. Note that if a patient tests positive for a recombinant
224 MBP-Hev b component, the clinical relevance has to be confirmed with a negative MBP result,
225 especially if this patient displays a high total IgE value.

226

Allergen	Trivial name	Localization	MW (kDa)	pI	Gly	Predicted physiological role	Glove users	Spina bifida	References
<i>Hev b 1</i>	Rubber elongation factor	Large rubber particles	14.7	5.0	-	Rubber synthesis	Minor	Major	[50]
<i>Hev b 2</i>	β -1,3 glucanase	Lutoids	35.1	9.5	+	Defence-related protein	Minor	Minor	[49, 51, 52]
<i>Hev b 3</i>	Small rubber particle protein	Small rubber particles	22.4	4.8	-	Rubber synthesis	Minor	Major	[53]
<i>Hev b 4</i>	Lecithinase homologue	Lutoids	53-55	4.5	+	Microhelix component	Minor	Minor	[54, 55]
<i>Hev b 5</i>	Acidic latex protein	Cytoplasm	16	3.5	-	Structural protein	Major	Major/minor	[56]
<i>Hev b 6</i>	Hevein and its precursors*	Lutoids	21	5.6	-	Lectin, latex coagulation	Major	Minor	[52, 57-59]
<i>Hev b 7</i>	Patatin homologue (esterase)	Lutoids Cytoplasm	42 44	4.8	+	Defence-related protein	Minor	Minor	[60, 61]
<i>Hev b 8</i>	Profilin	Cytoplasm	15	4.9	-	Cytoskeletal actin binding	Minor	Minor	[62]
<i>Hev b 9</i>	Enolase	Cytoplasm	47.7	5.6	-	Glycolytic enzyme	Minor	Minor	[47]
<i>Hev b 10</i>	Superoxide dismutase (MnSOD)	Mitochondria	26	6.3	-	Enzyme, radical destruction	Minor	Minor	[46, 63]
<i>Hev b 11</i>	Class I endochitinase	Lutoids	33	5.1	-	Defence-related protein	Minor	Minor	[64, 65]
<i>Hev b 12</i>	Nonspecific lipid transfer protein 1	Latex membranes	9.3	10.8	+	Defence-related protein	Minor	Minor	[66, 67]
<i>Hev b 13</i>	Esterase / early nodule specific protein	Lutoids	43	5.0	+	Defence-related protein	Minor	Minor	[49, 68]
<i>Hev b 14</i>	Hevamine (chitinase)	Lutoids	29.5	8.4	-	Defence-related protein	Minor	Minor	[69, 70]
<i>Hev b 15</i>	Serine protease inhibitor	Cytoplasm	7.5	4.8	-	Defence-related protein	Minor	Minor	[71, 72]

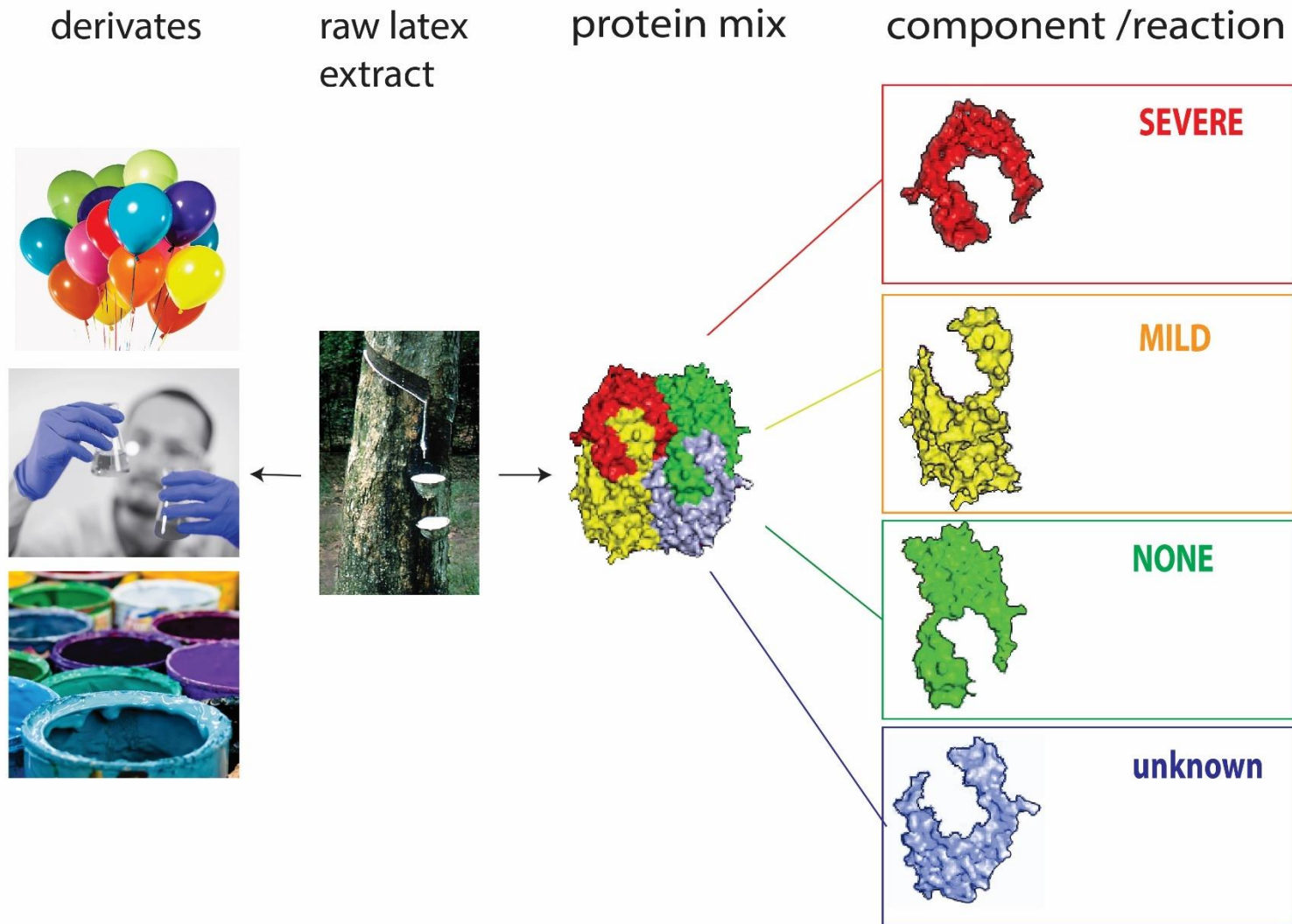
Commercially available components for sIgE testing are out in *italics*. * Hev b 6 comprised initially the 21 kDa precursor prohevein (Hev b 6.01), the 4.7 kDa hevein (Hev b 6.02), and the 14 kDa C-domain of prohevein (Hev b 6.03). Actually, the commercial used name Hev b 6 is synonymous with Hev b 6.02. Lutoids = B-serum, Cytoplasm = C serum. Gly: glycosylation. For accession N° see [73].

For more details: see <http://iuis.org>

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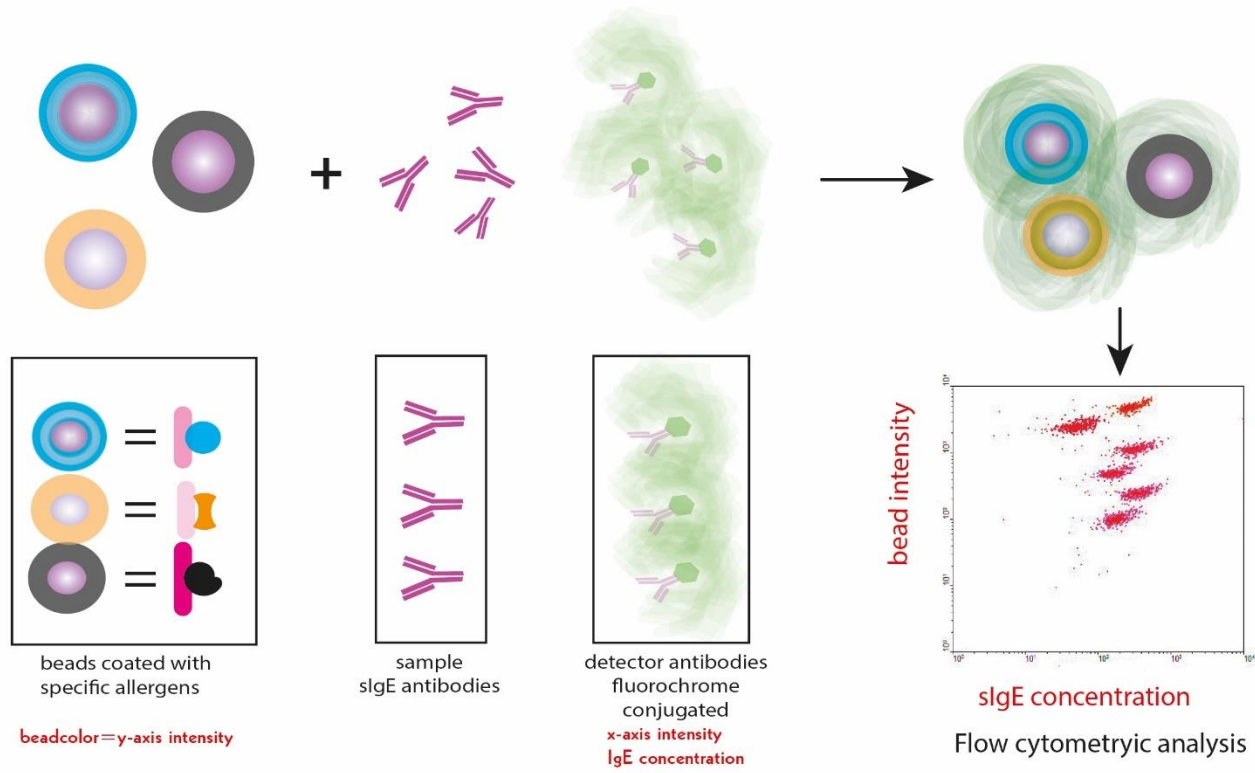
229 **Figure 1:**



231

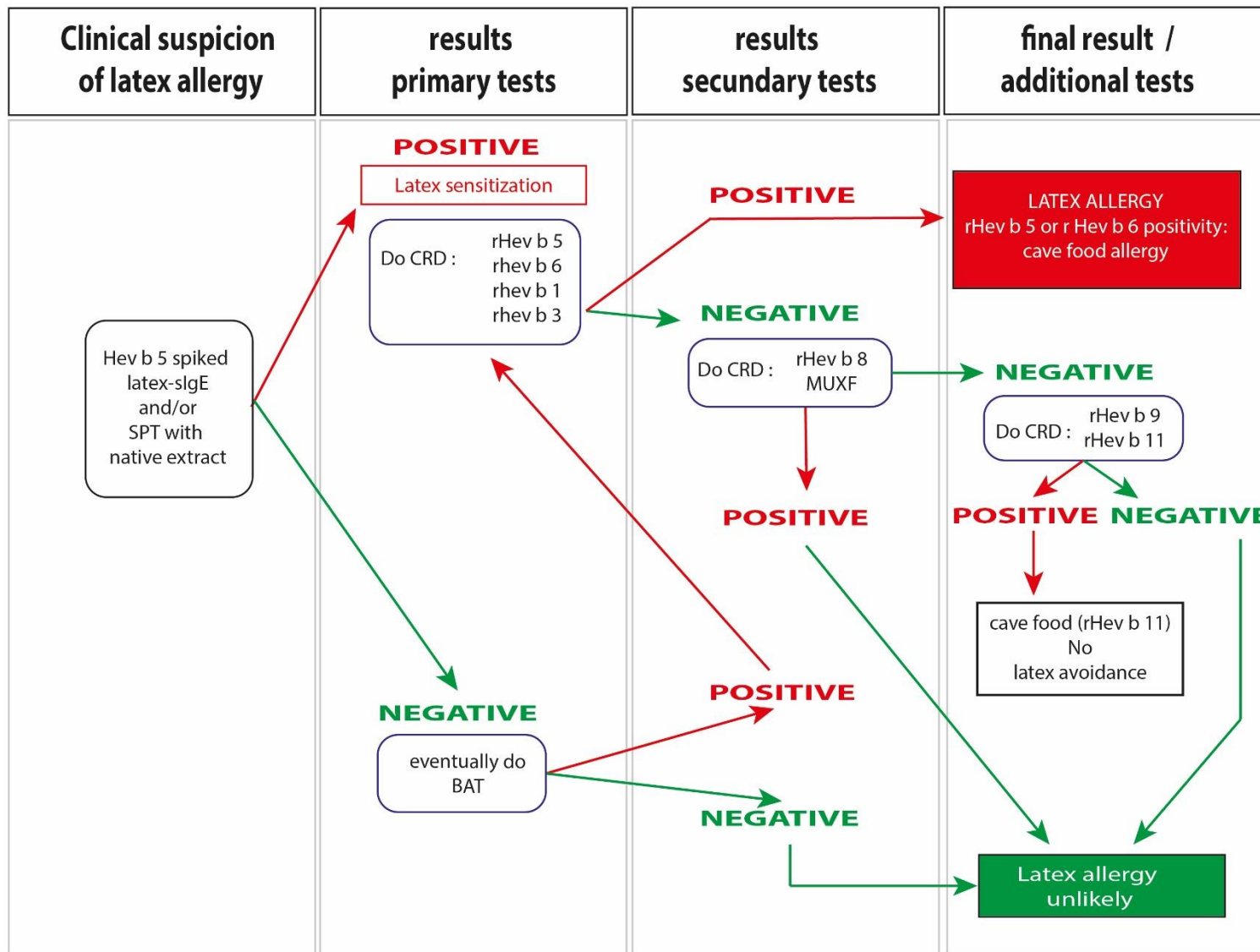
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233 **Figure 2:**



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240 **Figure 3**



242 **Declaration of fundings and conflict of interest**

243

244 The authors declare to have received no fundings and to have no conflict of interest

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249 **References**

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