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Chemical communication in the lacertid lizard ****Podarcis muralis**** : the functional significance of testosterone

Reference:

Baeckens Simon, Huyghe Katleen, Palme Rupert, Van Damme Raoul.- Chemical communication in the lacertid lizard ****Podarcis muralis**** : the functional significance of testosterone

Acta zoologica - ISSN 0001-7272 - 98:1(2017), p. 94-103

Full text (Publisher's DOI): <http://dx.doi.org/doi:10.1111/AZO.12160>

To cite this reference: <http://hdl.handle.net/10067/1328770151162165141>

1 TITLE: **Chemical communication in the lacertid lizard *Podarcis muralis*: the**
2 **functional significance of testosterone**

3 RUNNING HEAD: **Testosterone, and lizard chemical communication**

4

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33 Baeckens, S., Huyghe, K., Palme, R. and Van Damme, R. 2017. Chemical communication in
34 the lacertid lizard *Podarcis muralis*: the functional significance of testosterone. — *Acta*
35 *Zoologica* **98**: 94–103.

36

37 **Abstract.** Chemical signals are essential for intersexual communication in many animals,
38 including lizards. While faeces have been suggested to contain socially relevant chemical
39 stimuli, epidermal gland secretions are generally believed to be the leading source of
40 chemosignals involved in lizard communication. Early research has shown that sex hormones
41 affect epidermal gland activity, with androgens stimulating gland/pore size and/or gland
42 productivity. However, the functional significance of hormone-induced glandular activity in
43 lizard chemical communication remains unclear. In this study, we manipulated testosterone
44 (T) concentrations in male *Podarcis muralis* lizards. While T supplementation did not change
45 pore size, it did increase secretion production substantially. Chemosensory tests showed that
46 female conspecifics tongue-flick at a higher rate and more quickly towards the secretion of
47 males with experimentally-increased T levels than towards the secretion of control males,
48 suggesting that females can discriminate between males with dissimilar T levels based on
49 chemical cues of secretion alone. Based on the scent of faeces, however, females were unable
50 to discriminate between males with differential T levels. Also, females reacted more quickly
51 when offered larger amounts of secretion — irrespective of whether secretions were obtained
52 from control or T-increased males. This result indicates that secretion quantity affects
53 chemosignal detectability in *Podarcis muralis*.

54

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57

58 *Keywords:* chemical signals; epidermal glands; femoral secretions; pheromones, pores.

59

60

61 **Introduction**

62 Chemical signalling is the most widespread mode of communication between
63 organisms and plays a key role in intersexual communication in many vertebrate taxa
64 (Müller-Schwarze 2006; Wyatt 2014), including reptiles (Mason and Parker 2010). In
65 lizards, it is generally believed that epidermal gland secretions are the leading source
66 of chemosignals (Martín and López 2014; Mayerl *et al.* 2015), although there is some
67 evidence that faeces, cloacal secretions and skin lipids may also contain socially
68 relevant chemical stimuli (Cooper and Vitt 1984; Mason and Gutzke 1990; Cooper
69 1995; Labra 2008; Moreira *et al.* 2008). Epidermal glands of most lizards develop at
70 the onset of sexual maturity, and their activity is often greatest in male lizards and
71 during the reproductive season (Smith 1946; Cole 1966). These early observations
72 strongly suggest that the epidermal glands are controlled by androgens.

73 Experimental research indicates that castration of male lizards causes
74 glandular atrophy in *Lacerta agilis*, *Podarcis muralis* and *Cnemidophorus inornatus*
75 (Matthey 1929; Regamey 1932; Padoa 1933; Lindzey and Crews 1997). Testosterone
76 (T) supplementation restores gland activity in castrated *Goniurosaurus lichtenfelderi*
77 lizards (Golinski *et al.* 2015) and stimulates gland development in females of the
78 species *Gekko gecko* and *Urosaurus ornatus* (Chiu *et al.* 1970; Hews and Moore
79 1995). Oestrogen supplementation in males causes glandular atrophy in *Sceloporus*
80 sp. and *Hemidactylus bowringii*, a condition that can be reversed by exogenous
81 androgen administration (Forbes 1941; Chiu *et al.* 1975). Gland activity also
82 correlates strongly with testicular activity in *Sceloporus undulatus* and *Lacerta agilis*
83 (Regamey 1932; Altland 1941), and testis size in *Karusasaurus polyzonus* (Van Wyk
84 1990). Most of these studies have focused on proxies of glandular activity, such as
85 epidermal pore size (e.g. Hews and Moore 1995; Lindzey and Crews 1997; Rhen *et*

86 *al.* 2005); only a limited number of researchers scored secretion activity more directly
87 by measuring glandular size on histological sections (e.g. Chiu *et al.* 1975; Van Wyk
88 1990) or by collecting and weighing the produced secretion (e.g. Fergusson *et al.*
89 1985; Alberts *et al.* 1992). In addition, the functional significance of increased
90 glandular activity remains understudied. Martín *et al.* (2007b) established that T
91 implants alter the chemical composition of the epidermal secretions of male Iberian
92 wall lizards (*Podarcis hispanicus*) and that female conspecifics can discriminate
93 between T-implanted and control males based on the scent of male secretions. Martins
94 *et al.* (2006) found that seasonal increases in gland production enable male
95 *Sceloporus graciosus* lizards to scent-mark more localities rather than to mark any
96 single site more heavily, which may facilitate the expansion of a lizard's territory.
97 We, however, propose an additional advantage of the production of large amounts of
98 epidermal secretion in lizards. It is known that the absolute amount of volatile
99 compounds determines the absolute amount of airborne chemicals in a specific
100 environment (Apps *et al.* 2015). Based on the assumption that the proportion of
101 volatile compounds in the epidermal gland secretion remains constant with increasing
102 secretion quantity, we hypothesize that large amounts of secretion will increase the
103 active space of airborne chemosignals and, therefore, that high secretion quantities
104 will be detected more rapidly by lizard (vomero)olfaction.

105 Most research on chemical signalling in lizards has focused on chemosignals
106 originating from either epidermal gland secretions or faeces, while studies integrating
107 chemical cues of both origins are scant (but see e.g. Labra *et al.* 2002). Incorporating
108 multiple chemosignal sources within a study may shed light on the importance of
109 specific chemical sources, and on the multicomponent character of a message (Bro-
110 Jørgensen 2010).

111 In this study, we investigate the effect of epidermal gland secretion quantity
112 on signal transmission in a lacertid lizard, and test if chemosignals originating from
113 epidermal gland secretions and faeces convey similar information. We manipulate T
114 concentrations in male *Podarcis muralis* lizards using a non-invasive technique
115 described by Knapp and Moore (1997). Our objectives are (i) to quantify the effect of
116 T on the secretory activity of male epidermal glands, (ii) to test the ability of *Podarcis*
117 *muralis* females to discriminate between control males and males with manipulated T
118 levels based only on chemical cues (originating from epidermal secretion and faeces),
119 and (iii) to determine if the amount of epidermal secretion affects signal transmission
120 and detectability. The potential benefits of high secretion production rates in lizards
121 will be discussed.

122

123 **Material and methods**

124 *Animals and their maintenance*

125 The common wall lizard *Podarcis muralis* is a diurnal saxicolous lacertid (adult
126 snout-vent length (SVL) 51 – 65 mm; this study), widely distributed throughout
127 central and southern Europe (Gassert *et al.* 2013; Michaelides *et al.* 2015). *Podarcis*
128 *muralis* lizards have on average 15 pore-bearing scales on each thigh, the so-called
129 ‘femoral pores’ (Baeckens *et al.* 2015). Males of the species are highly territorial
130 (Stamps 1977; Edsman 1986) and use scent to mark their territory (Edsman 1990).
131 Based on chemical cues alone, female *Podarcis muralis* lizards are able to
132 discriminate between male conspecifics of different size (Heathcote *et al.* 2014).
133 Females of several lacertid species are known to exhibit chemically-mediated mate
134 choice (Martín and Lopéz 2015).

135 Early in their reproductive season (April 2015), 40 adult *Podarcis muralis*
136 lizards (21 males; 19 females) were captured by noose at two localities in the north of
137 Belgium ('Heverlee' and 'Muizen') and transported in cotton bags to our laboratory at
138 the University of Antwerp. Females lacked ventral mating scars, indicating that
139 females most probably had not mated yet. Males were individually housed in glass
140 terraria (50 x 40 x 50 cm) to avoid physical or visual contact between them, whereas
141 females were kept in group (maximum four females per 100 x 40 x 50 cm terrarium).
142 Each terrarium contained a layer of sandy substrate, some vegetation and slate stones.
143 A 60-watt bulb suspended above one end of the terrarium provided light (12L:12D)
144 and heat that enabled the lizards to maintain a body temperature within their preferred
145 range (i.e. between 30 °C – 35 °C; Bauwens *et al.* 1995). The larger terraria were
146 equipped with two light bulbs. Lizards had access to drinking water at all times, and
147 were fed moth larvae (*Galleria mellonella*) or crickets (*Acheta domesticus*) dusted
148 with multivitamin powder thrice a week.

149

150 *Hormonal treatments*

151 In this study, male hormone levels were manipulated using a non-invasive technique
152 based on the methodology described by Knapp and Moore (1997). Elevation of
153 sustained T levels was obtained by transdermal administration of a mixture of the
154 steroid hormone and sesame oil to the lizard. The lipid-rich character of the lizard
155 skin enables lipophilic molecules to pass through the scales of the integument into the
156 blood stream (Quay 1986). Unfortunately, protocolised T administration doses for
157 *Podarcis muralis* are not available in the literature. Therefore, we based our T dose on
158 a study by Belliure *et al.* (2004) ensuring that a chosen T dose increases T levels —
159 while maintaining levels within the species' estimated physiological range. In Belliure

160 *et al.* (2004), T levels were manipulated in two lacertid species, i.e. *Acanthodactylus*
161 *erythrurus* and *Psammodromus algirus*, which are both comparable to the common
162 wall lizard in body size. In accordance with their study, we created a hormone
163 solution by diluting testosterone powder (4-androsten-17 β -ol-3-one; Sigma #86500)
164 in commercial sesame oil (4 μ g testosterone / μ L sesame oil). Prior to the
165 experiments, we assigned males to one out of two treatment groups: a group with
166 experimentally manipulated T levels ('T-males', $n = 11$), or a control group ('C-
167 males', $n = 10$). Males from the two treatment groups did not differ in SVL
168 (ANOVA; $F_{1,20} = 0.090$; $P = 0.768$).

169 Males of the experimental T-group received 4 μ L of the hormone dilution
170 every two days over a period of four weeks (June 2015), while C-males were
171 administered the same amount of sesame oil as a control. Without disturbing the
172 lizards, a droplet of the T mixture or control oil (depending on the treatment group)
173 was deposited onto the lizard's dorsum using a volumetric pipet. To minimize stress,
174 administration was carried out in the early evening when the lights were off and the
175 lizards had become inactive. To validate the hormonal manipulation, we measured
176 male testosterone levels before and after the treatment period using a non-invasive
177 steroid analysis based on faecal droppings. Following extraction of the droppings with
178 60% methanol (0.5 mL per 20 mg; Palme *et al.* 2013) testosterone (metabolites) were
179 analysed with a testosterone enzyme immunoassay (details see Palme and Möstl
180 1994). Dry droppings were collected twice for every individual: two weeks before the
181 start of the treatment, and three weeks after the start of the treatment. We used forceps
182 for faecal collection, which were rinsed in 70% ethanol, to avoid cross contamination.
183 Any visible sand and uric acid remains on the dry droppings were manually scraped

184 off, and individual faecal samples were placed in labelled plastic bags and stored at -
185 20 °C.

186

187 *Femoral pore size and secretion mass*

188 Snout-vent length was measured using digital callipers (Mitutoyo, CD-15CPX,
189 accuracy = 0.01 mm). Female and male pore size was estimated by digitising (ImageJ,
190 Abràmoff *et al.* 2004) the circumference of each of the three most proximal pores of
191 the left femur on images obtained with a stereomicroscope (Leica M165 C). Only
192 male *Podarcis muralis* lizards possess active epidermal glands, but merely for
193 quantitative comparative purposes, we measured the pore size of individuals of both
194 sexes.

195 Next, we collected epidermal gland secretions (from all the glands of the left
196 thigh) of male lizards, by gently squeezing the femoral pores with a forceps until each
197 gland was completely emptied and all secretion yielded. Secretions were directly
198 weighed on a microbalance (Mettler Toledo MT5, accuracy = 1 μ g). The
199 measurements were conducted on the day preceding the hormonal treatment and
200 repeated 21 days after the start of the hormonal treatment. The obtained secretions
201 (after 21 days) were also used for the consecutive chemosensory tests.

202

203 *Chemosensory tests*

204 Squamates (amphisbaenians, lizards and snakes) react to a variety of chemical stimuli
205 with increased and differential rates of tongue extrusions. Tongue flick (TF) rates can,
206 therefore, be used as a quantitative bioassay for the degree of chemosensory
207 excitation (Burghardt 1970, 1980; Cooper 1994, 1998). To examine whether females
208 can distinguish between T- and C-males on the basis of their scent, we observed

209 female TF behaviour towards different stimuli presented on cotton-tipped applicators.
210 This technique is a widely used method in studies on chemical detection in squamates
211 (Cooper 1994, 1998, 2007). This particular part of the study was conducted 21 days
212 after the start of the T- or C-administration.

213 Cotton swabs were presented to 19 females. These swabs were impregnated
214 with (i) a single portion of male epidermal gland secretion (i.e. all secretion obtained
215 from all glands of one thigh), (ii) a double portion of male secretion (i.e. all secretion
216 obtained from all glands of both thighs) and (iii) male faeces. All three treatments
217 were carried out in twofold, i.e. with excretions extracted from C-males and from T-
218 males. Also, each female was subjected to an odourless control: a cotton swab
219 impregnated with deionized water (Cooper and Burghardt 1990). In total, each female
220 was subjected to seven different observation trials: one odourless control and six trials
221 with stimuli derived from six different males. We did not implement a pungency
222 control, as it is already well established that lacertid lizards can distinguish the scent
223 of conspecifics from ecologically irrelevant scents (e.g. Cooper and Pèrez-Mellado
224 2002). As a default preparation, we dipped the cotton tip of the wooden applicator in
225 deionized water before applying secretion or faeces onto the tip. Epidermal gland
226 secretions were extracted by pressing gently with forceps around the pores, and were
227 gathered on the swab after the secretions were weighed. Pending on the trial, we
228 collected secretion from only one (single portion) or both (double portion) thighs. For
229 the faecal trial, we made use of the fact that lizards often defecate when handled. As
230 such, fresh male faeces were applied onto the cotton swab.

231 Prior to the experiments, female lizards were placed individually in a novel
232 cage (50 x 25 x 30 cm), which contained slate stones under which the lizards could
233 seek refuge. Each female was exposed to the scent stimuli in a randomized order, and

234 with a minimum interval of 30 minutes between subsequent trials. Females were
235 solely subjected to the scent of males of their own population. All observations were
236 performed during the lizards' peak activity hours (10:00-16:00).

237 After preparation of the swab, one of the researchers (SB) quietly approached
238 the female's cage and moved the cotton swab slowly in a straight-line towards the
239 lizard until it was approximately 2 cm anterior to the snout (Fig. 1). Subsequently,
240 two variables were scored: (i) the time period — or 'latency' — to the first elicit TF,
241 starting from the moment the cotton swab was in position; (ii) the total number of
242 elicited TFs during 60 seconds, starting from the first TF.

243

244 *Statistics*

245 Prior to analyses, all variables were \log_{10} -transformed to meet assumptions of
246 normality (Shapiro-Wilks test with $W \geq 0.95$). Within each lizard, femoral pore size –
247 determined as pore surface area – did not differ between pore number one, two and
248 three (repeated-measure ANOVA, Wilk's $\lambda = 0.973$, $F_{2,19} = 0.234$, $P = 0.794$).
249 Therefore, the average size of all three pores was calculated for each individual and
250 used in further analyses. Relationships between average femoral pore size, secretion
251 mass and SVL were explored using Pearson's correlations. To test for changes in pore
252 size, secretion mass and testosterone levels before and after the treatment period, we
253 used repeated-measure ANOVAs with 'time' (before vs. after) as within-subject
254 variable, and 'treatment' (C vs. T) as between-subject variable testing for differences
255 in changes between C- and T-males. When necessary, an ANCOVA was used with
256 SVL as covariate. Repeated-measures ANOVAs were used to examine differences in
257 TF behaviour with 'scent stimuli' as within-subject factor. Pairwise comparisons were
258 carried out using Tukey's honestly significant difference (HSD) tests. All statistical

259 analyses were conducted in IBM SPSS Statistics for Macintosh v. 22.0 (Chicago, IL,
260 USA) and probabilities lower than 0.05 were considered statistically significant.

261

262 **Results**

263 *Validation of the hormonal treatment*

264 Prior to the administration period, T levels did not differ between C- and T-males
265 (average faecal steroid concentration \pm SE: 31.3 \pm 10.1 ng/g for C-males versus 30.4
266 \pm 6.0 ng/g for T-males; ANOVA, $F_{1,18} = 0.224$, $P = 0.642$). After three weeks of
267 treatment, lizards from the T-group showed a significant increase in faecal
268 testosterone concentration, whereas males from the control group exhibited even a
269 minor decrease in testosterone levels (repeated-measure ANOVA, ‘treatment’ x
270 ‘time’; $F_{1,18} = 11.484$, $P = 0.001$), with 10.7 \pm 5.6 ng/g for C-males versus 96.9 \pm 23.3
271 ng/g for T-males (Fig. 2). The small T-drop in the control group is most probably
272 linked with an inevitable stress increase due to captive conditions: a common
273 phenomenon, observed in various reptile taxa (e.g. Lance and Elsey 1986; Moore *et*
274 *al.* 1991).

275

276 *Femoral pore size and secretion mass*

277 Average femoral pore size differed significantly between sexes, with males having
278 larger pores than females (average pore size \pm SE: 638 \pm 45 μm^2 for males versus
279 364 \pm 18 μm^2 for females; ANOVA, $F_{1,39} = 44.596$, $P < 0.001$). Neither male nor
280 female pore size correlated with SVL (male, $r^2 = 0.033$, $P = 0.427$; female, $r^2 < 0.001$,
281 $P = 0.982$). Prior to the administration period, femoral pore size did not differ
282 between C- and T-males (ANOVA; $F_{1,20} = 0.729$, $P = 0.405$). After an administration
283 period of three weeks, pore size had not changed (repeated-measure ANOVA, ‘time’-

284 effect; $F_{1,20} = 1.325, P = 0.266$), and did not differ between treatments (repeated-
285 measure ANOVA, ‘treatment’-effect; $F_{1,20} = 1.439, P = 0.247$).

286 Larger males produced larger quantities of epidermal gland secretions ($r^2 =$
287 $0.263, P = 0.009$, Fig. 3). Prior to the administration period, the amount of secretion
288 that could be obtained from the males’ glands did not differ between the C- and the T-
289 group (average secretion mass \pm SE: 2.067 ± 0.204 mg for C-males versus 2.156 ± 0.302
290 mg for T-males; ANCOVA; $F_{1,20} = 0.001, P = 0.861$). After three weeks of C- and T-
291 administration, lizards from the T-group showed an increase in secretion production,
292 whereas C-males showed a decrease (repeated-measure ANCOVA, ‘treatment’ x
293 ‘time’; $F_{1,20} = 13.325, P = 0.002$; Fig 4). Compared to the sampling prior to the
294 administration period, the amount of secretion that could be obtained from the glands
295 was lower in C-males (-9.51%), and higher in T-males (+9,76%). Larger males did
296 not show a higher increase in secretion production (correlation of ‘difference in
297 secretion mass before-after’ over ‘SVL’, $r^2 = 0.008, P = 0.405$).

298

299 *Chemosensory tests*

300 All female lizards responded to swabs by tongue flicking. Mean latency to the first TF
301 differed significantly among conditions (repeated-measures ANOVA; Wilk’s $\lambda =$
302 $0.064, F_{6,13} = 31.612, P < 0.001$; Fig 5a). Latencies to the first TF in response to male
303 stimuli were significantly shorter than to water (Tukey post-hoc test, all $P < 0.002$).
304 Latency times in response to scent stimuli were similar for C- and T-males
305 (comparing secretions from one thigh: $P = 0.222$; from both thighs: $P = 0.772$; from
306 faeces: $P = 0.191$). However, females reacted more rapidly when offered large
307 amounts of epidermal secretions (from both thighs) than when offered smaller
308 quantities (from one thigh only) (comparing C-males: $P < 0.001$; comparing T-males:

309 $P < 0.001$). TF latency times did not differ with origin (one thigh, two thighs, faeces)
310 when stimuli were taken from C-males ($P \geq 0.191$). T-males faeces and secretions
311 from both thighs elicited earlier reactions than T-male secretions from one thigh ($P <$
312 0.001).

313 The mean number of TFs differed among stimulus conditions (repeated-
314 measures ANOVA, Wilk's $\lambda = 0.026$, $F_{6,13} = 81.282$, $P < 0.001$; Fig 5b). Swabs
315 impregnated with male epidermal gland secretions or faeces elicited significantly
316 more TFs than control swabs with water (all $P < 0.001$). Regardless of whether
317 secretions were obtained from a single thigh or from both, swabs with epidermal
318 secretions from T-males elicited more TFs than secretions from C-males ($P < 0.001$).
319 The amount of male secretion did not matter (quantity-effect in C-males: $P = 0.120$;
320 in T-males: $P = 0.164$). Females did not TF more when confronted with faeces from
321 T-males than with C-male faeces ($P = 0.253$). Similar numbers of TFs were
322 performed in response to C-male epidermal secretions and C-male faeces ($P = 0.120$),
323 but epidermal secretions of T-males elicited more TFs than T-male faeces ($P <$
324 0.001).

325

326 **Discussion**

327 *Testosterone and the epidermal gland system*

328 Firstly, the results of this study show that artificial T administration does not cause
329 any changes in the size of male femoral pores in *Podarcis muralis*. Pore size also did
330 not correlate with body size or secretion production. Interestingly, while the results of
331 this study does not support the idea that femoral pore size can be used as a proxy for
332 the intensity of secretion production in male wall lizards, studies on other lizard
333 species found opposite results. Femoral pore size in male green iguanas correlates

334 strongly with secretion mass, indicating that pore size probably provides a good
335 estimate of gland productivity in *Iguana iguana* (Alberts *et al.* 1992). More in line
336 with our results, Van Wyk (1990) did not find a relationship between testis volume
337 and femoral pore size in the cordylid lizard *Karusasaurus polyzonus*. Although, based
338 on histological section of the glands, the researcher did find a correlation between
339 testis volume and the depth of the epidermal glands. While we are not certain on the
340 true origin of these disparate findings, it is possible that they derive from interspecific
341 differences in gland morphology (e.g. tubular, tubulo-acinar, tubulo-alveolar), where
342 the form of the glands might determine if high secretory activity affect the expansion
343 of the pore-bearing scales. Detailed histological studies on this topic should be
344 encouraged. Nevertheless, pore size can easily be used as a tool for sex discrimination
345 in *Podarcis muralis*, for male femoral pores were almost twice the size of female
346 pores. Male pores are known to be notability larger than those of females in most
347 lizard taxa (Valdecantos *et al.* 2014; Baeckens *et al.* 2015; Mayerl *et al.* 2015). This
348 sexual difference may be due to increased plasma oestrogen levels associated with
349 female reproductive maturity, since administration of exogenous oestrogen inhibits
350 glandular activity in other lizard species (Forbes 1941; Chiu *et al.* 1975).

351 Secondly, our results show that T supplementation provokes substantial
352 changes in the secretory production of epidermal glands in male *Podarcis muralis*
353 lizards: males with artificially increased T concentrations produced on average 19.3%
354 more secretion than control males. In castrated *Ctenophorus ornatus*, daily T
355 injections incite a 23.4% increase in secretion production (Fergusson *et al.* 1985).
356 Research on seasonal T fluctuations and peak secretory activity in dominant *Iguana*
357 *iguana* males showed that during the month in which the highest plasma T
358 concentrations were observed (January), epidermal secretion production was

359 approximately 150% higher than during the period with the lowest T concentration
360 (May) (Alberts *et al.* 1992). Martins *et al.* (2006) have argued that an increased
361 secretion production during the mating season may help male *Sceloporus graciosus*
362 lizards to mark larger territories. As T levels are highest at the start of the
363 reproductive season (Manzo *et al.* 1994; Oppliger *et al.* 2004), they may stimulate
364 secretion production and, in some cases, lizard mobility (e.g. *Lacerta agilis*, Olsson *et*
365 *al.* 2000). Then, it could be assumed that as lizards move around more often, they
366 may release more glandular material over a larger area — which is made possible by
367 the high (T-induced) secretion production rate — and, therefore, expand their scent
368 marking space substantially. However, more research is necessary to confirm this.

369 In this study, large *Podarcis muralis* males produced larger amounts of
370 secretion than small males. Similarly, dominant *Iguana iguana* males (equipped with
371 large heads) produce gland secretions at a higher rate than subordinates with small
372 heads (Alberts *et al.* 1992). Because secretion production is most probably an
373 energetically costly affair (Martín and López 2015), it is very likely that only
374 individuals in a good condition can afford high secretory activity rates. Therefore,
375 large males with high T levels – which are often considered as most dominant
376 (Sinervo *et al.* 2000; Huyghe *et al.* 2007, 2009; Husak *et al.* 2009) and in best
377 condition (Tokarz 1985; Jakob *et al.* 1996), may produce high amounts of secretion,
378 and are thus able to mark more sites and create a more widespread territory than small
379 lizards. This is the case with sagebrush lizards (*Sceloporus graciosus*): territorially
380 aggressive males deposit high amounts of epidermal secretion, and this over a large
381 area (Martins *et al.* 2006).

382

383 *Testosterone and signal detectability*

384 Squamate chemoreception is said to work in a hierarchical fashion, with chemicals
385 being received initially through the nares, processed by the nasal organs, and finally
386 triggering tongue-flick mediated vomerolfaction (Halpern 1992; Cooper 1994;
387 Schwenk 1995). As such, different latencies to the first TF or the number of TFs
388 elicited toward stimuli indicate distinct chemosensory activities and, thus, stimuli
389 discrimination (Cooper 1994). Our chemosensory experiments suggest that female
390 *Podarcis muralis* can detect and distinguish between epidermal gland secretion of C-
391 and T-males, as they react more quickly and more strongly with TF behaviour to
392 secretion of T-males than to secretion of C-males. This result corroborates the
393 findings by Martín *et al.* (2007b) in the closely related *Podarcis hispanicus*. Females
394 may exploit T-induced differences in the chemical composition of the epidermal
395 gland secretion or changes in the concentrations of relevant chemicals in the
396 secretion. The ability to discriminate between differential T levels may be an
397 important tool in lizard mate assessment, as male T concentration is commonly
398 known to correlate strongly with various lizard performance traits, such as sprint
399 speed (Klukowski *et al.* 1998) and bite force (Husak *et al.* 2007; Huyghe *et al.* 2009,
400 2010), and with overall dominant behaviour (Greenberg and Crews 1990; Pratt *et al.*
401 1992).

402 Although the amount of epidermal secretion did not affect female TF rates,
403 females did respond more rapidly when offered double-loaded swabs —
404 irrespectively of whether secretions were obtained from C- or T-males. These
405 findings imply that the absolute amount of detectable airborne chemicals increases
406 with increasing secretion quantity. As the absolute amount of airborne chemicals in
407 the environment is determined by the amount of volatile chemicals in the secretion
408 (Apps *et al.* 2015), we can infer that an increase in secretion quantity corresponds

409 with an increase in the absolute amount of volatiles. High amounts of epidermal gland
410 secretion may, therefore, increase signal detectability, with lizards responding more
411 rapidly by vomerolfactory behaviour. These results indicate that secretion quantity —
412 rather than T-induced changes in the physico-chemical properties of the secretion —
413 affect chemosignal transmission and detectability in *Podarcis muralis* lizards.
414 Accordingly, Escobar *et al.* (2001, 2003) found that *Liolaemus* lizards occurring at
415 high altitudes and low latitudes are equipped to produce high amounts of secretion, a
416 hypothesized compensation strategy to prevent signal loss due to the harsh signalling
417 environment. In addition to changes in secretion quantity, changes in the chemical
418 profile of the secretion may affect signal detectability too. For example, natural
419 seasonal changes in T level concentrations increase the proportion of unsaturated
420 lipids in the secretions of male *Iguana iguana*, which are postulated to increase
421 volatility and detectability of secretions (Alberts 1992). A recent study by Martín *et*
422 *al.* (2015) established interpopulational differences in secretion composition in
423 *Podarcis hispanicus*, enabling maximum signal detectability (and persistence) under
424 different climatic conditions.

425 Our results also show that although *Podarcis muralis* females showed interest
426 in male faeces, they could not discriminate between males with dissimilar T levels
427 based on scent of faeces alone. Previous research has already established the ability of
428 lacertid lizards to discriminate chemicals from faeces, and even suggested faeces as a
429 player in intraspecific lizard communication. Tongue-flick assays showed that
430 Carpetan rock lizards (*Iberolacerta cyreni*) are able to distinguish between own and
431 conspecific faeces (López *et al.* 1998), and that Iberian rock lizards (*Iberolacerta*
432 *monticola*) react differently when confronted with juvenile or adult faecal deposits
433 (Moreira *et al.* 2008). Although faecal scent elicited substantial vomerolfactory

434 behaviour in female *Podarcis muralis* in this study, the fact that females were unable
435 to detect T level differences in faeces (while the T enzyme immunoassay revealed
436 clear T level variation in the faecal pellets), suggests that faeces may not only serve as
437 a chemical signal. It has been suggested that lizards use faecal deposits to mark their
438 territory, in which individuals exploit both the chemical and visual aspect of the mark
439 to convey information. Male *Iberolacerta cyreni* typically deposit their excrements on
440 rock-tops, which is thought to facilitate visual detection by conspecifics (López *et al.*
441 1998). After first locating and approaching the signal visually, a lizard could
442 investigate the mark more closely through chemosensory channels (Duvall *et al.*
443 1987; Alberts 1989). It is generally assumed that — when the lizard moves —
444 femoral pores scrape against the substrate and small portions of glandular material are
445 transferred onto the substrate (Maderson 1986; Jared *et al.* 1999; Khannoon *et al.*
446 2013; de Villiers *et al.* 2015). Following this line of thought, it is probable that sites
447 with faecal deposits will also be labelled with epidermal secretions. Martins *et al.*
448 (2006) found a strong association between the amount of epidermal secretion and the
449 amount of faecal matter on a certain marking spot in *Sceloporus graciosus*. It is,
450 therefore, reasonable to hypothesize that lizards use faecal deposits as an initial long-
451 range visual signal providing the first cue, and carrying minimal chemical information
452 (e.g. own vs. conspecific) once encountered. The neighbouring epidermal gland
453 secretion deposits may act as chemosignal-reservoirs containing more elaborate
454 information on an individuals' condition, e.g. hormonal level, parasitic infection,
455 immune response, diet quality or fighting ability (López *et al.* 2003; Labra 2006;
456 Martín and Lopéz 2006a,b; Martín *et al.* 2007a,b, 2008; Khannoon *et al.* 2010, 2011;
457 Kopena *et al.* 2014).

458

459 *In conclusion*

460 Our data demonstrate that testosterone affects epidermal gland activity in male
461 *Podarcis muralis* lizards with an increased secretory production, but without any
462 changes in the size of the pore-bearing scales. Based on the scent of male epidermal
463 gland secretion alone, and not on the scent of faecal deposits, female conspecifics are
464 able to discriminate between males with different testosterone levels using
465 vomerolfaction behaviour. Moreover, females react more quickly towards higher
466 amounts of secretion, implying that epidermal gland secretion quantity affects lizard
467 chemosignal detectability. Still, more research on lizard scent marking behaviour is
468 necessary to elucidate the importance of secretion quantity in a natural setting.

469

470 **Acknowledgments**

471 We thank J. Daans, T. Locus, J. Mertens, W. Müller, A. Sannen and J. Scholliers for
472 practical assistance, J. Meaney for linguistic proof reading, and D. Tibax for graphical
473 support. Also, our moms. Two anonymous reviewers made valuable comments,
474 which helped to improve the quality of the manuscript considerably. K. Huyghe is a
475 postdoctoral fellow of the FWO-Flanders. All work was done in accordance with
476 University of Antwerp (Belgium) animal welfare standards and protocols (ECD 2015-
477 09).

478

479 **References**

480 Abràmoff, M. D., Magalhães, P. J. and Sunanda R. J. 2004. Image processing with
481 ImageJ. *Biophotonics International* **11**: 36–42.
482 Alberts, A. C. 1989. Ultraviolet visual sensitivity in desert iguanas: implications for
483 pheromone detection. *Animal Behaviour* **38**: 129–137.

484 Alberts, A. C. 1992. Constraints on the design of chemical communication systems in
485 terrestrial vertebrates. *American Naturalist* **139**: 62–89.

486 Alberts, A. C., Pratt, N. C. and Phillips, J. A. 1992. Seasonal productivity of lizard
487 femoral glands: relationship to social dominance and androgen levels.
488 *Physiology & Behavior* **51**: 729–733.

489 Altland, P. D. 1941. Annual reproductive cycle of the male fence lizard. *Journal of*
490 *the Elisha Mitchell Scientific Society* **57**: 73–83.

491 Apps, P. J., Weldon, P. J. and Kramer, M. 2015. Chemical signals in terrestrial
492 vertebrates: search for design features. *Natural Product Reports* **32**: 1131–
493 1153.

494 Baeckens, S., Edwards, S., Huyghe, K. and Van Damme R. 2015. Chemical signalling
495 in lizards: an interspecific comparison of femoral pore numbers in Lacertidae.
496 *Biological Journal of the Linnean Society* **114**: 44–57.

497 Bauwens, D., Garland, T. Jr, Castilla, A. M. and Van Damme, R. 1995. Evolution of
498 sprint speed in lacertid lizards: morphological, physiological, and behavioral
499 covariation. *Evolution* **49**: 848–863.

500 Belliure, J., Smith, L. and Sorci, G. 2004. Effect of testosterone on T cell-mediated
501 immunity in two species of Mediterranean lacertid lizards. *Journal of*
502 *Experimental Zoology. Part A, Comparative Experimental Biology* **301**: 411–
503 418.

504 Bro-Jørgensen, J. 2010. Dynamics of multiple signalling systems: animal
505 communication in a world in flux. *Trends in Ecology & Evolution* **25**: 292–
506 300.

- 507 Burghardt, G. M. 1970. Chemical perception in reptiles. In: Johnston J. W., Moulton,
508 D. G. and Turk, M. (Eds): *Communication by Chemical Signals*, pp. 241–308.
509 Appleton-Century-Crofts, New York.
- 510 Burghardt, G. M. 1980. Behavioral and stimulus correlates of vomeronasal
511 functioning in reptiles: feeding, grouping, sex, and tongue use. In: Müller-
512 Schwarze, D. and Silverstein, R. M. (Eds): *Chemical signals. Vertebrates and*
513 *Aquatic Invertebrates*, pp. 275–301. Plenum Press, New York.
- 514 Chiu, K. W., Lofts, B. and Tsui, H. W. 1970. Effect of testosterone on the sloughing
515 cycle and epidermal glands of female geckos (*Gekko gekko*). *General and*
516 *Comparative Endocrinology* **15**: 12–19.
- 517 Chiu, K. W., Maderson, P. F. A., Alexander, S. A. and Wong, K. L. 1975. Sex
518 steroids and epidermal glands in two species of gekkonine lizards. *Journal of*
519 *Morphology* **118**: 119–135.
- 520 Cole, C. 1966. Femoral glands in lizards: A review. *Herpetologica* **22**: 199–205.
- 521 Cooper, W. E. 1994. Chemical discrimination by tongue-flicking in lizards: a review
522 with hypotheses on its origin and its ecological and phylogenetic relationships.
523 *Journal of Chemical Ecology* **20**: 439–487.
- 524 Cooper, W. E. 1995. Effects of estrogen and male head coloration on chemosensory
525 investigation of female cloacal pheromones by male broad-headed skinks
526 (*Eumeces laticeps*). *Physiology & Behavior* **58**: 1221–1225.
- 527 Cooper, W. E. 1998. Evaluation of swab and related tests as a bioassay for assessing
528 responses by squamate reptiles to chemical stimuli. *Journal of Chemical*
529 *Ecology* **24**: 841–866.

530 Cooper, W. E. 2007. Lizard chemical senses, chemosensory behavior, and foraging
531 mode. In: Reilly, S., McBrayer, L. and Miles, D. (Eds): Lizard Foraging
532 Behavior, pp. 237–270. Cambridge University Press, Cambridge.

533 Cooper, W. E. and Vitt, L. J. 1984. Detection of conspecific odors by the female
534 broad-headed skink, *Eumeces laticeps*. *Journal of Experimental Zoology* **229**:
535 49–54.

536 Cooper, W. E. and Burghardt, G. M. 1990. A comparative analysis of scoring
537 methods for chemical discrimination of prey by squamate reptiles. *Journal of*
538 *Chemical Ecology* **16**: 45–65.

539 Cooper, W. E. and Pérez-Mellado, V. 2002. Pheromonal discriminations of sex,
540 reproductive condition, and species by the lacertid lizard *Podarcis hispanica*.
541 *Journal of Experimental Zoology* **292**: 523–527.

542 de Villiers, A., Flemming, A. and Mouton, P. Le F. N. 2015. Generation glands of
543 cordylid lizards: mechanism of secretion transfer to the environment.
544 *Amphibia-Reptilia* **36**: 351–360.

545 Duvall, D., Graves, B. M. and Carpenter, G. C. 1987. Visual and chemical composite
546 signalling effects of *Sceloporus* lizard fecal boli. *Copeia* **4**: 1028–1031.

547 Edsman, L. 1986. Territoriality and resource defense in wall lizards (*Podarcis*
548 *muralis*). In: Roček, Z. (Eds): Studies in Herpetology, pp. 601–604. Charles
549 University, Prague.

550 Edsman, L. 1990. Territoriality and competition in wall lizards, PhD thesis.
551 University of Stockholm Press, Stockholm.

552 Escobar, C. A., Labra, A. and Niemeyer, H. M. 2001. Chemical composition of
553 precloacal secretions of *Liolaemus* lizards. *Journal of Chemical Ecology* **27**:
554 1677–1690.

555 Escobar, C. M., Escobar, C. A., Labra, A. and Niemeyer, H. M. 2003. Chemical
556 composition of precloacal secretions of two *Liolaemus fabiani* populations:
557 are they different? *Journal of Chemical Ecology* **29**: 629–638.

558 Fergusson, B., Bradshaw, S. and Cannon, J. 1985. Hormonal control of femoral gland
559 secretion in the lizard, *Amphibolurus ornatus*. *General and Comparative*
560 *Endocrinology* **57**: 371–376.

561 Forbes, T. R. 1941. Observations on the urogenital anatomy of the adult male lizard,
562 *Sceloporus*, and on the activation of implanted pellets of testosterone and of
563 estrone. *Journal of Morphology* **68**: 31–65.

564 Gassert, F., Schulte, U., Husemann, M., Ulrich, W., Rödder, D., Hochkirch, A.,
565 Engel, E., Meyer, J. and Habel, J. C. 2013. From southern refugia to the
566 northern range margin: Genetic population structure of the common wall
567 lizard, *Podarcis muralis*. *Journal of Biogeography* **40**: 1475–1489.

568 Golinski, A., Kubička, L., John-Alder, H. and Kratochvíl, L. 2015. Androgenic
569 control of male-typical behavior, morphology and sex recognition is
570 independent of the mode of sex determination: a case study on Lichtenfelder's
571 gecko (Eublepharidae: *Goniurosaurus lichtenfelderi*). *Hormones and Behavior*
572 **72**: 49–59.

573 Greenberg, N. and Crews, D. 1990. Endocrine and behavioral responses to aggression
574 and social dominance in the green anole lizard, *Anolis carolinensis*. *General*
575 *and Comparative Endocrinology* **77**: 246–255.

576 Halpern, M. 1992. Nasal chemical senses in reptiles: Structure and function. In: Gans,
577 C. and Crews, D. (Eds): *Biology of the Reptilia*, Vol. 18: Physiology E, pp.
578 424–532. University of Chicago Press, Chicago.

579 Heathcote, R. J. P., Bell, E., d’Ettorre, P., While, G. M. and Uller, T. 2014. The scent
580 of sun worship: basking experience alters scent mark composition in male
581 lizards. *Behavioral Ecology and Sociobiology* **68**: 861–870.

582 Hews, D. K. and Moore, M. C. 1995. Influence of androgens on differentiation of
583 secondary sex characters in tree lizards, *Urosaurus ornatus*. *General and*
584 *Comparative Endocrinology* **97**: 86–102.

585 Husak, J. F., Irschick, D. J., Meyers, J. J., Lailvaux, S. P. and Moore, I. T. 2007.
586 Hormones, sexual signals, and performance of green anole lizards (*Anolis*
587 *carolinensis*). *Hormones and Behavior* **52**: 360–367.

588 Huyghe, K., Vanhooydonck, B., Herrel, A., Tadić, Z. and Van Damme, R. 2007.
589 Morphology, performance, behavior and ecology of three color morphs in
590 males of the lizard *Podarcis melisellensis*. *Integrative and Comparative*
591 *Biology* **47**: 211–220.

592 Huyghe, K., Husak, J. F., Herrel, A., Tadić, Z., Moore, I. T., Van Damme, R. and
593 Vanhooydonck, B. 2009. Relationships between hormones, physiological
594 performance and immunocompetence in a color-polymorphic lizard species,
595 *Podarcis melisellensis*. *Hormones and Behavior* **55**: 488–494.

596 Huyghe, K., Husak, J. F., Moore, I. T., Vanhooydonck, B., Van Damme, R., Molina-
597 Borja, M. and Herrel, A. 2010. Effects of testosterone on morphology,
598 performance and muscle mass in a lizard. *Journal of Experimental Zoology*
599 *Part A: Ecological Genetics and Physiology* **313**: 9–16.

600 IBM Corp. Released 2013. IBM SPSS Statistics for Macintosh, Version 22.0. IBM
601 Corp. Armonk, New York.

602 Jakob, E. M., Marshall, S. D. and Uetz, G. W. 1996. Estimating fitness: a comparison
603 of body condition indices. *Oikos* **77**: 61–67.

604 Jared, C., Antoniazzi, M. M., Silva, J. and Freymuller, E. 1999. Epidermal glands in
605 Squamata: microscopical examination of precloacal glands in *Amphisbaena alba*
606 (Amphisbaenia, Amphisbaenidae). *Journal of Morphology* **241**: 197–206.

607 Khannoon, E. R., Breithaupt, T., El-Gendy, A. and Hardege, J. D. 2010. Sexual
608 differences in behavioural response to femoral gland pheromones of
609 *Acanthodactylus boskianus*. *Herpetological Journal* **20**: 225–229.

610 Khannoon, E. R., Flachsbarth, B., El-Gendy, A., Mazik, K., Hardege, J. D. and
611 Schulze, S. 2011. New compounds, sexual differences, and age-related variations
612 in the femoral gland secretions of the lacertid lizard *Acanthodactylus boskianus*.
613 *Biochemical Systematics and Ecology* **39**: 95–101.

614 Khannoon, E. R., Dollahon, N. R. and Bauer, A. M. 2013. Comparative study of the
615 pheromone-manufacturing femoral glands in two sympatric species of lacertid
616 lizards (*Acanthodactylus*). *Zoological Science* **30**: 110–117.

617 Klukowski, M., Jenkinson, N. M. and Nelson, C. E. 1998. Effects of testosterone on
618 locomotor performance and growth in field-active northern fence lizards,
619 *Sceloporus undulates hyacinthinus*. *Physiological Zoology* **71**: 506–514.

620 Knapp, R. and Moore, M. C. 1997. A non-invasive method for sustained elevation of
621 steroid hormone levels in reptiles. *Herpetological Review* **28**: 33–36.

622 Kopena, R., López, P. and Martín, J. 2014. Relative contribution of dietary
623 carotenoids and vitamin E to visual and chemical sexual signals of male Iberian
624 green lizards: an experimental test. *Behavioral Ecology and Sociobiology* **68**:
625 571–581.

626 Labra, A. 2006. Chemoreception and the assessment of fighting abilities in the lizard
627 *Liolaemus monticola*. *Ethology* **112**: 993–999.

- 628 Labra, A. 2008. Multi-contextual use of chemosignals by *Liolaemus* lizards. In: Hurst,
629 J. L., Beynon, R. J., Roberts, S. C. and Wyatt, T. D. (Eds): Chemical Signals in
630 Vertebrates 11, pp. 357–365. SpringerLink, New York.
- 631 Labra, A., Escobar, C. A., Aguilar, P. M. and Niemeyer, H. M. 2002. Sources of
632 pheromones in the lizard *Liolaemus tenuis*. *Revista Chilena de Historia Natural*
633 **75**: 141–147.
- 634 Lance, V. A. and Elsey, R. M. 1986. Stress-induced suppression of testosterone
635 secretion in male alligators. *Journal of Experimental Zoology* **239**: 241–246.
- 636 Lindzey, J. and Crews, D. 1997. Effects of progesterone and dihydrotestosterone on
637 stimulation of androgen-dependent sex behavior, accessory sex structures, and in
638 vitro binding characteristics of cytosolic androgen receptors in male whiptail
639 lizards (*Cnemidophorus inornatus*). *Hormones and Behavior* **27**: 269–281.
- 640 López, P., Aragón, P. and Martín, J. 1998. Iberian rock lizards (*Lacerta monticola*
641 *cyreni*) assess conspecific information using composite signals from faecal
642 pellets. *Ethology* **104**: 809–820.
- 643 López, P., Aragón P., and Martín, J. 2003. Responses of female lizards, *Lacerta*
644 *monticola*, to males' chemical cues reflect their mating preference for older
645 males. *Behavioral Ecology and Sociobiology* **55**: 73–79.
- 646 Maderson, P. F. A. 1986. Tetrapod epidermis: a system protoadapted as a
647 semiochemical source. In: Duvall D., Müller-Schwarze, D. and Silverstein, R.
648 M. (Eds): Chemical Signals in Vertebrates 4: Ecology, Evolution, and
649 Comparative Biology, pp. 13–26. Plenum Press, New York.
- 650 Manzo, C., Zerani, M., Gobbetti, A., Di Fiore, M. M. and Angelini, F. 1994. Is
651 corticosterone involved in the reproductive processes of the male lizard,
652 *Podarcis sicula sicula*? *Hormones and Behavior* **28**: 117–129.

653 Martín, J. and López, P. (2006a) Vitamin D supplementation increases the
654 attractiveness of males' scent for female Iberian rock lizards. *Proceedings of the*
655 *Royal Society B Biological Sciences* **273**: 2619–2624.

656 Martín, J. and López, P. (2006b) Age-related variation in lipophilic chemical
657 compounds from femoral gland secretions of male lizards *Psammodromus*
658 *algirus*. *Biochemical Systematics and Ecology* **34**: 691–697.

659 Martín, J. and López, P. 2014. Pheromones and other chemical communication in
660 animals. In: Rheubert, J. L., Siegel, D. S. and Trauth, S.E. (Eds): *Reproductive*
661 *Biology and Phylogeny of Lizards and Tuatara*, pp. 43–77. CRC Press, New
662 York.

663 Martín, J. and López, P. 2015. Condition-dependent chemosignals in reproductive
664 behavior of lizards. *Hormones and behavior* **68**: 14–24.

665 Martín J., Civantos, E., Amo, L. and López, P. (2007a) Chemical ornaments of male
666 lizards *Psammodromus algirus* may reveal their parasite load and health state to
667 females. *Behavioral Ecology and Sociobiology* **62**: 173–179.

668 Martín, J., López, P., Gabirot, M. and Pilz, K. M. 2007b. Effects of testosterone
669 supplementation on chemical signals of male Iberian wall lizards: consequences
670 for female mate choice. *Behavioral Ecology and Sociobiology* **61**: 1275–1282.

671 Martín J., Amo, L. and López, P. 2008. Parasites and health affect multiple sexual
672 signals in male common wall lizards, *Podarcis muralis*. *The Science of Nature*
673 **95**: 293–300.

674 Martín J., Ortega, J. and López P. 2015. Interpopulational variations in sexual
675 chemical signals of Iberian wall lizards may allow maximizing signal efficiency
676 under different climatic conditions. *Plos One* **10**: e0131492.

677 Martins, E. P., Ord, T. J., Slaven, J., Wright, J. L. and Housworth, E. A. 2006.
678 Individual, sexual, seasonal, and temporal variation in the amount of sagebrush
679 lizard scent marks. *Journal of Chemical Ecology* **32**: 881–893.

680 Mason, R. T. and Gutzke, W. H. N. 1990. Sex recognition in the leopard gecko,
681 *Eublepharis macularius* (Sauna: Gekkonidae): possible mediation by skin
682 derived semiochemicals. *Journal of Chemical Ecology* **16**: 27–36.

683 Mason, R. T. and Parker, M. R. 2010. Social behavior and pheromonal
684 communication in reptiles. *Journal of comparative physiology. A,*
685 *Neuroethology, sensory, neural, and behavioral physiology* **196**: 729–749.

686 Matthey, R. 1929. Caractères sexuels secondaires du lézard mâle. *Bulletin de la*
687 *Société Vaudoise des Sciences Naturelles* **57**: 71–81.

688 Mayerl, C., Baeckens, S. and Van Damme, R. 2015. Evolution and role of the
689 follicular epidermal gland system in non-ophidian squamates. *Amphibia-Reptilia*
690 **36**: 185–206.

691 Michaelides, S., Cornish, N., Griffiths, R., Groombridge, J., Zajac, N., Walters, G. J.,
692 Aubret, F., While, G. M. and Uller, T. 2015. Phylogeography and conservation
693 genetics of the common wall lizard, *Podarcis muralis*, on islands at its northern
694 range. *Plos One* **10**: e0117113.

695 Moore, M., Thompson, C. and Marler, C. 1991. Reciprocal changes in corticosterone
696 and testosterone levels following acute and chronic handling stress in the tree
697 lizard, *Urosaurus ornatus*. *General and Comparative Endocrinology* **81**: 217-
698 226.

699 Moreira, P. L., López, P. and Martín, J. 2008. Discrimination of conspecific faecal
700 chemicals and spatial decisions in juvenile Iberian rock lizards (*Lacerta*
701 *monticola*). *Acta Ethologica* **11**: 26–33.

702 Müller-Schwarze, D. 2006. Chemical Ecology of Vertebrates. Cambridge University
703 Press, Cambridge.

704 Olsson, M., Wapstra, E., Madsen, T. and Silverin, B. 2000. Testosterone, ticks and
705 travels: A test of the immunocompetence-handicap hypothesis in free-ranging
706 male sand lizards. *Proceedings of the Royal Society B Biological Sciences* **267**:
707 2339–2343.

708 Oppliger, A., Giorgi, M. S., Conelli, A., Nembrini, M. and John-Alder, H. B. 2004.
709 Effect of testosterone on immunocompetence, parasite load, and metabolism in
710 the common wall lizard (*Podarcis muralis*). *Canadian Journal of Zoology* **82**:
711 1713–1719.

712 Padoa, E. 1933. Ricerche sperimentali sui pori femorali e sull'epididimo della
713 lucertola (*Lacerta muralis* Laur.) considerati come caratteri sessuali secondari.
714 *Archivio Italiano di Anatomia e di Embriologia* **31**: 205–252.

715 Palme, R. and Möstl, E. 1994. Biotin-streptavidin enzyme immunoassay for the
716 determination of oestrogens and androgens in boar faeces. In: Görög, S. (Eds):
717 Advances of Steroid Analysis, pp. 111–117. Akadémiai Kiadó, Budapest.

718 Palme, R., Touma, C., Arias, N., Dominchin, M. F. and Lepschy, M. 2013. Steroid
719 extraction: get the best out of faecal samples. *Wiener Tierärztliche*
720 *Monatsschrift* **100**: 238–246.

721 Pratt, N. C., Alberts, A. C., Fulton-Medler, K. G. and Phillips, J. A. 1992. Behavioral,
722 physiological and morphological components of dominance and mate
723 attraction in male green iguanas. *Zoo Biology* **11**: 153–163.

724 Quay, W. B. 1986. The skin of reptiles: glands. In: Bereiter-Hahn, J., Matoltsy, A.G.
725 and Richards, K. S. (Eds): *Biology of the Integument, Vol. 2: Vertebrates*, pp.
726 188–193. Springer-Verlag, Berlin.

- 727 Regamey, J. 1932. Caractères sexuels secondaires du *Lacerta agilis* Linné. *Bulletin de*
728 *la Société Vaudoise des Sciences Naturelles* **57**: 589–591.
- 729 Rhen, T., Sakata, J. T. and Crews, D. 2005. Effects of gonadal sex and incubation
730 temperature on the ontogeny of gonadal steroid concentrations and secondary
731 sex structures in leopard geckos, *Eublepharis macularius*. *General and*
732 *Comparative Endocrinology* **142**: 289–296.
- 733 Schwenk, K. 1995. Of tongues and noses: chemoreception in lizards and snakes.
734 *Trends in Ecology & Evolution* **10**: 7–12.
- 735 Sinervo, B., Miles, D. B., Frankino, W. A., Klukowski, M. and DeNardo, D. F. 2000.
736 Testosterone, endurance, and Darwinian fitness: Natural and sexual selection
737 on the physiological bases of alternative male behaviors in side-blotched
738 lizards. *Hormones and Behavior* **38**: 222–233.
- 739 Smith, H. M. 1946. Handbook of Lizards: Lizards of the United States and of Canada.
740 Cornell University Press, New York.
- 741 Stamps, J. A. 1977. Social behavior and spacing patterns in lizards. In: Gans, C. and
742 Tinkle, D. W. (Eds): *Biology of the Reptilia*, Vol. 7: Ecology and Behaviour
743 A, pp. 265–334. Academic Press, New York.
- 744 Tokarz, R. R. 1985. Body size as a factor determining dominance in staged agonistic
745 encounters between male brown anoles (*Anolis sagrei*). *Animal Behaviour* **33**:
746 746–753.
- 747 Valdecantos, S., Martinez, V. and Labra, A. 2014. Comparative morphology of
748 *Liolaemus* lizards precloacal glands. *Acta Herpetologica* **9**: 147–158.
- 749 Van Wyk, J. H. 1990. Seasonal testicular activity and morphometric variation in the
750 femoral glands of the lizard *Cordylus polyzonus polyzonus* (Sauria:
751 Cordylidae). *Journal of Herpetology* **24**: 405–409.

752 Wyatt, T. D. 2014. Pheromones and Animal Behaviour: Chemical Signals and
753 Signatures. Cambridge University Press, Cambridge.

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782 **Figures**

783 **Fig. 1**— An illustration of the cotton-swab technique, designed to test lizard's tongue-flick
784 behaviour towards chemical stimuli. Drawn by Detlef Tibax.

785

786 **Fig. 2**— Mean (\pm SE) faecal testosterone (metabolite) concentrations of *Podarcis muralis*
787 males from the experimental group, which received a testosterone treatment (T-males) and
788 males from the control group (C-males). Means are shown 'before' and 'after' the three-week
789 administration period. Steroid concentration is presented as the mass of immunoreactive
790 testosterone metabolites per dried faecal mass. FTM = faecal testosterone metabolites.
791 Asterisks indicate statistical differences.

792

793 **Fig. 3**— Relationship between epidermal gland secretion mass and snout-vent length (SVL)
794 in *Podarcis muralis* males.

795

796 **Fig. 4**— Mean (\pm SE) difference in epidermal secretion mass (i.e. secretion mass after three-
797 week administration period subtracted by secretion mass before administration period) for
798 control males (C) and males with experimentally increased testosterone levels (T). Asterisks
799 indicate statistical differences.

800

801 **Fig. 5**— **(a)** Mean latency (in seconds) to the first TF and **(b)** mean number of tongue-flicks
802 (TF) elicited in 1 minute by female conspecifics in response to different stimuli (symbolized
803 by different background colours) originating from control males (C) and males with
804 experimentally increased testosterone levels (T). The stimuli are: deionized water (W); a
805 single portion of male epidermal gland secretion (red); a double portion of male epidermal
806 gland secretion (green); faeces (blue). Error bars represent SE. The same letters above the
807 bars denotes that means are not significantly different from each other.