
In-Source CID of Guanosine: Gas Phase Ion-Molecule Reactions

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In-source collision induced dissociation was applied to access second generation ions of protonated guanosine. The in-source gas-phase behavior of $[\text{BH}_2]^+\text{-NH}_3$ (m/z 135, $\text{C}_5\text{H}_3\text{N}_4\text{O}^+$) was investigated. Adduct formation and reactions with available solvent molecules (H_2O and CH_3OH) were demonstrated. Several addition/elimination sequences were observed for this particular ion and solvent molecules. Dissociation pathways for the newly formed ions were developed using a QqTOF mass spectrometer, permitting the assignment of elemental compositions of all product ions produced. Reaction schemes were suggested arising from the ring-opened intermediate of the protonated base moiety $[\text{BH}_2]^+$, obtained from fragmentation of guanosine. The mass spectral data revealed that the in-source CH_3OH -reaction product underwent more complex fragmentations than the comparable ion following reaction with H_2O . A rearrangement and a parallel radical dissociation pathway were discerned. Apart from the mass spectrometric evidence, the fragmentation schemes are supported by density functional theory calculations, in which the reaction of the ring-opened protonated guanine intermediate with CH_3OH and a number of subsequent fragmentations were elaborated. Additionally, an in-source transition from the ring-opened intermediate of protonated guanine to the ring-opened intermediate of protonated xanthine was suggested. For comparison, a low-energy collision induced dissociation study of xanthosine was performed. Its dissociation pathways agreed with our assumption. (J Am Soc Mass Spectrom 2006, 17, 1050–1062) © 2006 American Society for Mass Spectrometry

During the last decade, most fragmentation pathways were elaborated on triple quadrupole (QqQ) and particularly quadrupole ion-trap (QIT) instruments. The latter's capabilities of accessing MS^n product ions by applying repeated stages of mass selection and fragmentation proved to be very useful [1]. The maturation of the time of flight (TOF) technology and, more importantly, its use in hybrid-arrangements such as the popular orthogonal QqTOF mass spectrometer made accurate mass data of product ions more readily available [2]. In many cases, the elemental compositions of the product ions can unambiguously be

assigned when using a QqTOF mass spectrometer. Product ions lower in the genealogical ladder are also accessible by applying relatively high collision energies in the collision cell. Yet disentanglement of different dissociation pathways originating from a communal progenitor is—in general—still confined to QIT set ups, although the interpretation of energy resolved spectra also has proven its usefulness [3].

Recently we and others showed [4–6] that by making optimal use of in-source CID, combined with the inherent tandem MS capabilities of the QqTOF, what is in effect up to MS^5 data can be acquired. Compared to QIT data the QqTOF data obtained is superior in respect of higher mass accuracy. On the other hand QIT-CID experiments, in general, access almost exclusively the dissociation reaction channels of the lowest energy of activation [7]. This energy-dependent information can

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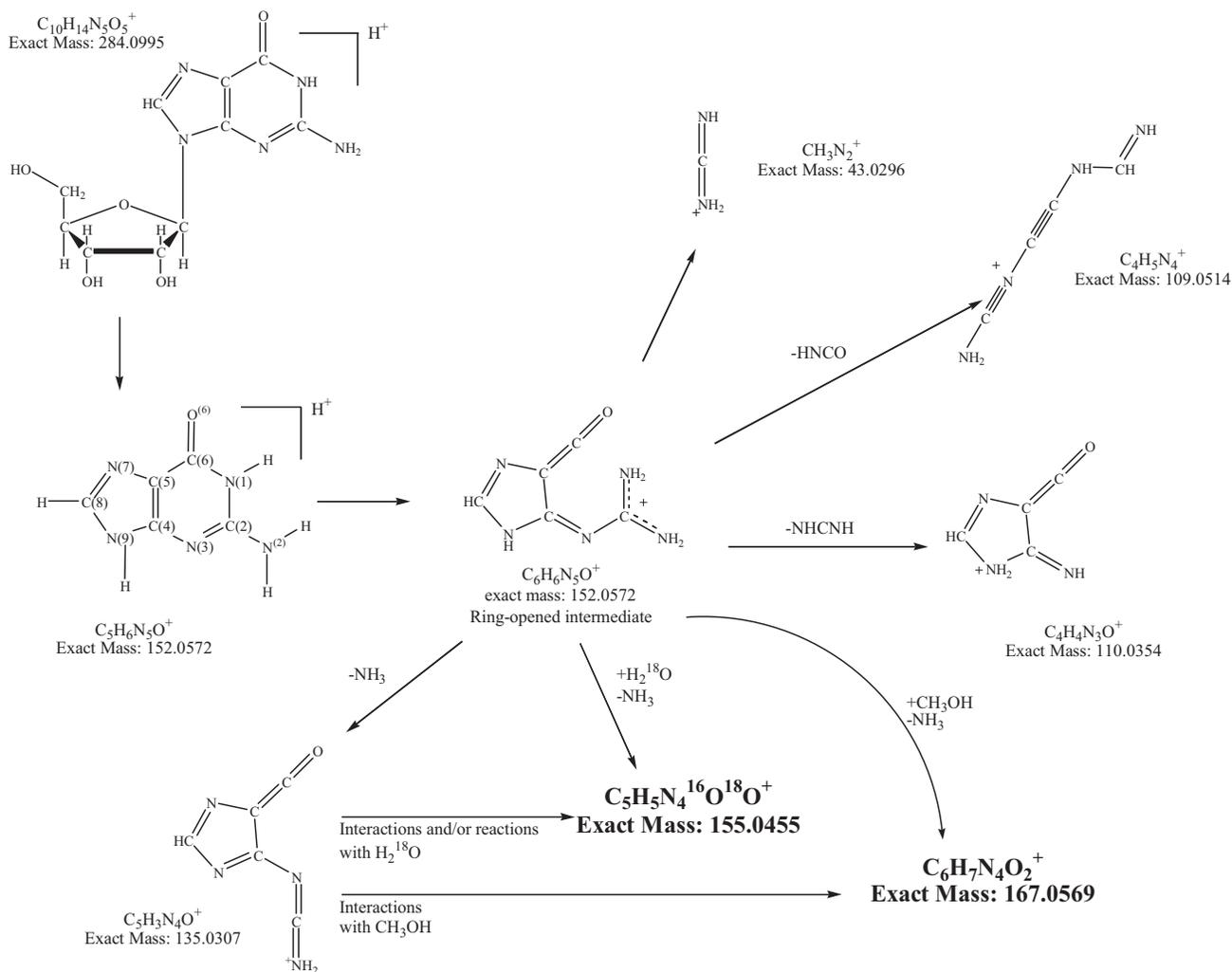


Figure 1. The main dissociation pathways of protonated guanosine up to the second generation product ions, together with the ions at m/z 155 and 167, respectively, formed upon interaction/reaction with $H_2^{18}O$ or CH_3OH . For this study the latter ions were generated in-source by applying adjusted CV-values.

pass unrecognized in QqTOF tandem MS experiments due to a less controlled energy deposit on the parent ions. However, in QqTOF experiments fragmentation pathways with higher activation energies become within reach due to higher energy-transfer in a single collision at high CE settings [6, 7].

During an in-depth study of the fragmentation behavior of the nucleoside guanosine with the optimized in-source CID-QqTOF MS approach it became clear that some of its product ions have a high affinity for H_2O . Additionally, some other product ions arose, which suggested even more sophisticated ion-molecule reactions [5]. It has to be noted that in-source addition of solvent molecules has been observed before by Gabelica et al. for benzylpyridinium cations [8].

This work focuses on these gas-phase ion-molecule interactions and reactions of one particular product ion of protonated guanosine. By in-source CID of the protonated guanosine the second generation product ion

$C_5H_3N_4O^+$ at m/z 135 (Figure 1), was produced [5, 9]. Its in-source formed adducts with $H_2^{18}O$ and CH_3OH were subsequently submitted to tandem MS analysis by QqTOF MS and accurate mass data of the according product ions was acquired. Different neutral gain/neutral loss sequences involving H_2O and CH_3OH were distinguished.

Based on the product ions, an in-source formation of ring-opened xanthine was predicted, emanating from the reaction with H_2O of the ring opened protonated guanine. To support this assumption the complete low-energy CID fragmentation study of xanthosine was elaborated.

The product ion data of the CH_3OH adduct indicated that a reaction of the same ring opened protonated guanine with CH_3OH occurred. Product ions were observed resulting from the transfer of the methyl group from the methanol to the heterocyclic base. The tentative reaction- and fragmentation schemes deduced

from the mass spectral data were confirmed by quantumchemical calculations in which theoretical information on the anticipated gas-phase reaction and the ensuing fragmentation pathways were obtained at the DFT level. To this end, equilibrium geometries were obtained starting from the tentative product ion structures, and the reaction energies ΔE_r between the reagents/precursor ions and products/fragments were determined. In addition, for all reactions studied, the transition states were localized and the corresponding activation energies were calculated. The results illustrated that for all reactions studied, the theoretical data compared favorably with the tentative schemes. In addition, the DFT calculations provided additional insights with respect to proton positions and the proton transfers.

Experimental

Chemicals

Guanosine, xanthosine, and N7-methylguanosine were purchased from Sigma (Bornem, Belgium). ^{18}O -labeled water, H_2^{18}O (^{18}O , 95%) was obtained from Cambridge Isotope Laboratories, Inc. (Andover, MA). Methanol, CH_3OH , (HPLC grade), and phosphoric acid, H_3PO_4 , were acquired from Acros Organics (Geel, Belgium). H_2O [when the isotope label for oxygen is not mentioned, it is adopted to be ^{16}O] (HPLC grade) was purchased from Fisher Scientific (Loughborough, Leicestershire, UK). Formic acid, HCOOH , (99–100%; ultrapure) was obtained from VWR International (Leuven, Belgium).

1 mL stock solutions of 10^{-3}M of each nucleoside were prepared in 50/50 (vol/vol) $\text{CH}_3\text{OH}/\text{H}_2\text{O}$ or $\text{CH}_3\text{OH}/\text{H}_2^{18}\text{O}$ (only guanosine). Working solutions were obtained by diluting the stocks further to 10^{-5}M in 50/49.9/0.1 (vol/vol) $\text{CH}_3\text{OH}/\text{H}_2\text{O}/\text{HCOOH}$ or 50/49.9/0.1 (vol/vol) $\text{CH}_3\text{OH}/\text{H}_2^{18}\text{O}/\text{HCOOH}$. For this purpose, a 10% formic acid solution in H_2O or H_2^{18}O was used.

Instrumentation and MS Conditions

A Q-TOF II mass spectrometer (Waters, Manchester, UK) was equipped with a standard pneumatically-assisted electrospray probe and a Z-spray source (Waters). Electrospray mass spectra were recorded in the positive ion mode. The applied experimental settings were: spray voltage 3.5 kV, source temperature 80 °C, and desolvation temperature 100 °C. A cone gas flow-rate of ca. 60 L/h and a desolvation gas flow-rate of ca. 150 L/h were applied. In MS/MS experiments, the precursor ions were selected with an isolation window of ± 0.5 Da, adequately removing isotopic (^{13}C) interferences from the spectrum. Scan time and inter-scan time were, respectively 1.0 s and 0.1 s. For each MS/MS experiment the nucleoside solution was infused for 50 min at a flow rate of 5 $\mu\text{L}/\text{min}$ via a syringe pump

Table 1. Applied experimental conditions for the different ionic species subjected to MS/MS

		<i>m/z</i>	<i>CV</i> (V)	<i>CE-range</i> (eV)
Guo	$[\text{BH}_2]^+-\text{NH}_3$	135	65	5–40
	$[\text{BH}_2]^+-\text{NH}_3+\text{H}_2^{18}\text{O}$	155	65	5–40
	$[\text{BH}_2]^+-\text{NH}_3+\text{CH}_3\text{OH}$	167	65	5–50
Xan	$[\text{M}+\text{H}]^+$	285	20	5–60
	$[\text{BH}_2]^+$	153	30	5–50
	$[\text{BH}_2]^+-\text{NH}_3$	136	50	5–40
	$[\text{BH}_2]^+-\text{HNCO}$	110	65	5–40

(Harvard Apparatus, Natick, MA). During the MS/MS experiment the collision energy was varied in a cyclic manner (5°eV° intervals, 9° ranges cf. $^\circ$ Table 1) permitting the construction of collision energy profiles. Before the MS/MS experiments of the in-source formed product ions the cone voltage was optimized for each ion of interest. Therefore, a nucleoside solution was infused at 5 $\mu\text{L}/\text{min}$ and full scan spectra were recorded (*m/z* 100–600) with a cone voltage varying between 15 and 90 V (5 V intervals, each 30 s). The optimal cone voltage (CV) for each ion of interest was selected upon the evaluation of the extracted ion chromatograms and the corresponding spectra (cf. isobaric interferences).

For high mass accuracy the QqTOF MS was calibrated using 0.1% phosphoric acid in 50:50 (vol/vol) $\text{MeOH}/\text{H}_2\text{O}$. Any instrument drift was compensated by applying a lock mass correction. Depending on the experiment the protonated nucleoside $[\text{M} + \text{H}]^+$, the protonated base $[\text{BH}_2]^+$, or other product ions were used as lock mass. In the latter case only these fragments of which the elemental composition was confirmed by us and others were utilized. The TOF analyzer was programmed to record the *m/z* range 50–400 for the protonated nucleosides and the *m/z* range 50–250 for $[\text{BH}_2]^+$ and the other ionic species studied. Only nominal masses are given in the discussions, accurate mass data is available in the Supplementary Information, (which can be found in the electronic version of this article).

The data were processed using MassLynx version 3.5. The following criteria were applied to distinguish “true” product ions from interferences in the spectra: (1) only ions with intensity above 1% of the base peaks intensity were included; (2) the product ions must be present in the spectra for at least two collision energy values applied to any given precursor ion; (3) the product ions must yield credible empirical formulas in respect of the parent ion from which they arose (taking into account possible adduct formation in the collision cell as described previously) [5, 10].

Calculations

Density functional theory (DFT) calculations were performed using Gaussian 03 [11], as installed on the computing cluster CalcUA. Equilibrium geometries and transition states were obtained at the B3LYP/6-31G(d)

level. Additional evidence supporting the nature of the stationary was obtained by calculating the corresponding Hessian, and by carefully analyzing the number of imaginary vibrational frequencies derived from them. The Cartesian coordinates, charge distributions, and vibrational frequencies for the geometries obtained are available from the authors upon request.

It should be noted that for the reagent/parent ions and the product ions, computational evidence of donor-acceptor interactions between the most electrophilic region of the ions with an electron-rich region of the incoming (CH_3OH) or leaving (H_2O , NH_3 , HOCN , or CH_3NH_2) species was found. The appearance of these intermolecular interactions, evidently, influences the calculated energies. In this study, the activation energies and the reaction energies were calculated by comparing the energies of the initial and final adducts, without correcting for the complexation energies present.

Results and Discussion

H₂O- and CH₃OH-Adduct Formation of In-Source CID Product ions

As a starting point of this study a cone voltage (CV) optimization was performed: the CV was gradually increased while infusing an acidified guanosine solution into the ESI-source. Increasing the CV resulted firstly in a well known break up of the anomeric bond, leading to $[\text{BH}_2]^+$. Further CV increase gave rise to the in-source formation of the initial product ions of the known $[\text{BH}_2]^+-\text{NH}_3$ (m/z 135; $\text{C}_5\text{H}_3\text{N}_4\text{O}^+$) and $[\text{BH}_2]^+-\text{NHCNH}$ pathways (m/z 110; $\text{C}_4\text{H}_4\text{N}_3\text{O}^+$) and the minor $[\text{BH}_2]^+-\text{HNCO}$ (m/z 109; $\text{C}_4\text{H}_5\text{N}_4^+$) pathway (Figure 1). Earlier experiments, augmented with DFT calculations, showed that both the m/z 110 and m/z 135 product ions interacted strongly with H_2O present in the source and with traces of H_2O present in the collision cell [5]. In the latter study, the occurrence of a non-covalent (hydrogen bonded) adduct m/z 128 ($\text{C}_4\text{H}_4\text{N}_3\text{O}^+\cdot\text{H}_2\text{O}$), was derived from the MS/MS-spectra and supported by the DFT calculations. However further characterization of the m/z 153 due to $\text{C}_5\text{H}_3\text{N}_4\text{O}^+\cdot\text{H}_2\text{O}$ with MS/MS revealed several new product ions in addition to the known fragments of m/z 135 [5].

Inspection of the full scan spectra clearly illustrated that adduct formation with methanol also occurred for the product ions m/z 110 and m/z 135. In agreement with earlier observations suggesting that CH_3OH is a much stronger Lewis base than H_2O , the formation of molecular complexes with CH_3OH is much more pronounced than with H_2O (Figure 2). Thus a significant fraction of these, by in-source CID produced, second generation product ions of guanosine was promptly transformed to ions with higher m/z ratios. The correlation between the in-source formed product ion at m/z 135 and its H_2^{18}O - and CH_3OH -adducts (respectively, at m/z 155 and 167) is clear from Figure 2 with increasing CV

parallel appearances of the m/z ratios of $[\text{BH}_2]^+-\text{NH}_3$, $[\text{BH}_2]^+-\text{NH}_3 + \text{H}_2^{18}\text{O}$, $[\text{BH}_2]^+-\text{NH}_3 + \text{CH}_3\text{OH}$ occur. Moreover, also in the product ion spectra of the in-source formed ion at m/z 167 ($\text{C}_5\text{H}_3\text{N}_4\text{O}^+ + \text{CH}_3\text{OH}$), product ions are seen which cannot be explained solely by a noncovalent ion-neutral interaction: gas-phase reactions have to be assumed. In contrast the product ion spectra of m/z 142, ($\text{C}_4\text{H}_4\text{N}_3\text{O}^+ + \text{CH}_3\text{OH}$), shows no other product ions than these corresponding to the initial ions from m/z 110.

C₅H₅N₄¹⁶O¹⁸⁺: A Result of H₂¹⁸O-Adduct Formation and H₂¹⁸O-Gas-Phase Reactions

To investigate the phenomena arising from the solvent-adducts of m/z 135 ($\text{C}_5\text{H}_3\text{N}_4\text{O}^+$) it is favorable to replace H_2^{16}O by H_2^{18}O in the sample, since the H_2^{16}O -adduct at m/z 153 nominally coincides with the natural ¹³C-isotopic signal of the $[\text{BH}_2]^+$ of guanosine. However, it has to be recognized that considerable amounts of H_2^{16}O are still present during in-source CID in agreement with the quoted commercial purity of the H_2^{18}O (¹⁸O, 95%) and the atmospheric conditions in the ESI-source.

Application of a cone voltage of 65 V generated in-source the ion at m/z 135 together with its H_2^{18}O -adduct at m/z 155 ($\text{C}_5\text{H}_5\text{N}_4^{16}\text{O}^{18+}$). Both the ions at m/z 135 and 155 were monoisotopically selected by the quadrupole filter and accurate mass energy-resolved spectra were collected in the range of 5–40 eV. Exemplary product ion spectra of both ions at CE 15 eV and CE 30 eV are shown in Figure 3. The accurate mass data confirmed that m/z 155 ($\text{C}_5\text{H}_5\text{N}_4^{16}\text{O}^{18+}$) corresponded with the addition of H_2^{18}O to m/z 135 ($\text{C}_5\text{H}_3\text{N}_4^{16}\text{O}^+$). The prominent presence of the m/z 135 ($\text{C}_5\text{H}_3\text{N}_4^{16}\text{O}^+$) in the spectra of m/z 155 at low CE agrees with the non-covalent nature of the interaction between $\text{C}_5\text{H}_3\text{N}_4^{16}\text{O}^+$ and H_2^{18}O . At the same time these spectra demonstrated the stability of the H_2^{18}O -adduct: for m/z 155 at CE 15 eV little product ions, apart from m/z 135, were produced, whereas a fairly extensive fragmentation was apparent at CE 15 eV in the product ion spectrum of m/z 135 itself. The typical product ions of m/z 135 only appeared in the product ion spectra of m/z 155 at higher CEs. The product ions related with the $[\text{BH}_2]^+-\text{NH}_3$ pathway have already been discussed [5, 9], but a variety of new product ions were also observed in the MS/MS spectra of m/z 155 in the current study. All the product ions associated with the dissociation of m/z 155 can be found in the Supplementary Materials section, which can be found in the online version of this article.

The product ions of m/z 155 at m/z 138 and 137 could not be explained by a simple ion-molecule cluster. The unexpected yet abundant ion at m/z 137 ($\text{C}_5\text{H}_3\text{N}_4^{18}\text{O}^+$) is found upon loss of H_2^{16}O from m/z 155. The elemental composition of m/z 137 requires that the ¹⁶O originally present in the carbonyl group (C6) of the guanine is

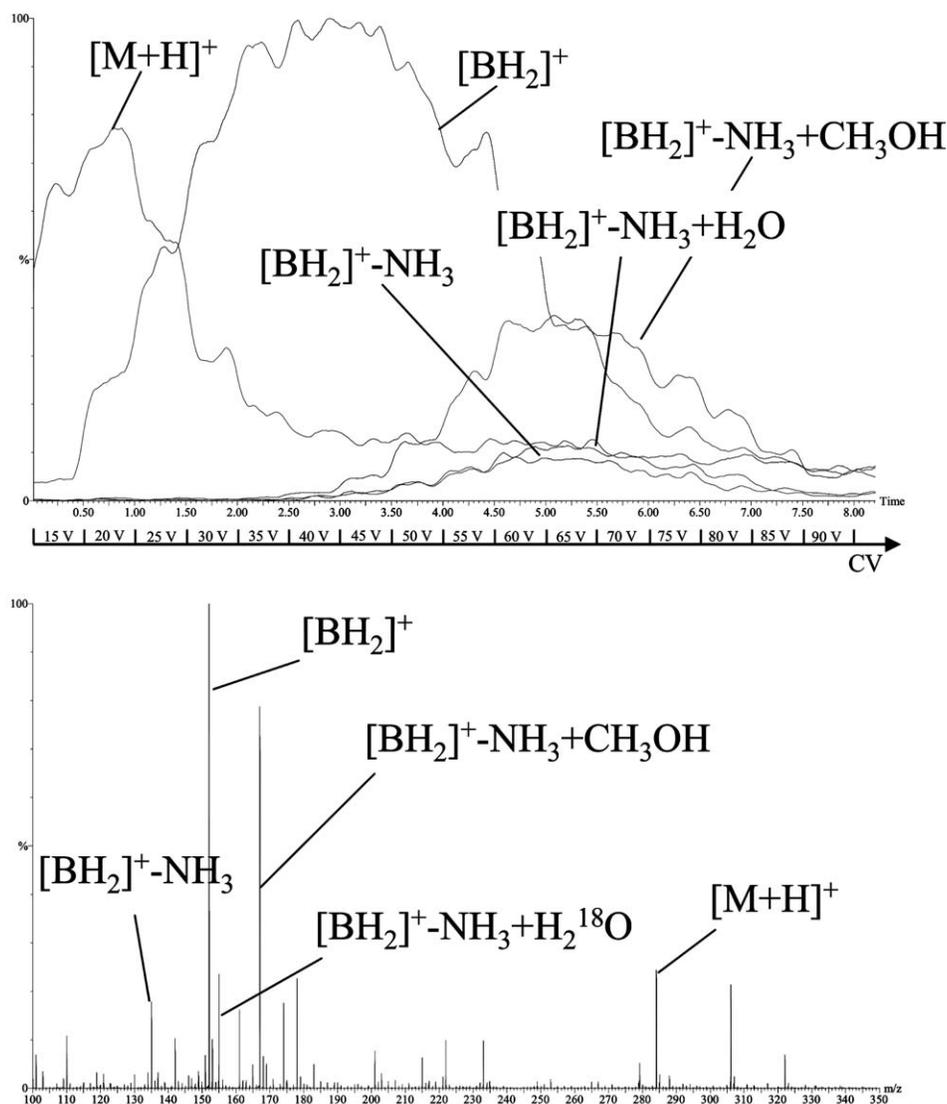


Figure 2. Top: The extracted ion profiles corresponding to the $[M + H]^+$, $[BH_2]^+$, the m/z 135 ($[BH_2]^+ - NH_3$) and the latter's $H_2^{18}O$ and CH_3OH -adducted ions in function of increasing cone voltage (CV; CV increased 5 V every 30 s). Bottom: The accumulated spectrum of the 4.5–5.5 min time span.

removed by formation of the carbonyl hydrate ($H_2^{18}O$, see Scheme 1, and 2), of m/z 135 in-source (m/z 155) and dehydration ($H_2^{16}O$) in the collision cell. Further support for this $^{16}O/^{18}O$ -exchange at C6 was found in the ion at m/z 95 $C_4HN_2^{18}O^+$. This ion corresponded with the product ion m/z 93 ($C_4HN_2^{16}O^+$), a well characterized fragment of m/z 135^[9], which conserves the initial oxygen of m/z 135 (^{18}O : m/z 137 and m/z 95). All the other product ions of m/z 137 coincided with the product ions in the original $[BH_2]^+ - NH_3$ (m/z 135) pathway since none of them retains the oxygen atom.

The accurate mass data for m/z 138 pointed to the elemental composition $C_5H_2N_3^{16}O^{18}O^+$, which agreed with a loss of NH_3 . Its interrelationship with m/z 155 requires a covalently bounded ^{18}O for $C_5H_5N_4^{16}O^{18+}$, and not solely a noncovalent ion-molecule interaction. Most likely an addition/elimination reaction occurs at C(2) of the open-ring intermediate of protonated gua-

anine structure proposed by Gregson and McCloskey^[9]. A nucleophilic attack by $H_2^{18}O$ onto the electron deficient C(2) can result in a subsequent elimination of NH_3 , leading to the ion at m/z 155 with the observed elemental composition. A second loss of NH_3 from the selected ion m/z 155 will result in the product ion at m/z 138.

Alternatively, a loss of $HNCO$ was easily envisioned from the newly formed ion at m/z 155: product ions at m/z 110 and 112, depending on the position of the ^{18}O -label, were anticipated. Indeed, an ion ($C_4H_4N_3O^+$) was detected as a low abundant ion at m/z 112 (^{18}O). The ^{16}O analogue at m/z 110 was not seen since it was obscured by a new abundant ion as explained below. The structure resulting from the loss of $HNCO$ starting from m/z 155 was thought to be the same as that emanating from $[BH_2]^+ - NHCNH$ in the fragmentation of guanine^[9]. This was confirmed by the presence of

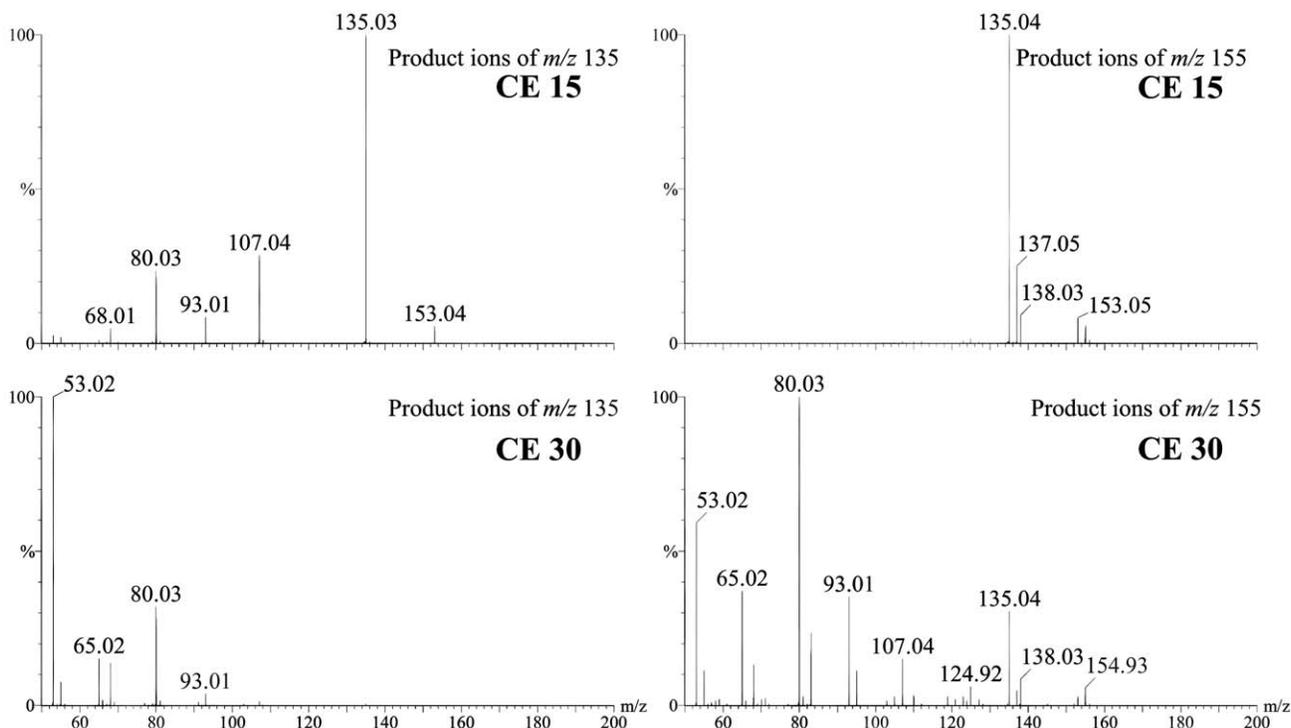


Figure 3. Left: Product ion spectra of the in-source generated ion at m/z 135 ($C_5H_4N_3O^+$) outgoing protonated guanosine (CV 65 V and CE 15 eV (upper) and 30 eV (lower); no lock mass correction, uncentered). Right: Product ion spectra of the in-source generated ion at m/z 155 ($C_5H_4N_3O^+ + H_2^{18}O$) outgoing protonated guanosine (CV 65 V and CE 15 eV (upper) and 30 eV (lower); no lock mass correction, uncentered).

product ions known from the $[BH_2]^+-NHCNH$ fragmentation pathway resulting from two consecutive losses of HCN, at m/z 85 $C_3H_3N_2^{18}O^+$ and m/z 58 $C_2H_2N^{18}O^+$. The corresponding ^{16}O ions were not detected either due to overlapping signals or low signal intensity. Loss of carbon monoxide ($C^{18/16}O$) from m/z 112/110 results in the known ion at m/z 82 which further loses HCN.

Clearly by in-source addition of H_2O ($H_2^{18}O$) to the $[BH_2]^+-NH_3$ ion and the subsequent loss of HNCO from m/z 153 (m/z 155), a blending in of the two major known fragmentation pathways ($[BH_2]^+-NH_3$ (m/z 135) and the $[BH_2]^+-HNCNH$ (m/z 110)) occurs, that are normally separate for the fragmentation of protonated Gua [5,9].

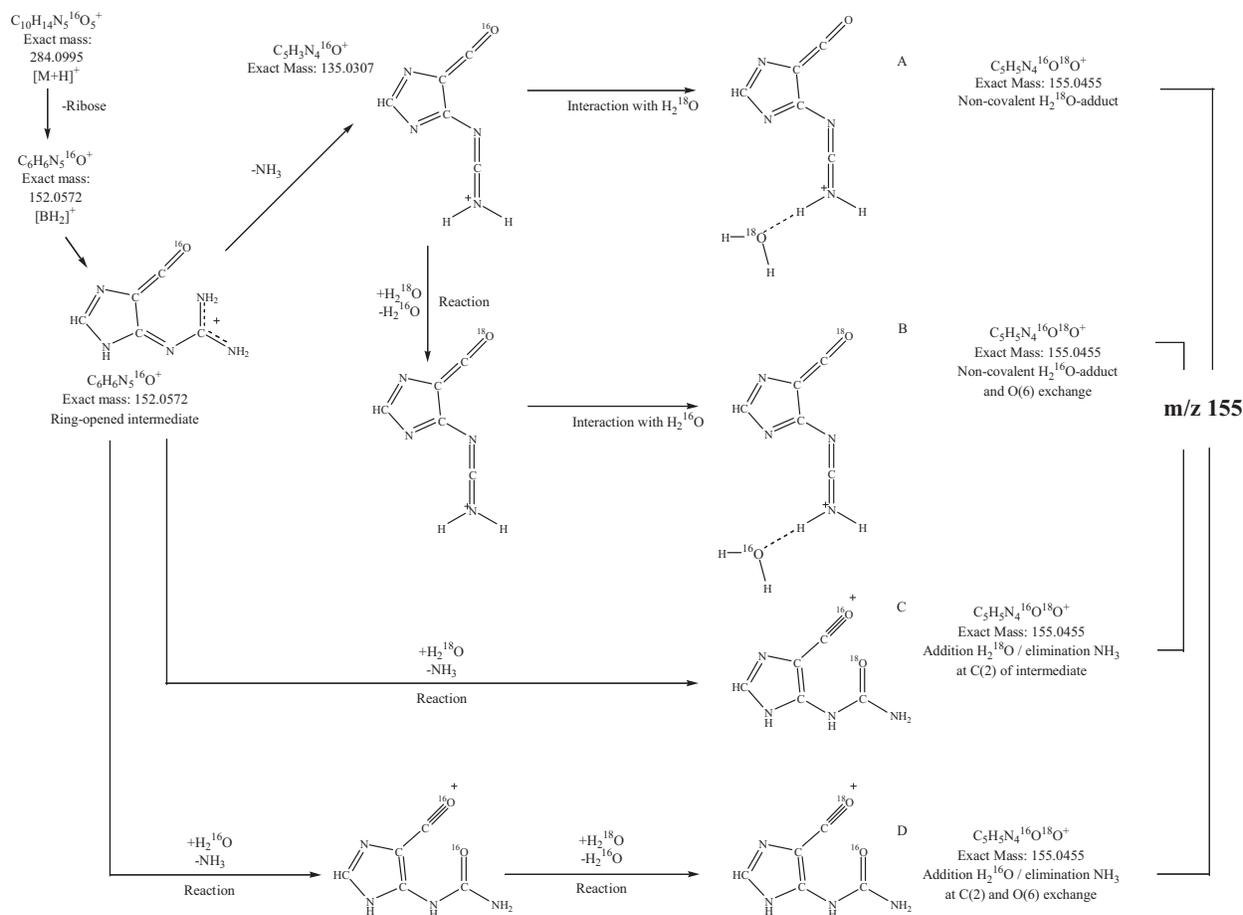
In Scheme 2 the other product ions arising from m/z 138 are given. The expulsion of $H^{16/18}OCN$ leads to the pair of product ions at m/z 93/95, with the same elemental compositions as the product ions derived from the ions at m/z 135 and m/z 137. An additional loss of carbon monoxide ($C^{16/18}O$) from the ions at m/z 93/95 also produces an ion at m/z 65 ($C_3HN_2^+$) which again was common to a product ion appearing in the $[BH_2]^+-NH_3$ pathway. Evidence to support these convergent pathways will follow further. At the same time a second dissociation pathway developed from the ion at m/z 138 ($C_5H_2N_3^{16}O^{18}O^+$). An initial loss of carbon monoxide ($^{18/16}O$) led to the ions at m/z 110 and 108 (very low abundance). From these ions a subsequent

loss of HCN was also observed in the energy resolved spectra giving rise to the unique ions at m/z 83 and m/z 81.

$C_6H_7N_4O_2^+$: A Result of CH_3OH -Adduct Formation and CH_3OH -Gas-Phase Reactions

As mentioned previously and indicated in Figure 2 a significant fraction of the in-source generated ion at m/z 135, ensuing in-source CID of protonated guanosine, was converted into its CH_3OH -adduct (m/z 167). It was found interesting to verify if neutral gain/neutral loss sequences similar to those observed for the H_2O -adduct would occur as well for this CH_3OH -adduct. Infusing a Guo solution under the application of a CV of 65V produced high levels of the ion at m/z 167 in-source of which energy resolved spectra were recorded. As there was no isobaric interference there was no need to use isotopic labeled CH_3OH . In Figure 4 the accumulated product ion spectra of m/z 167 at CE 15 eV and 30 eV are shown; additionally the accurate mass data and elemental compositions of all product ions are available in the Supplementary Information. The accurate mass measurement of m/z 167 ($C_6H_7N_4O_2^+$) confirmed the addition of CH_3OH to $C_5H_3N_4O^+$.

Noncovalent interaction of methanol with the in-source formed fragment at m/z 135 was indicated by the



high abundance of the latter in the product ion spectra of m/z 167 at low CE. With increased CE, the typical product ions of the $[\text{BH}_2]^+ \text{-NH}_3$ dissociation pathway also appeared. The high affinity of m/z 135 for H_2^{16}O is seen in the product ion observed at m/z 153, the result of scavenging traces of H_2^{16}O available in the collision cell by the product ion at m/z 135 as reported earlier [5].

In analogy with the elimination of NH_3 and HNCO from the H_2^{18}O -adduct m/z 135, the methanol adduct m/z 167 showed the loss of NH_2CH_3 (m/z 136) and CH_3OCN (m/z 110), and corresponding product ions derived thereof (Scheme 3). The presence of these ions suggested that also for the adducts with methanol, an addition (CH_3OH)/elimination (NH_3) reaction departing from the ring-opened protonated guanine occurred similar to that observed for H_2^{18}O . Theoretical data obtained from DFT calculations, in which the reagents and the reactions products are studied, confirmed this reaction pathway. The equilibrium geometry of the ring opened intermediate of protonated guanine (m/z 152) of the product ion (m/z 167) and of typical product ions resulting from further fragmentation of m/z 167 are summarized in Scheme 4. The transition states involved and the corresponding values for E_a and ΔE_r are collected in Table 2.

The geometry optimizations of protonated ring-opened guanine suggested that the positive charge (proton) is preferentially located at the cyanamide moiety rather than at the carbonyl at the (6) position [9]. The DFT results revealed that the addition/elimination reactions are preceded by the formation of a donor-acceptor complex in which an incoming methanol molecule interacts with the ring opened guanine. Subsequently, a concerted reaction takes place: covalent bonding of the oxygen of CH_3OH at C(2) proton transfer from $\text{CH}_3\text{O}^*\text{H}$ to N(1)/(2)N and formation of NH_3 proceeds in a single step. The transition-state for the concerted reaction, TS1° is shown in Table 2. The corresponding values for ΔE_r and E_a are -1.7 and 223.6 kJ mol^{-1} . The activation (E_a) and reaction energies (ΔE_r) (Table 2) are similar to those obtained for proton transfers [12] and collision induced dissociations [13] in the product ions formation of other nucleosides. The experimental settings, i.e., the collision energy, allow for the calculated transitions.

As the methanol-product was formed with a higher abundance, the product ions m/z 136 and 110 as well as their fragmentation products were rather easily identified. The loss of methylamine leading to m/z 136 requires a rearrangement of the ion, in which the methyl

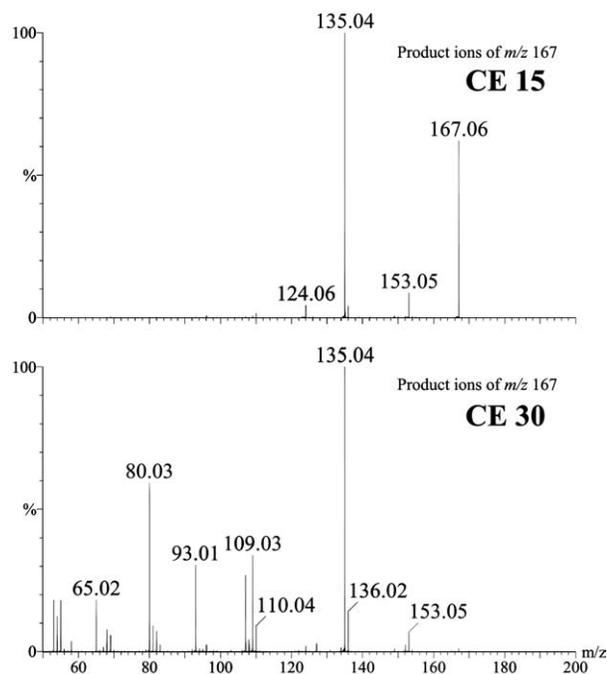
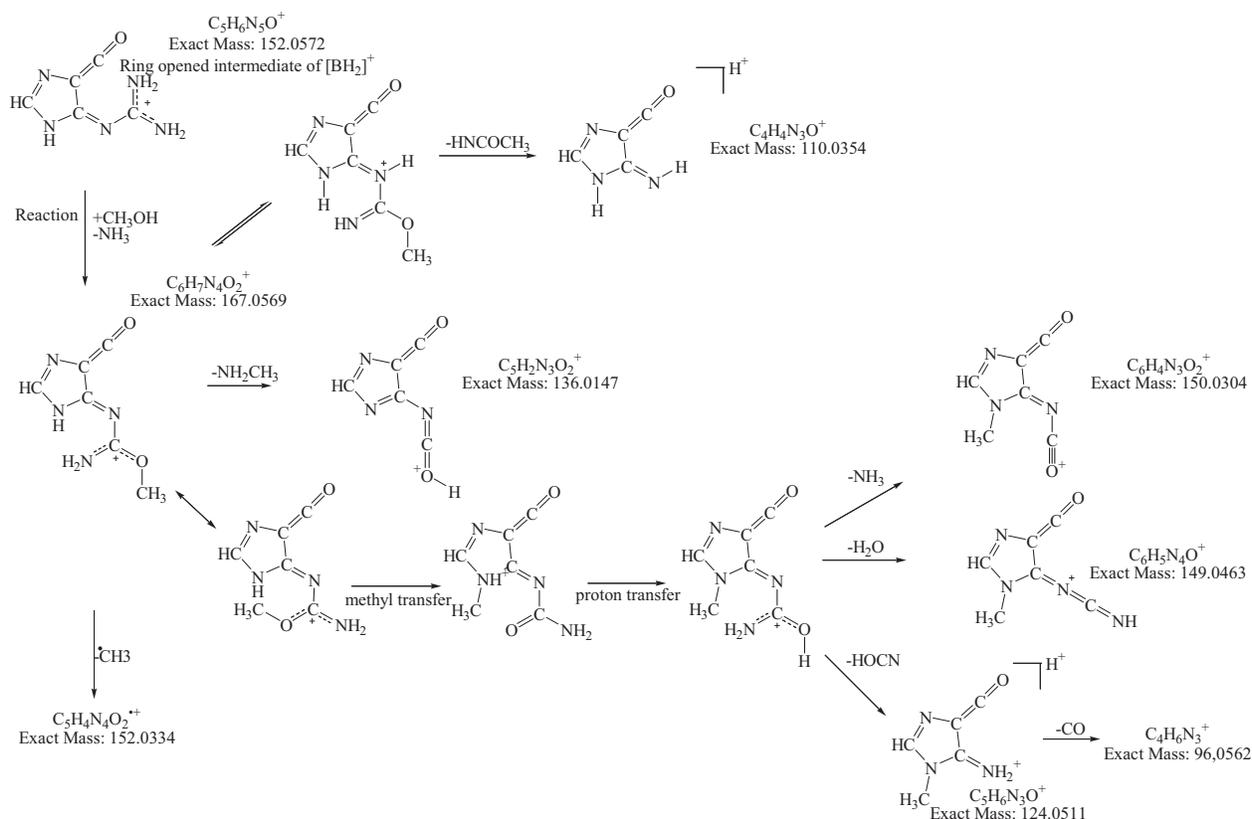


Figure 4. Product ion spectra of the in-source generated ion at m/z 167 ($C_5H_4N_3O^+ + CH_3OH$) outgoing protonated guanosine (CV 65 V and CE 15 eV (upper) and 30 eV (lower); no lock mass correction, uncentered).

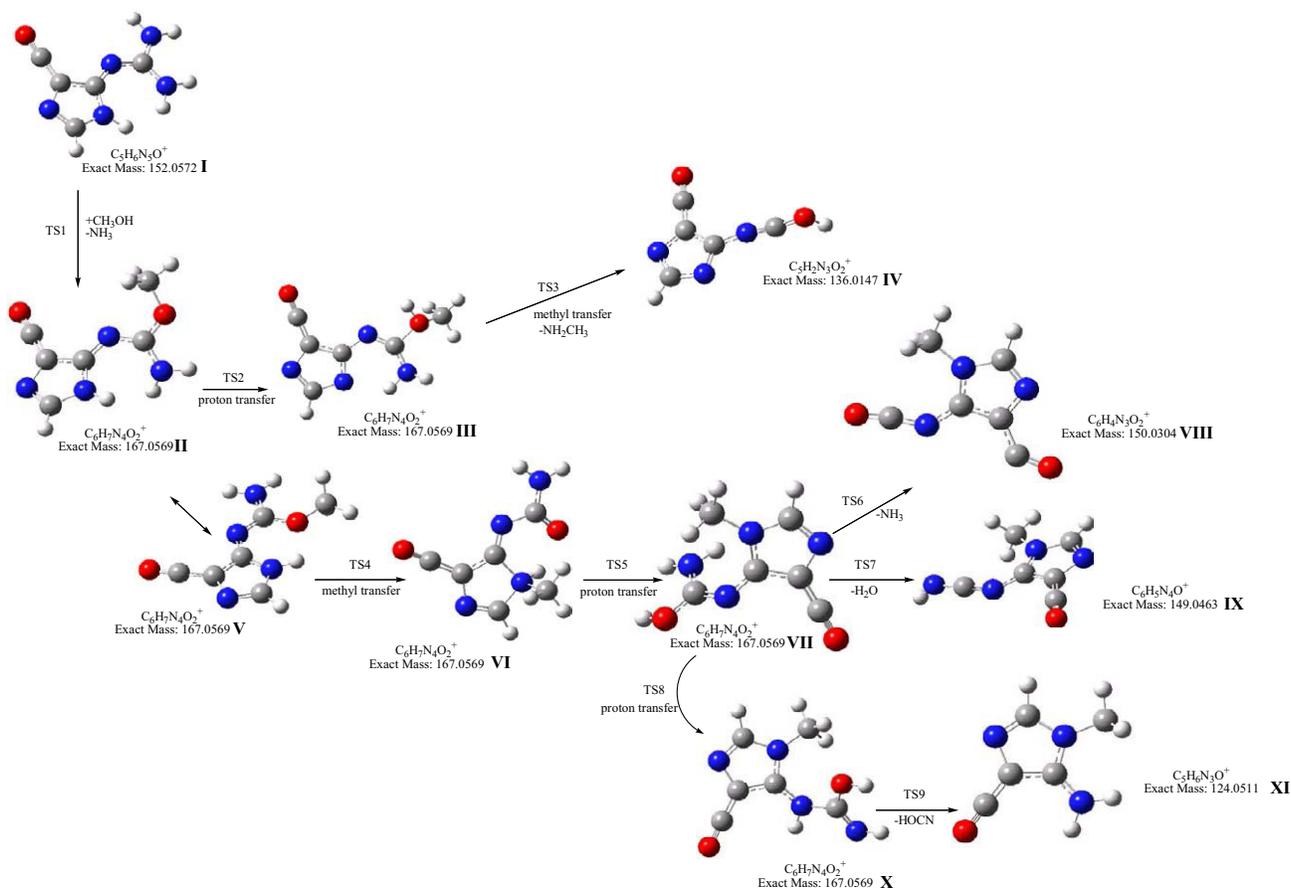
group is transferred from the methanol oxygen to the nitrogen N(1)/(2)N (cf) randomization of N(2)/(2)N subsequent ring-opening⁹ (Scheme 3). The occurrence of this transfer is confirmed by the DFT calculations (Scheme 4 and Table 2).

Apart from the features described above, new, less abundant product ions appeared at m/z 154 $C_5H_4N_3O_3^+$, m/z 152 $C_5H_4N_4O_2^+$, m/z 150 $C_6H_5N_3O_2^+$, m/z 149 $C_6H_5N_4O^+$ and m/z 124 $C_5H_6N_3O^+$. In agreement with the results reported earlier⁵, the ion m/z 154 $C_5H_4N_3O_3^+$ resulted from the addition of H_2O in the collision cell to m/z 136. The ions m/z 149 and 150 were assigned to species produced after elimination of H_2O , NH_3 , and $HOCN$ from the m/z 167 ion.

The latter experimental data are easily rationalized by the obtained equilibrium geometries and transition states derived from the DFT calculations. The elimination of H_2O , NH_3 , and $HOCN$ develops via a unique intermediate (VI), which emanates from V by a methyl transfer from the methanol oxygen to the N(9) position of the original guanine (TS4), followed by a proton transfer (TS5) from VI to VII. Literature and our own (unpublished) mass spectral data further supported the presence of the methyl group on the imidazole ring: product ions at m/z 149 and 124 showed to be distinctive for a methyl group positioned on a nitrogen of the imidazole part of a purine (e.g., N7- and N9-methyl-guanine)¹⁴. The further eliminations of H_2O and NH_3



Scheme 3

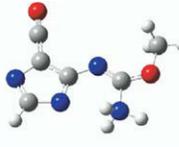


Scheme 4

from **VII** proceed via a single step, involving either TS7 or TS6. In contrast, the elimination of HOCN requires for two different subsequent proton transfers: a first transfer from **VII** to **X** through TS8 and a second transfer from **X** to **XI** through TS9 resulting finally in the expulsion of HOCN. An additional product ion at m/z 96 ($C_4H_6N_3^+$) corresponded an extra loss of CO from the ion at m/z 124.

Interestingly there also appeared a parallel dissociation pathway comprising only radical product ions in the energy resolved spectra of m/z 167. This series of radical cations starts off with the loss of a methyl radical from the ion at m/z 167 to m/z 152 $C_5H_4N_4O_2^+$. Two different pathways evolved from there, starting with the loss of H_2O yielding the ion at m/z 134 ($C_5H_2N_4O^+$) or the loss of HNCN leading to the ion at m/z 109 ($C_4H_3N_3O^+$), with the latter being the most abundant. From these two product ions multiple losses of HCN and/or CO showed, as commonly seen in the dissociation pathways of nucleosides [5,6,9,15,16]. It has to be noted that once more an addition of H_2O in the collision cell was observed; the radical ion at m/z 109 formed an adduct with H_2O yielding the radical ion at m/z 127 ($C_4H_5N_3O_2^+$). Many of these radical product ions have nominal masses that coincide with ions of other dissociation pathways (e.g., $C_4H_3N_4^+$ calc. m/z 107.0358,

Table 2. The optimized B3LYP/6-31G(d) transition states of the fragmentation of m/z 167 as presented in Scheme 4. Energies are given in kJ mol^{-1}

		
TS 1 E_a : 223.61 ΔE_R : -1.7	TS 2 E_a : 118.3 ΔE_R : 95.4	TS 3 E_a : 125.58 ΔE_R : -132.6
		
TS 4 E_a : 277.6 ΔE_R : 111.7	TS 5 E_a : 8.1 ΔE_R : -81.2	TS 6 E_a : 177.0 ΔE_R : -49.9
		
TS 7 E_a : 240.7 ΔE_R : 101.3	TS 8 E_a : 184.4 ΔE_R : 50.46	TS 9 E_a : 225.84 ΔE_R : 13.7

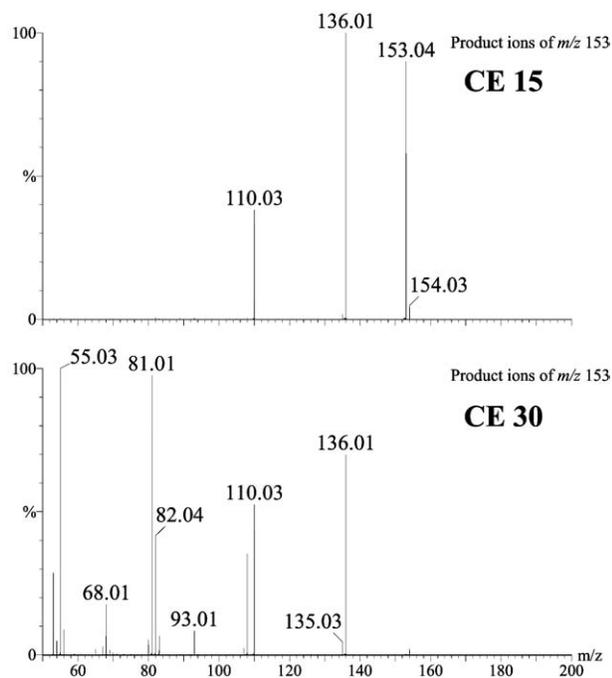


Figure 5. Product ion spectra of the in-source generated $[\text{BH}_2]^+$ ion of xanthosine at m/z 153 ($\text{C}_5\text{H}_5\text{N}_4\text{O}_2^+$) (CV 20 V and CE 15 eV (upper) and 30 eV (lower); no lock mass correction, uncentered).

meas. m/z 107.0362 and $\text{C}_4\text{HN}_3\text{O}^+$ calc. m/z 107.0120, meas. m/z 107.0127). They could only be discerned because of the sufficient resolving power and accuracy of the QqTOF MS.

In-Source Generation of the Ring-Opened Protonated Xanthine

The hydrolytic deamination of guanine to xanthine is well documented: an enzyme guanine deaminase catalyses the addition of H_2O and elimination of NH_3 .^[17] There is a clear similarity with the above described phenomena in the product ion spectra of guanosine. To probe this similarity, an in-depth CID study of the nucleoside xanthosine was performed by means of combined in-source and in-collision cell CID by using the QqTOF mass spectrometer. To the best of our knowledge, no extensive ES-MS fragmentation study of xanthosine has been carried out before.

Initially the energy resolved spectra of protonated xanthosine $[\text{M} + \text{H}]^+$ were collected between CE 5–60 eV and based on the accurate mass data elemental compositions assigned (cf. Supplementary Data). Akin to the majority of nucleosides the initial energy deposit caused a break of the anomeric bond. Though unlike other nucleosides an important part of the positive charge was channelled into the sugar-moiety leading to the product ion S^+ at m/z 133 ($\text{C}_5\text{H}_9\text{O}_4^+$). Further dissociation of this sugar-moiety led to a variety of ions. A plethora of losses of different small neutrals (i.e., water, carbon monoxide, formaldehyde, acetaldehyde) was displayed.

More important to this study was the simultaneous generation of the $[\text{BH}_2]^+$ ion at m/z 153, essentially being protonated xanthine ($\text{C}_5\text{H}_5\text{N}_4\text{O}_2^+$). By applying a CV of 30 V the protonated base moiety (m/z 153) was formed in-source and its product ion spectra recorded. In Figure 5, the product ion spectra of the protonated xanthine base at CE 15 eV and 30 eV are shown. The protonated xanthine primarily dissociated into the ions at m/z 136 ($\text{C}_5\text{H}_2\text{N}_3\text{O}_2^+$) and m/z 110 ($\text{C}_4\text{H}_4\text{N}_3\text{O}^+$) and a minor fragment at m/z 135 ($\text{C}_5\text{H}_3\text{N}_4\text{O}^+$). These fragments could be rationalized via a ring-opened intermediate of protonated xanthine in analogy to the ring opened protonated guanine (Scheme 2). This ring opened intermediate (m/z 153) is identical to the structure following the H_2O -addition/ NH_3 -elimination on the ring-opened intermediate of protonated guanine. In-source generation of the MS³-like product ions at m/z 110 and 136 (respective CVs: 55 V and 50 V) and their subsequent product ions in collision cell CID study supported this conclusion. The plots of the energy resolved spectra of the product ions related to m/z 110 from protonated xanthine matched exactly the plots of the m/z 110 originating from protonated guanine (not shown). The further dissociation of the xanthine-fragment at m/z 136 coincided with the represented pathway (Scheme 2).

In Table 3 a summary of all relevant product ions of the discussed spectra is given, which stresses once more the interrelations between the product ions of $[\text{BH}_2]^+$, $[\text{BH}_2]^+ - \text{NH}_3$, and $[\text{BH}_2]^+ - \text{HNCO}$ of protonated xanthosine and the $[\text{BH}_2]^+ - \text{NH}_3$, $[\text{BH}_2]^+ - \text{NH}_3 + \text{H}_2^{18}\text{O}$ and $[\text{BH}_2]^+ - \text{NH}_3 + \text{CH}_3\text{OH}$ of protonated guanosine. The absence of some of the predicted product ions in the corresponding product ion spectra of m/z 135 + H_2^{18}O (m/z 155, $\text{C}_5\text{H}_5\text{N}_4^{16}\text{O}^{18+}$) is attributed to too low abundances. Clearly, the m/z 155 ion population was rather limited. Moreover, the signal of m/z 155 was composed of several ionic structures and adducts, all fragmenting differently.

Conclusions

This study demonstrated that in-source CID and product ion spectra thereof must be handled with care, due to the creation of various solvent dependent ionic species. Initial fragment information might be lost as some of the in-source generated product ions might—to greater or lesser extent—be channelled directly into solvent-adducts before their actual MS/MS analysis. The reaction of the product ions with methanol and water occurs both in the electrospray source and, especially for water, also in the collision cell. Depending on the experimental settings, product ions resulting from reaction of guanosine derived ions with H_2O and CH_3OH could yield false positive results for the presence of xanthosine and/or methylated guanosines. Similar interactions with solvents might be expected for example for the nucleoside cytidine yielding uridine

Table 3. Summary of the product ions following the dissociation of the $[\text{BH}_2]^+$ (m/z 153), $[\text{BH}_2]^+-\text{NH}_3$ (m/z 136), $[\text{BH}_2]^+-\text{HNCO}$ (m/z 110) of protonated xanthosine and the product ions of the $[\text{BH}_2]^+-\text{NH}_3$ (m/z 135), $[\text{BH}_2]^+-\text{NH}_3 + \text{H}_2^{18}\text{O}$ (m/z 155), $[\text{BH}_2]^+-\text{NH}_3 + \text{CH}_3\text{OH}$ (m/z 167) of protonated guanosine

Exact m/z	Elemental Compositions	Guo m/z ($135+\text{H}_2^{18}\text{O}$)	Guo m/z ($135+\text{CH}_3\text{OH}$) ^b	Guo m/z 135	Xan m/z 110	Xan m/z 136	Xan m/z 153	Elemental Compositions	Nucleoside Exact m/z
156.0296	$\text{C}_5\text{H}_4\text{N}_3^{16}\text{O}_2^{18}\text{O}^+$	X	X					$\text{C}_6\text{H}_7\text{N}_4\text{O}_2^+$	167.0569
154.0253	$\text{C}_5\text{H}_4\text{N}_3^{16}\text{O}_3^+$		X	X		X	X	$\text{C}_5\text{H}_4\text{N}_3\text{O}_3^+$	154.0253
155.0455	$\text{C}_5\text{H}_5\text{N}_4^{16}\text{O}^{18}\text{O}^+$	X							
153.0413	$\text{C}_5\text{H}_5\text{N}_4^{16}\text{O}_2^+$	X	X	X			X	$\text{C}_5\text{H}_5\text{N}_4\text{O}_2^+$	153.0413
138.0189	$\text{C}_5\text{H}_2\text{N}_3^{16}\text{O}^{18}\text{O}^+$	X							
136.0159	$\text{C}_5\text{H}_2\text{N}_3^{16}\text{O}_2^+$	X ^a	X	X			X	$\text{C}_5\text{H}_2\text{N}_3\text{O}_2^+$	136.0147
137.0349	$\text{C}_5\text{H}_3\text{N}_4^{18}\text{O}^+$	X							
135.0307	$\text{C}_5\text{H}_3\text{N}_4^{16}\text{O}^+$	X	X	X			(X)	$\text{C}_5\text{H}_3\text{N}_4\text{O}^+$	135.0307
112.0397	$\text{C}_4\text{H}_4\text{N}_3^{18}\text{O}^+$	X							
110.0354	$\text{C}_4\text{H}_4\text{N}_3^{16}\text{O}^+$		X		X		X	$\text{C}_4\text{H}_4\text{N}_3\text{O}^+$	110.0354
110.0240	$\text{C}_4\text{H}_2\text{N}_3^{18}\text{O}^+$	X							
108.0198	$\text{C}_4\text{H}_2\text{N}_3^{16}\text{O}^+$	X	X			X	X	$\text{C}_4\text{H}_2\text{N}_3\text{O}^+$	108.0198
107.0358	$\text{C}_4\text{H}_3\text{N}_4^+$	X	X	X			X	$\text{C}_4\text{H}_3\text{N}_4^+$	107.0358
95.0131	$\text{C}_4\text{HN}_2^{18}\text{O}^+$	X							
93.0089	$\text{C}_4\text{HN}_2^{16}\text{O}^+$	X	X	X		X	X	$\text{C}_4\text{HN}_2\text{O}^+$	93.0089
85.0288	$\text{C}_3\text{H}_3\text{N}_2^{18}\text{O}^+$	X							
83.0245	$\text{C}_3\text{H}_3\text{N}_2^{16}\text{O}^+$		X		X		X	$\text{C}_3\text{H}_3\text{N}_2\text{O}^+$	83.0245
82.0405	$\text{C}_3\text{H}_4\text{N}_3^+$	X	X		X		X	$\text{C}_3\text{H}_4\text{N}_3^+$	82.0405
83.0131	$\text{C}_3\text{HN}_2^{18}\text{O}^+$	X							
81.0089	$\text{C}_3\text{HN}_2^{16}\text{O}^+$	X	X				X	$\text{C}_3\text{HN}_2\text{O}^+$	81.0089
80.0249	$\text{C}_3\text{H}_2\text{N}_3^+$	X	X	X			X	$\text{C}_3\text{H}_2\text{N}_3^+$	80.0249
69.0089	$\text{C}_2\text{HN}_2^{16}\text{O}^+$		X			X	X	$\text{C}_2\text{HN}_2\text{O}^+$	69.0089
68.0136	$\text{C}_3\text{H}_2\text{N}^{16}\text{O}^+$	X	X	X		X	X	$\text{C}_3\text{H}_2\text{NO}^+$	68.0136
68.0253	$\text{C}_2\text{H}_2\text{N}_3^+$			X*				$\text{C}_2\text{H}_2\text{N}^+$	68.0253
67.0296	$\text{C}_3\text{H}_3\text{N}_2^+$		X		X		X	$\text{C}_3\text{H}_3\text{N}_2^+$	67.0296
65.0140	C_3HN_2^+	X	X	X		X	X	C_3HN_2^+	65.0140
58.0197	$\text{C}_2\text{H}_2\text{N}^{18}\text{O}^+$	X							
56.0136	$\text{C}_2\text{H}_2\text{N}^{16}\text{O}^+$		X		X	X	X	$\text{C}_2\text{H}_2\text{NO}^+$	56.0136
55.0296	$\text{C}_2\text{H}_3\text{N}_2^+$	X	X	X	X		X	$\text{C}_2\text{H}_3\text{N}_2^+$	55.0296
53.9980	C_2NO^+	X				X	X	C_2NO^+	53.9980
53.0140	C_2HN_2^+	X	X	X	X*	X	X	C_2HN_2^+	53.0140

(X) Following a transition between different fragmentation pathways

^aMinor abundance^bOnly product ions shown which are common with the protonated xanthosine- or the m/z 135 out of protonated guanosine pathways

after hydrolytic deamination and methylated uridine after reaction with methanol.

The unexpected presence of radical cations in the spectra of these protonated molecules further complicates the obtained product ion pattern. High-resolution product ion spectra are required to resolve these radical cations from isobaric even-electron product ions and correct interpretation of the obtained data (Supplementary Material Tables 4, 5, and 6).

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References

1. March, R. E. An introduction to quadrupole ion trap mass spectrometry. *J. Mass Spectrom.* **1997**, *32*, 351–369.
2. Chernushevich, I. V.; Loboda, A. V.; Thomson, B. A. An introduction to quadrupole-time-of-flight mass spectrometry. *J. Mass Spectrom.* **2001**, *36*, 849–865.
3. Lemièrre, F.; Vanhoutte, K.; Jonckers, T.; Marek, R.; Esmans, E. L.; Claeys, M.; Van den Eeckhout, E.; Van Onckelen, H. Differentiation between isomeric phenylglycidyl ether adducts of 2'-deoxyguanosine and 2'-deoxyguanosine-5'-monophosphate using liquid chromatography/electrospray tandem mass spectrometry. *J. Mass Spectrom.* **1999**, *34*, 820–834.
4. Nunez, O.; Moyano, E.; Galceran, M. T. High mass accuracy in-source collision-induced dissociation tandem mass spectrometry and multi-step mass spectrometry as complementary tools for fragmentation studies of quaternary ammonium herbicides. *J. Mass Spectrom.* **2004**, *39*, 873–883.
5. Tuytten, R.; Lemièrre, F.; Van Dongen, W.; Esmans, E. L.; Witters, E.; Herrebout, W.; Van, D.V.; Dudley, E.; Newton, R. P. Intriguing mass spectrometric behavior of guanosine under low energy collision-induced dissociation: H₂O adduct formation and gas-phase reactions in the collision cell. *J. Am. Soc. Mass Spectrom.* **2005**, *16*, 1291–1304.
6. Dudley, E.; Tuytten, R.; Bond, A.; Lemièrre, F.; Brenton, A. G.; Esmans, E. L.; Newton, R. P. Study of the mass spectrometric fragmentation of pseudouridine: Comparison of fragmentation data obtained by matrix-assisted laser desorption/ionization post-source decay, electrospray ion trap multistage mass spectrometry, and by a method utilizing electrospray quadrupole time-of-flight tandem mass spectrometry and in-source fragmentation. *Rapid Commun. Mass Spectrom.* **2005**, *19*, 3075–3085.
7. Sleno, L.; Volmer, D. A. Ion activation methods for tandem mass spectrometry. *J. Mass Spectrom.* **2004**, *39*, 1091–1112.
8. Gabelica, V.; Lemaire, D.; Laprevote, O.; De Pauw, E. Kinetics of solvent addition on electrosprayed ions in an electrospray source and in a quadrupole ion trap. *Int. J. Mass Spectrom.* **2001**, *210*, 113–119.
9. Gregson, J. M.; McCloskey, J. A. Collision-induced dissociation of protonated guanine. *Int. J. Mass Spectrom.* **1997**, *165*, 475–485.
10. Frycak, P.; Huskova, R.; Adam, T.; Lemr, K. Atmospheric pressure ionization mass spectrometry of purine and pyrimidine markers of inherited metabolic disorders. *J. Mass Spectrom.* **2002**, *37*, 1242–1248.
11. Turecek, F.; Chen, X. H. Protonated adenine: Tautomers, solvated clusters, and dissociation mechanisms. *J. Am. Soc. Mass Spectrom.* **2005**, *16*, 1713–1726.
12. Yao, C. X.; Cuadrado-Peinado, M. L.; Polasek, M.; Turecek, F. Gas-phase tautomers of protonated 1-methylcytosine. Preparation, energetics, and dissociation mechanisms. *J. Mass Spectrom.* **2005**, *40*, 1417–1428.
13. Curcuruto, O.; Tarzia, G.; Hamdan, M. Differentiation between isomeric methylated purine derivatives by means of chemical ionization and collision-induced dissociation. *Rapid Commun. Mass Spectrom.* **1992**, *6*, 596–600.
14. Nelson, C. C.; McCloskey, J. A. Collision-induced dissociation of uracil and its derivatives. *J. Am. Soc. Mass Spectrom.* **1994**, *5*, 339–349.
15. Boos, K. S.; Grimm, C. H. High-performance liquid chromatography integrated solid-phase extraction in bioanalysis using restricted access pre-column packings. *TRAC-Trends Anal. Chem.* **1999**, *18*, 175–180.
16. Roberts, E. L. L.; Newton, R. P. Estimation of guanine deaminase using guanosine as a "prosubstrate". *Anal. Biochem.* **2004**, *324*, 250–257.