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Performance and stability of a dynamically controlled EBPR anaerobic/aerobic granular sludge reactor

**Reference:**

De Vleeschauwer Flinn, Caluwé Michel, Dobbeleers Thomas, Stes Hannah, Dockx Lennert, Kiekens Filip, D' aes Jolien, Copot Cosmin, Dries Jan.-  
Performance and stability of a dynamically controlled EBPR anaerobic/aerobic granular sludge reactor  
Bioresource technology - ISSN 0960-8524 - (2019), p. 1-29  
Full text (Publisher's DOI): <https://doi.org/10.1016/J.BIORTECH.2019.02.052>  
To cite this reference: <https://hdl.handle.net/10067/1570620151162165141>

1 **Performance and Stability of a Dynamically controlled EBPR Anaerobic/Aerobic**  
2 **Granular Sludge Reactor.**

3  
4 Short title: EBPR Anaerobic/aerobic AGS system with an anaerobic and  
5 aerobic dynamically controlled step

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19  
20 **Abstract**

21 Treatment of rapidly varying wastewaters in anaerobic/aerobic  
22 aerobic granular sludge (AGS) systems remains problematic. This  
23 study investigated AGS formation and the impact of varying COD  
24 and phosphorus concentrations on an enhanced biological  
25 phosphorus removal (EBPR) AGS SBR with a conductivity based  
26 anaerobic and OUR based aerobic dynamically controlled step.  
27 Phase 1 investigated the development of AGS. Phase 2 examined  
28 the flexibility of the dynamic control strategy and AGS efficiency while  
29 rapidly altering the influent composition. AGS was formed  
30 successfully in phase 1: the DV50 increased to 285 µm., and the  
31 SVI5 and SVI30 decreased to 51 and 40 ml/g respectively. In phase  
32 2 the effluent COD and PO<sub>4</sub>-P concentration remained low at  
33 respectively 58 ± 27 mg/L and 0.53 ± 0.77 mg/L. With an anaerobic  
34 DOC uptake efficiency of 98.4 ± 0.9 %.

35 Keywords: Anaerobic dynamic control strategy; synthetic wastewater; phosphate  
36 accumulating organisms (PAO)

## 37 1.0 Introduction

38 In the past 2 decades, the aerobic granular sludge (AGS) technology has  
39 shown to have high potential in the treatment of wastewater. A total of 42 full-  
40 scale sewage wastewater treatment plants and 6 full scale industrial wastewater  
41 treatment plants have been reported worldwide (website Nereda). The large  
42 difference between the number of domestic and industrial full-scale AGS  
43 wastewater treatment plants can be explained by the difficulty of treating  
44 complex and variable wastewater such as industrial wastewater. A possible  
45 AGS operational method for the treatment of industrial wastewater is a  
46 anaerobic/aerobic AGS sequencing batch reactor (SBR) with a low hydraulic  
47 selection pressure. An anaerobic/aerobic operation stimulates the growth of  
48 granules with slow growing polyphosphate accumulating and glycogen  
49 accumulating organisms (PAO and GAO) (de Kreuk et al 2004). This kind of  
50 AGS SBRs requires a complete COD uptake during the anaerobic step by PAO  
51 and GAOs. A weakness to this kind of systems is floccular sludge formation due  
52 to “aerobic COD leakage” resulting from an incomplete COD uptake in the  
53 anaerobic step. As such, aerobic COD leakage feeds the aerobic floccular  
54 sludge (de Pronk et al. 2015). With a static anaerobic step in the treatment  
55 process, the risk of aerobic COD leakage increases when using varying COD  
56 concentrations due to a possible incomplete COD uptake in the anaerobic step.

57 An innovative solution is the use of sensor guided dynamically controlled  
58 anaerobic/aerobic AGS systems. Such AGS systems would automatically

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59 elongate or shorten anaerobic and/or aerobic steps to ensure an optimal  
60 anaerobic and aerobic efficiency, hereby reducing aerobic COD leakage and  
61 improving the AGS formation.

62 Extensive research in static controlled anaerobic/aerobic AGS systems  
63 has been carried out (e.g. De Kreuk et al. 2004). In contrast, only a few reports  
64 on dynamically controlled anaerobic/aerobic AGS systems have been  
65 published. In a simultaneous nitrification denitrification (SND) anaerobic/aerobic  
66 AGS system treating potato industrial wastewater, the oxygen uptake rate  
67 (OUR) and the pH profile have been used successfully to control the duration of  
68 the aerobic step (Dobbeleers et al. 2017). Stes et al. (2018) have reported a  
69 successful implementation of an OUR based strategy in an anaerobic/aerobic  
70 AGS system treating brewery wastewater.

71 In an EBPR system the conductivity profile can be used to control the  
72 anaerobic step. The change in conductivity is well correlated with the change in  
73 the anaerobic  $PO_4$ -P release rate because of the additional release of  
74 potassium and magnesium ions during the anaerobic  $PO_4$ -P release. (Maurer et  
75 al. 1995). Kishida et al. (2008) have used the anaerobic conductivity profile and  
76 aerobic pH profile to dynamically control the anaerobic and aerobic steps in an  
77 EBPR AGS system, resulting in stable nitrogen and phosphorus removal even  
78 during 8 days of influent variations. Long term studies on AGS systems with  
79 changing influent compositions and a conductivity based anaerobic dynamically  
80 controlled step have not yet been reported.

81 The current study investigates the performance of an EBPR AGS SBR  
82 using a conductivity based anaerobic dynamic control strategy and an OUR

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83 based aerobic dynamic control strategy. The study was carried out in two  
84 phases. Phase 1 focuses on the formation and performance of an EBPR AGS  
85 SBR while applying an anaerobic and aerobic dynamic control strategy and  
86 feeding the reactor with an influent with a constant composition. Phase 2  
87 investigates the prolonged effect on the EBPR AGS system efficiency and the  
88 impact on the anaerobic and aerobic dynamic control strategy while feeding the  
89 reactor an influent with rapidly varying compositions.

## 90 **2.0 Material and methods**

### 91 **2.1 Reactor set-up and operation**

92 The study was carried out in two phases. Phase 1 (reactor A) investigated  
93 AGS formation in an anaerobic/aerobic dynamically controlled EBPR AGS SBR.  
94 Phase 2 (reactors B and C) examined the flexibility of the dynamic control  
95 strategy and the AGS efficiency when feeding influent with rapidly varying  
96 compositions. Reactor C was the control reactor which was fed a constant  
97 influent composition.

98 In the first phase the reactor had a working volume of 11.1L. For the  
99 second phase both reactors had a working volume of 6L. The reactors were  
100 equipped with an influent peristaltic pump (Watson Marlow®), a mixer  
101 (Heidolph® RZR2020), a discharge valve (Eriks RX ER10.X33.S00) and an  
102 aeration system consisting of an aeration pump (koi flow 60, Aquadistri China®)  
103 and a 13 cm aeration disc (Aquadistri China®). Furthermore the reactors were  
104 equipped with a luminescent dissolved oxygen (LDO) sensor (Hang Lange®), a  
105 pH sensor (JUMO® BlackLine) and a 3798-S digital inductive conductivity

106 sensor (Hach Lange®). All sensor values were logged. At the end of each  
107 cycle the values were archived.

108 The SBRs were operated with a custom built National Instruments  
109 LabVIEW® program, a Siemens PLC and a Phoenix IO. The reactor cycles had  
110 9 steps; idle step (10 seconds); aerated pre-step (30 minutes); unaerated step  
111 (10 minutes); anaerobic influent feeding step (flow dependent); anaerobic step  
112 (dynamic); aerobic step (dynamic); sludge settling step (10 minutes); discharge  
113 step (5 minutes); inactive step (1 minute). The duration of the feeding step  
114 varied between 1 and 2 minutes.

115 Reactor A was seeded with 10.1L sludge from a local EBPR activated  
116 sludge municipal wastewater treatment plant (Aquafin Antwerpen Zuid). The  
117 seed sludge MLSS, MLVSS, SVI30 and DV50 values were respectively 3.712  
118  $\pm 0.011$  g/L, 2.682  $\pm 0.216$  g/L, 76 ml/g and 128  $\pm 2.16$   $\mu$ m. Reactor B and C were  
119 seeded with sludge from reactor A.

120 During phase 1 reactor A was fed 1L influent per cycle, had a working  
121 volume of 11.1L and a volume exchange rate (VER) of about 9%. The VER of  
122 reactor B and C in phase 2 was approximately 8% with a influent feed of 0.48 L  
123 and a working volume of 6L. The sludge retention time (SRT) was kept at 30  
124 days throughout the study. The hydraulic retention time (HRT) and organic  
125 loading rate (OLR) varied due to the dynamic cycle durations and the varying  
126 influent composition in phase 2.

## 127 **2.2 Influent composition**

128 The reactors were fed with synthetic wastewater. The influent composition  
129 of reactor A and C was kept constant (Influent 1): 1250 mgCOD/L (Na-

130 propionate), 25 mgPO<sub>4</sub>-P/L (K<sub>2</sub>HPO<sub>4</sub>), 25 mgNH<sub>4</sub>-N/L (NH<sub>4</sub>Cl), 75 mgCO<sub>3</sub><sup>2-</sup>/L  
131 (NaHCO<sub>3</sub><sup>2-</sup>), 26 mgMg/L (MgSO<sub>4</sub>·6H<sub>2</sub>O), 56 mgK<sup>1+</sup>/L (KCl) and 1ml/L trace  
132 solution (Vishniac and Santer 1957). To inhibit nitrification, N-allylthiourea (ATU)  
133 was added to the influent at 10 mg/L.

134 The influent composition of reactor B was varied. The impact of three  
135 influent variation strategies (IVS) was investigated. 1) IVS<sub>1</sub> (day 8 to day 62):  
136 The COD and PO<sub>4</sub>-P concentrations were varied step by step, first with small  
137 COD and PO<sub>4</sub>-P variations (± 15% compared to influent 1, COD concentration  
138 between 1156 and 1437 mg/L) followed by increasing variations (up to ± 40%  
139 compared to influent 1, COD concentration between 750 and 1750 mg/L).  
140 Meanwhile the COD/P ratio was kept constant at 50. 2) IVS<sub>2</sub> (day 63 to day  
141 99): the PO<sub>4</sub>-P concentration was varied step by step (± 60% compared to  
142 influent 1, PO<sub>4</sub>-P concentration between 10 and 40 mg/L) while the COD  
143 concentration was kept constant at 1250 mg/L with Na-propionate. 3) IVS<sub>3</sub> (day  
144 100 to day 115): The influent composition strategies of IVS<sub>1</sub> and IVS<sub>2</sub> were  
145 combined. The COD/P ratio was varied (between 21 and 116) by changing both  
146 the COD and PO<sub>4</sub>-P concentrations step by step (± 40% compared to influent  
147 1). The ATU and CO<sub>3</sub><sup>2-</sup> concentrations and the COD/N/K/Mg ratios were kept  
148 constant.

### 149 **2.3 Dynamic control strategy**

150 The anaerobic and aerobic dynamically controlled systems both worked as  
151 closed-loop control systems. Sensor profile patterns determined the duration of  
152 the anaerobic and aerobic step.

153 The aerobic step was dynamically controlled by means of the oxygen  
154 uptake rate (OUR). The OUR calculation and the aerobic dynamic control  
155 strategy were used as described in Dobbeleers et al. (2017). The oxygen level  
156 in the reactors were controlled with an on/off aeration control, keeping the  
157 oxygen concentration between 1 and 2 mg/L. This resulted in an oxygen  
158 profile with alternating positive and negative flanks. A custom built LabVIEW  
159 program calculated the OUR (mgO<sub>2</sub>/L.h) during the negative flanks. The  
160 aerobic dynamic control strategy used two set point, SP<sub>OUR1</sub> and SP<sub>OUR2</sub>. When  
161 a fixed number (SP<sub>OUR1</sub>) of consecutive OUR values dropped below a minimal  
162 set point (SP<sub>OUR2</sub>), the aerobic step was terminated. The SP<sub>OUR1</sub> and SP<sub>OUR2</sub>  
163 were periodically adjusted to ensure a complete PO<sub>4</sub>-P uptake. SP<sub>OUR2</sub> was  
164 adjusted to ensure that the specific respiration speed (SOUR) decreased to 2.5  
165 mg/g<sub>MLVSS</sub>.h. SP<sub>OUR2</sub> was increased or decreased to guarantee that the effluent  
166 PO<sub>4</sub>-P was below 1 mg/L. To rule out premature or overdue terminations of the  
167 aerobic step, the aerobic dynamic control strategy had a built-in minimal and  
168 maximal duration of respectively 1 and 5 hours. The aeration step in reactor A, B  
169 and C were operated with the same aerobic dynamic control strategy.

170 As described by Kishida et al. (2008), the duration of the anaerobic step in  
171 an EBPR AGS SBR can be controlled by the conductivity slope (μS/m.d). The  
172 change in conductivity is well correlated to the PO<sub>4</sub>-P concentration. The  
173 additional release of magnesium and potassium ions during the phosphate  
174 release contributes to the increase in the conductivity (Maurer et al. 1995).  
175 When the anaerobic phosphate release terminates, the conductivity slope  
176 becomes equal to zero, hereby giving a good indication of the PO<sub>4</sub>-P  
177 concentration profile.



178 The anaerobic dynamic control system calculates the moving conductivity slope  
179 (MCS) during the anaerobic step. The control strategy uses 4 calculation  
180 variables (CV); the intervals of the sensor value ( $CV_1$ ) and the number of sensor  
181 values per MCS calculation ( $CV_2$ ). The cut-off point is determined by a minimal  
182 MCS ( $CV_3$ ) and the number of MCS's ( $CV_4$ ) that meet the cut-off requirements.  
183 In total the anaerobic dynamic control strategy ran through 3 loops to  
184 dynamically control the duration of the anaerobic step (Figure 1).

185 To obtain an accurate and stable anaerobic step termination the optimal  
186 CVs set points were tested by looking at the conductivity, pH, DOC uptake and  
187  $PO_4$ -P release profiles of an AGS SBR. The use of higher  $CV_1$  and  $CV_2$  values  
188 resulted in smooth MCS profiles (high stability) but caused a longer delay on the  
189 cut-off point (low accuracy). A  $CV_1$  value of 60 seconds and a  $CV_2$  value of 10  
190 resulted in the stable MCS profile with an acceptable delay ( $\pm 10$  minutes delay).  
191 During phase 1 the  $CV_3$  value was adjusted depending on the evolution of the  
192 DOC uptake and  $PO_4$ -P release profiles. The  $CV_4$  value was kept constant at  
193 20. The  $CV_3$  and  $CV_4$  values remained unchanged in phase 2. To rule out  
194 premature or overdue terminations of the anaerobic step, the anaerobic  
195 dynamic control strategy had a built-in minimal and maximal duration of  
196 respectively 20 and 120 minutes. The calculated MCS values were logged and  
197 archived at the end of each cycle.

## 198 **2.4 Analytical Methodology**

199 The chemical analyses (total and soluble chemical oxygen demand (COD,  
200 sCOD), phosphorus-orthophosphate ( $PO_4$ -P), and nitrogen-ammonium ( $NH_4$ -N))  
201 were analysed with test kits from Hanna Instruments Belgium respectively

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202 HI93754A-25, HI93754B-25, HI93706-01 and HI93715-01. The potassium ( $K^{1+}$ )  
203 and Magnesium ( $Mg^{2+}$ ) concentrations were analysed with Hach Lange® test  
204 kits, respectively LCK 328 and LCK 326. The dissolved organic carbon (DOC)  
205 was analysed with a Sievers InnovOx® laboratory total organic carbon  
206 analyser. All parameters except the COD were measured on pre-filtered  
207 samples (VWR® glass microfibers filter, 693, particle retention: 1.2 $\mu$ m).

208 The sludge mixed liquor suspended (volatile) solids (ML(V)SS) and the  
209 sludge volume index (SVI) were measured according to the standard methods  
210 (APHA, 1998). Microscopic images were taken with a MOTIC BA310  
211 microscope (Opti-service, Belgium).

212 The sludge particle size distribution (DV50) was measured with a Malvern  
213 Mastersizer 3000 as described by Caluwé et al. (2017). To study the evolution  
214 of the particle size classes (>200 $\mu$ m), the volume fraction larger than 200  $\mu$ m  
215 ( $VF_{>200\mu m}$ ) was calculated.

216 Reactor C was operated as a control reactor, only the sludge particle size  
217 distribution (DV50) was analysed.

## 218 **2.5 In-situ cycle measurement**

219 To test the effectiveness of the anaerobic dynamic control strategy, the  
220 DOC uptake efficiency and the  $PO_4$ -P release efficiency were measured at  
221 different moments during the cycle. During the anaerobic step 3 grab samples  
222 were taken (Figure 2): at sample point a (SPa) the start of the anaerobic step; at  
223 sample point b (SPb), during the anaerobic step (conductivity profile dependent,  
224 when the conductivity slope changed from positive to negative); at sample point  
225 c (SPc), at the end of the anaerobic step. During the aerobic step in phase 1, a

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226 sample was taken every hour and at the end of the aerobic step. The aerobic  
227 sample frequency was altered in phase 2 to study the relationship between the  
228 conductivity profile and the phosphate concentration. Two samples were taken  
229 during the aerobic **step: one at** sample point d (SPd) (conductivity profile  
230 **dependent, when the conductivity slope changed from negative to positive) and**  
231 **one** at sample point e (SPE) at the end of the aerobic step (Figure 2). The DOC  
232 and PO<sub>4</sub>-P concentrations were measured. The anaerobic DOC uptake  
233 efficiency and the anaerobic PO<sub>4</sub>-P release efficiency were calculated with  
234 following formulas:

$$\text{Anaerobic DOC uptake (\% at (b))} = 100x \frac{DOC_{(a)} - DOC_{(b)}}{DOC_{(a)} - DOC_{(e)}}$$

$$\text{Total anaerobic DOC uptake (\% at (c))} = 100x \frac{DOC_{(a)} - DOC_{(c)}}{DOC_{(a)} - DOC_{(e)}}$$

$$\text{Anaerobic PO}_4\text{(P) release (\% at (b))} = 100x \frac{PO_4\text{(P)}_{(b)} - PO_4\text{(P)}_{(a)}}{PO_4\text{(P)}_{(c)} - PO_4\text{(P)}_{(a)}}$$

237 With (a) 30 seconds after influent feed, (b) at the sample points

239 (conductivity profile dependent), (c) at the end of the anaerobic step and (e) at  
240 the end of the aerobic step (Figure 2). To take into account immediately  
241 adsorbed DOC, the DOC concentration at SPa was theoretically calculated  
242 using the influent DOC, the effluent DOC and VER.

## 243 **2.6 DNA extraction**

244 For molecular PAO and GAO community quantification and gene  
245 sequencing analysis, 250 µl AGS samples were taken from the reactor

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246 periodically and preserved at -80°C. The DNA was extracted according to the  
247 method described in Mcilroy et al. (2008) and stored at -80°C.

## 248 **2.7 qPCR molecular quantification**

249 A PAO541f/PAO846r primer and a GAOQ989f/GAM1278r primer were  
250 used to **target respectively** PAO (16S rRNA Candidatus Accumulibacter  
251 phosphatis) and GAO (16S rRNA Candidatus Competibacter phosphatis)  
252 organisms. Universal 1055f/Universal 1392r primers were used to measure the  
253 total amount of **bacterial DNA** (16S rRNA Universal bacteria). The amount of  
254 target cells per gMLVSS were calculated from de measured DNA  
255 concentrations. **The DNA concentrations were quantified** according to the  
256 method described in Caluwé et al. (2017).

## 257 **2.8 Microbial community composition**

258 The microbial community composition was obtained by 16S rRNA gene  
259 **amplification**. Barcode primers (IDT) and Phusion High-Fidelity DNA  
260 polymerase (Thermo Scientific) were used to generate amplicons targeting the  
261 V 1-3 region of the 16S rRNA gene as described by Kozich et al. (2013). The  
262 purification of the PCR products was carried out **with the S**equalPrep  
263 Normalization plate kit (Invitrogen) before being pooled. Afterwards NucleoSpin  
264 Gel and PCR Clean-up (Macherey Nagel) was used to obtain an enhanced  
265 purified library. Before amplicon sequencing, the library was diluted resulting in  
266 a 2 nM library. At last the amplicon sequencing was executed on an Illumina  
267 Miseq system at the Centre for Medical Genetics (Edegem, Belgium) with the  
268 MiSeq Reagent Kit v2 (Illumina). The UPARSE pipeline (Edgar et al. 2013) was  
269 used to process the paired-end reads. The **results were** compared, for

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270 taxonomy predictions of the operational taxonomic unit (OTU) sequences, to a  
271 reference database; MiDAS (version 2.1). A name is proposed for all the  
272 abundant genus-level taxa present in activated sludge, anaerobic digesters and  
273 influent wastewater. The reference database is a manually curated SILVA 16S  
274 rRNA taxonomy library (release 1.23 Ref NR99) (McIlroy et al. 2015).

## 275 **3.0 Results and Discussion**

### 276 **3.1 Phase 1**

277 During phase 1 the formation of granules in an EBPR AGS SBR with  
278 dynamically controlled anaerobic and aerobic steps was investigated for 105  
279 days. The duration of the anaerobic step was controlled using the conductivity  
280 MCS. The duration of the aerobic step was controlled using the OUR. AGS was  
281 formed successfully during phase 1 with an anaerobic and aerobic dynamic  
282 control strategy.

#### 283 **3.1.1 Reactor operation and removal efficiency**

284 The first three days the anaerobic step was controlled statically with a  
285 fixed duration of 2 hours. From the fourth day onward the anaerobic dynamic  
286 control strategy was applied. The anaerobic dynamic set points  $CV_1$ ,  $CV_2$  and  
287  $CV_4$  were kept respectively at 60(sec), 10 (#) and 20 (#). To optimize the  
288 anaerobic step, the cut-off point ( $CV_3$ ) was systematically increased depending  
289 on the stabilization of the conductivity profile.

290 The reliability, or degree of success, of the anaerobic and aerobic dynamic  
291 control strategy was 82.4 %. The control detected the endpoint of the anaerobic

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292 step and terminated the step accordingly. Two factors had a negative effect on  
293 the reliability. First, the sporadic activation of the minimal and maximal  
294 anaerobic and aerobic step durations occurred due to ineffective filling steps  
295 resulting from detached influent tubing. Secondly, a continuous aeration from  
296 day 34 to 35 due to an aeration pump malfunction resulted in the loss of the  
297 anaerobic PAO conductivity profile. In-situ PO<sub>4</sub>-P analysis showed no anaerobic  
298 PO<sub>4</sub>-P release. A possible cause could be the loss of the microbial stored poly-  
299 PO<sub>4</sub>-P due to aerobic release and sequential wash-out of the PO<sub>4</sub>-P in the  
300 effluent. The anaerobic step was operated statically (1.25 h) from day 36-42. On  
301 day 43 the anaerobic PAO conductivity profile was re-established and the  
302 anaerobic dynamic control strategy was reactivated.

303 The anaerobic conductivity increase is strongly correlated to the PO<sub>4</sub>-P  
304 release (Maurer et al. 1995). Therefore the evolution of the MCS profile is a  
305 good representation of the evolution of the PAO activity. During the first 56  
306 days, the maximal MCS values did not evolve, stabilizing at  $1.55 \pm 0.14$   
307  $\mu\text{S}/\text{cm}\cdot\text{d}$ . As time progressed the maximal MCS values increased to  $4.04 \pm 0.36$   
308  $\mu\text{S}/\text{cm}\cdot\text{day}$  (day 91-110). The increase in the conductivity MCS indicates an  
309 increased phosphate release rate. Furthermore the anaerobic step duration is  
310 by design strongly correlated to the conductivity MCS profile. While the  
311 maximal MCS values remained stable during the first  $\pm 56$  days, the anaerobic  
312 step duration continually decreased, starting at  $1.680 \pm 0.23$  hours (day 1-7)  
313 and systematically decreasing over a period of 73 days to  $0.790 \pm 0.076$  hours  
314 (day 68-73). The anaerobic step varied more during first 50 days, these  
315 variations are possible caused by an increasing sludge activity as the sludge

316 concentration increased and the sludge metabolism adapted to the influent  
317 composition and reactor operation strategy.

318 From day 65 to 110 the reactor showed a stabilized operation, the duration  
319 of the anaerobic step and the starting MCS value were respectively  $0.737 \pm$   
320  $0.069$  hour and  $3.42 \pm 0.59$   $\mu\text{S}/\text{cm}\cdot\text{day}$ .

321 The sludge PAO activity during stable operation was confirmed through in-  
322 situ measurements. The DOC uptake efficiency at SPc and SPd (Figure 2) were  
323 respectively  $96.9 \pm 1.8$  % and  $97.8 \pm 1.4$  %. The anaerobic  $\text{PO}_4\text{-P}$  release  
324 efficiency at SPc was  $94.4 \pm 0.3$  %.

325 As reported by Sato et al. (1992), Pijuan et al. (2005) and Oehmen et al.  
326 (2005) the anaerobic P release/COD uptake ratio (Pmol/Cmol) of an EBPR  
327 system varies between 0.30 to 0.45 Pmol/Cmol. During phase 1 the anaerobic  
328 P release/COD uptake ratio during the stable operation was  $0.24 \pm 0.04$   
329 Pmol/Cmol. This is lower than reported values. Oehmen et al. (2006) suggested  
330 that lower anaerobic P release/COD uptake ratios could indicate a possible  
331 GAO competition for the available substrate during the anaerobic step.

332 From day 1 to day 110 the aerobic step was dynamically controlled by  
333 means of the OUR.  $\text{SP}_{\text{OUR}2}$  was set at  $10 \text{ mgO}_2/\text{L}\cdot\text{h}$ . This was reduced to  $8$   
334  $\text{mgO}_2/\text{L}\cdot\text{h}$  after the first 7 days to improve the  $\text{PO}_4\text{-P}$  uptake efficiently.  $\text{SP}_{\text{OUR}2}$   
335 was kept at 3 for the entire phase 1. The aerobic step took on average  $3.09 \pm$   
336  $0.54$  h. The OUR profile was strongly correlated to the  $\text{PO}_4\text{-P}$  concentration with  
337  $R^2$  of 0.9992 and a p-value of  $< 0.0008$  (Alfa = 0.05).

338 During the aeration, the aerobic conductivity profile initially decreased but  
339 at a later stage increased again (Figure 3 (a)). In-situ measurements of

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340 magnesium ( $Mg^{2+}$ ) and potassium ( $K^{1+}$ ) show a decrease of  $Mg^{2+}$  and  $K^{1+}$   
341 concentrations during the conductivity decrease. However the  $K^{1+}$  concentration  
342 increased again at the end of the aerobic step. This indicates that the increase  
343 in  $K^{1+}$  is potentially the reason for the aerobic conductivity increase.

344 The anaerobic OLR increased as the sludge PAO activity increased,  
345 starting at  $1.726 \pm 0.225$  kgCOD/m<sup>3</sup>.d and from day 70 to 110 stabilizing at  
346  $3.839 \pm 0.148$  (Figure 3 (d)). This had little effect on the total OLR, which  
347 remained stable throughout phase 1 at  $0.620 \pm 0.077$  kgCOD/m<sup>3</sup>.d.

348 The COD removal efficiency was  $97.5 \pm 1.1\%$ . No change in the COD  
349 efficiency was observed. The aeration pump malfunction between day 33-35  
350 had no effect on the COD removal efficiency, while the impact on the PO<sub>4</sub>-P  
351 removal efficiency was significant, decreasing from  $90.6 \pm 5.9\%$  to  $81.2 \pm$   
352  $2.9\%$ . The removal efficiency increased again to  $90.7 \pm 8.5\%$  between day 65-  
353 110.

### 354 3.1.2 Aerobic Granular Sludge formation and qPCR analysis

355 The application of an EBPR operation and an anaerobic and aerobic  
356 dynamic control strategy resulted in the formation of AGS. On the basis of the  
357 MLSS, MLVSS, the particle size distribution (Figure 3 (c)) and microscopic  
358 images the sludge morphology experienced three distinct stages (stage 1, 2  
359 and 3): a start-up stage, an AGS germination stage and an AGS maturation  
360 stage.

361 During the first 20 days (stage 1) the MLSS and MLVSS decreased due to  
362 the adaptation of the sludge to an EBPR SBR operation and the implementation  
363 of a sludge retention time of 30 days. The DV50 increased and the  $VF_{>200\mu m}$



364 increased from 20,0 % to 40,6 %. Microscopic images show a decrease of small  
365 sludge particles.

366 **Between day 20 and 57** (stage 2) the MLSS and **MLVSS** increased and  
367 the DV50 stabilized. The  $VF_{>200\mu\text{m}}$  stabilized at  $37,7 \pm 3,1\%$ . Microscopic images  
368 show the reappearance of small sludge agglomerates, indicating the potential  
369 growth of small granule-like **structures**.

370 From day **57 until day 110** (stage 3) the MLSS and MLVSS stabilized. The  
371 DV50 and the  $VF_{>200\mu\text{m}}$  increased continually, from respectively 153  $\mu\text{m}$  to 285  
372  $\mu\text{m}$  and 40.1 % to 60.2%. The granules **increased** in size during this stage  
373 without an increase in MLSS indicating the development of AGS.

374 Additionally qPCR and microbial community composition analysis showed  
375 an increase in *Candidatus Accumulibacter phosphatis* a PAO **(Figure 4)**. The  
376 abundance **of** *Candidatus Competibacter phosphatis* (a GAO) remained low;  
377  $0.58 \pm 1.06\%$ . This indicates that the combination of the applied influent COD/P  
378 ratio **(approximately 50)** and an anaerobic/aerobic operational strategy was  
379 successful in selecting PAOs over GAOs. Weissbrodt et al. (2013) reported a  
380 similar OTU value of 25 % while feeding propionate based wastewater in an  
381 anaerobic fed anaerobic/aerobic AGS SBR.

## 382 **3.2 Phase Two**

### 383 **3.2.1 Reactor operation and removal efficiency**

384 During phase 2 the impact of rapidly varying influent concentrations on the  
385 efficiency of the anaerobic and aerobic dynamic control strategy and ASG  
386 efficiencies were investigated. The sludge **from** reactor A was divided in reactor  
387 B and C. Reactor C functioned as a **control and sludge backup reactor** and was

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388 fed the same influent composition as reactor A. In reactor B three influent  
389 composition variations were tested; IVS<sub>1</sub>, IVS<sub>2</sub> and IVS<sub>3</sub>.

390 The anaerobic dynamic control strategy was immediately implemented at  
391 the start of phase 2. For the entire phase 2, the aerobic and anaerobic control  
392 strategy of reactor B and C were operated with the same set-points (CV<sub>1</sub>, CV<sub>2</sub>,  
393 CV<sub>3</sub> and CV<sub>4</sub>) as reactor A at the end of phase 1.

394 In reactor B, the reliability of the anaerobic and aerobic dynamic control  
395 strategy during phase 2 was 98.2 %. Automatic activation of the set minimal  
396 and maximal anaerobic and aerobic step durations occurred sporadically due to  
397 ineffective filling steps.

398 The anaerobic control strategy was able to adjust the anaerobic step  
399 according to the influent COD. The anaerobic MCS profile increased and  
400 decreased with an increasing and decreasing influent COD concentration, as  
401 was previously reported by Kishida et al. (2008). During IVS<sub>1</sub> a linear correlation  
402 between the influent COD and anaerobic step duration was observed with an R<sup>2</sup>  
403 of 0.702 and a p-value of < 0.001 ( $\alpha = 0.05$ ), whereas no significant correlation  
404 was present during IVS<sub>2</sub> and IVS<sub>3</sub> when the COD/P ratio was varied. The  
405 influent PO<sub>4</sub>-P and COD/P ratio had no impact on the anaerobic step duration  
406 during IVS<sub>2</sub> and IVS<sub>3</sub> (Figure 5 (a) and (d)).

407 The anaerobic DOC uptake and PO<sub>4</sub>-P release efficiencies at grab sample  
408 points SPb and SPc (Figure 2) during phase 2 did not change significantly  
409 during IVS<sub>1</sub>, IVS<sub>2</sub> and IVS<sub>3</sub> (Figure 5(c)). While feeding rapidly varying influent  
410 compositions, the anaerobic dynamic control strategy was able to ensure a  
411 complete anaerobic DOC uptake.

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412 A significant linear **relation** between the anaerobic OLR and the influent  
413 COD was observed during IVS<sub>1</sub> and IVS<sub>3</sub>, **with** respectively **an** R<sup>2</sup> of 0.982 and  
414 a p-value of < 0.001, and **an** R<sup>2</sup> of 0.962 and a p-value of < 0.001 (Figure 5 (a)  
415 and (b)). Yet the anaerobic control reduced the impact of the influent COD  
416 variation on the anaerobic OLR (**Error! Reference source not found.(d)**).  
417 Whereas the influent COD variation of 40% would cause a theoretical ORL  
418 variation of around 31% in a static anaerobic step, the anaerobic ORL varied  
419 around 20%. The influent **PO<sub>4</sub>-P** concentration and the COD/P ratio had no  
420 impact on the anaerobic ORL. No linear **relation** was observed (Figure 5  
421 **5(a) and (b)**).

422 The anaerobic P release/COD uptake ratio remained high during phase 2,  
423 averaging at 0.42 ± 0.05 during IVS<sub>1</sub> and 0.35 ± 0.09 during IVS<sub>2</sub> and IVS<sub>3</sub>. The  
424 high anaerobic P release/COD uptake ratio indicates a mature PAO sludge  
425 (Oehmen et al. 2006).

426 The aerobic phase was dynamically controlled depending on the OUR.  
427 The influent COD had an impact on the aerobic step duration and the ORL  
428 (**Error! Reference source not found.**), but this impact was not significant. The  
429 influent PO<sub>4</sub>-P and COD/P ratio had no impact on the aerobic step duration or  
430 the ORL. The main objective of the OUR control was to ensure a complete  
431 PO<sub>4</sub>-P uptake during the aerobic step. **As shown in** Figure 5(d) **the aerobic step**  
432 **periodically showed wide variations. These were caused by the combination of**  
433 **(1) the slow OUR calculation rate (± 20 minutes per value at an endogenous**  
434 **OUR) and (2) the OUR control strategy requirement to have 3 OUR values**  
435 **(SP<sub>OUR1</sub>) below the endogenous set point. This resulted in elongated aerobic**  
436 **steps.**

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437 The rapid influent composition variations had no observed effect on the  
438 effluent quality. The control strategy was able to adjust the anaerobic and  
439 aerobic step durations to ensure optimal COD and PO<sub>4</sub>-P removal. The COD  
440 and PO<sub>4</sub>-P removal efficiency during the different influent variation strategies  
441 remained stable at respectively  $95.2 \pm 2.7\%$  and  $97.7 \pm 3.8\%$ . The average  
442 COD and PO<sub>4</sub>-P effluent concentrations during phase 2 were respectively  $58 \pm$   
443  $27 \text{ mg/L}$  and  $0.53 \pm 0.77 \text{ mg/L}$ . This is close to the reported effluent PO<sub>4</sub>-P  
444 concentration by Kishida et al. (2008), lower than  $< 0.3 \text{ mg/L}$ . The conductivity  
445 based anaerobic and OUR based aerobic dynamic control strategy was highly  
446 effective at achieving low COD and PO<sub>4</sub>-P concentrations even during a  
447 prolonged period of rapidly varying influent concentrations. Additionally varying  
448 COD/P ratios had little effect on the PO<sub>4</sub>-P removal efficiency. This increases  
449 the potential applications of the control strategy in EBPR systems for the  
450 treatment wastewater with a varying composition.

### 451 **3.2.2 Aerobic Granular Sludge morphology**

452 During phase 2 the AGS in reactor B and C evolved into a hybrid (granular  
453 and floccular) sludge. In reactor B the particle distribution (DV50) decreased  
454 from  $238 \mu\text{m}$  to  $129 \mu\text{m}$  indicating the formation of hybrid sludge. The same  
455 evolution was observed in the control reactor C, the particle distribution (DV50)  
456 decreased from  $200 \mu\text{m}$  to  $135 \mu\text{m}$ . The microscopic images confirmed the  
457 evolution into hybrid sludge. Granules remained visible throughout phase 2.  
458 The degradation of the AGS in reactor B was not due to applied feeding  
459 strategy because a similar AGS degradation was observed in the control reactor  
460 C. A possible factor that could have led to the AGS degradation is the reactor

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461 temperature, which was not controlled and thus varied depending on the room  
462 temperature..

## 463 4.0 Conclusion

464 The present study investigated a conductivity based anaerobic and OUR  
465 based aerobic dynamic control strategy for the removal of PO<sub>4</sub>-P in an EBPR  
466 AGS SBR which was first fed a constant and then by a rapidly varying  
467 wastewater. AGS was successfully formed. The effluent COD and PO<sub>4</sub>-P  
468 concentrations remained low during the whole study. The anaerobic and  
469 aerobic dynamic control strategy elongated and shortened the step durations in  
470 line with the influent COD. No aerobic COD leakage was observed.

471 E-supplementary data of this work can be found in the online version of  
472 the paper.

## 473 5.0 Acknowledgments

474 This work was financed by the University of Antwerp through the industrial  
475 research fund (IOF) for strategic basic research and was carried out in the  
476 research group BioGEM of the Faculty of Applied engineering.

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536 DESCRIPTION FIGURES

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3 537 Figure 1: (a) Schematic overview of the anaerobic dynamic control strategy,  
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5 538 consisting of 3 loops; Loop 1, intermittent logging (every  $CV_1$  seconds) of paired  
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7 539 conductivity and timestamp values and building conductivity and timestamp  
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9 540 arrays (until the number of pairs in the array reaches a set point value  $CV_2$ );  
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11 541 Loop 2, intermittent calculation (every  $CV_1$  seconds) of the MCS and comparing  
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13 542 the MCS to the maximal MCS set point (until the calculated MCS surpasses the  
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15 543 maximal MCS set point  $CV_3$ ); Loop 3, intermittent calculation (every  $CV_1$   
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17 544 seconds) of the MCS, comparing the MCS to the maximal MCS set point ( $CV_3$ )  
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19 545 and comparing the counted number of correct MCS to a set point amount to  
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21 546 determine anaerobic step termination (until the amount of correct MCS  
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23 547 surpasses the maximal amount of the correct MCS set point  $CV_4$ ). (b) The  
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25 548 anaerobic conductivity and MCS profile of the anaerobic dynamic control  
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27 549 strategy, (○) MCS, (■) conductivity.

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38 551 Figure 2: Cycle conductivity profile and grab sample points (SP); SPa, SPb,  
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40 552 SPc, SPd and SPe. Highlighted area is the anaerobic step.

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47 554 Figure 3: Data phase 1; (a) in-situ profiles of reactor A on day 86, (b) OUR and  
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49 555 anaerobic and aerobic conductivity MCS profiles on day 86, (c) MLSS, SVI5,  
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51 556 SVI30 and DV50 profile of reactor A, (d) Anaerobic time, total OLR and  
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53 557 anaerobic OLR profiles of reactor A. The highlighted area in figure (a) and (b)  
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55 558 are anaerobic.

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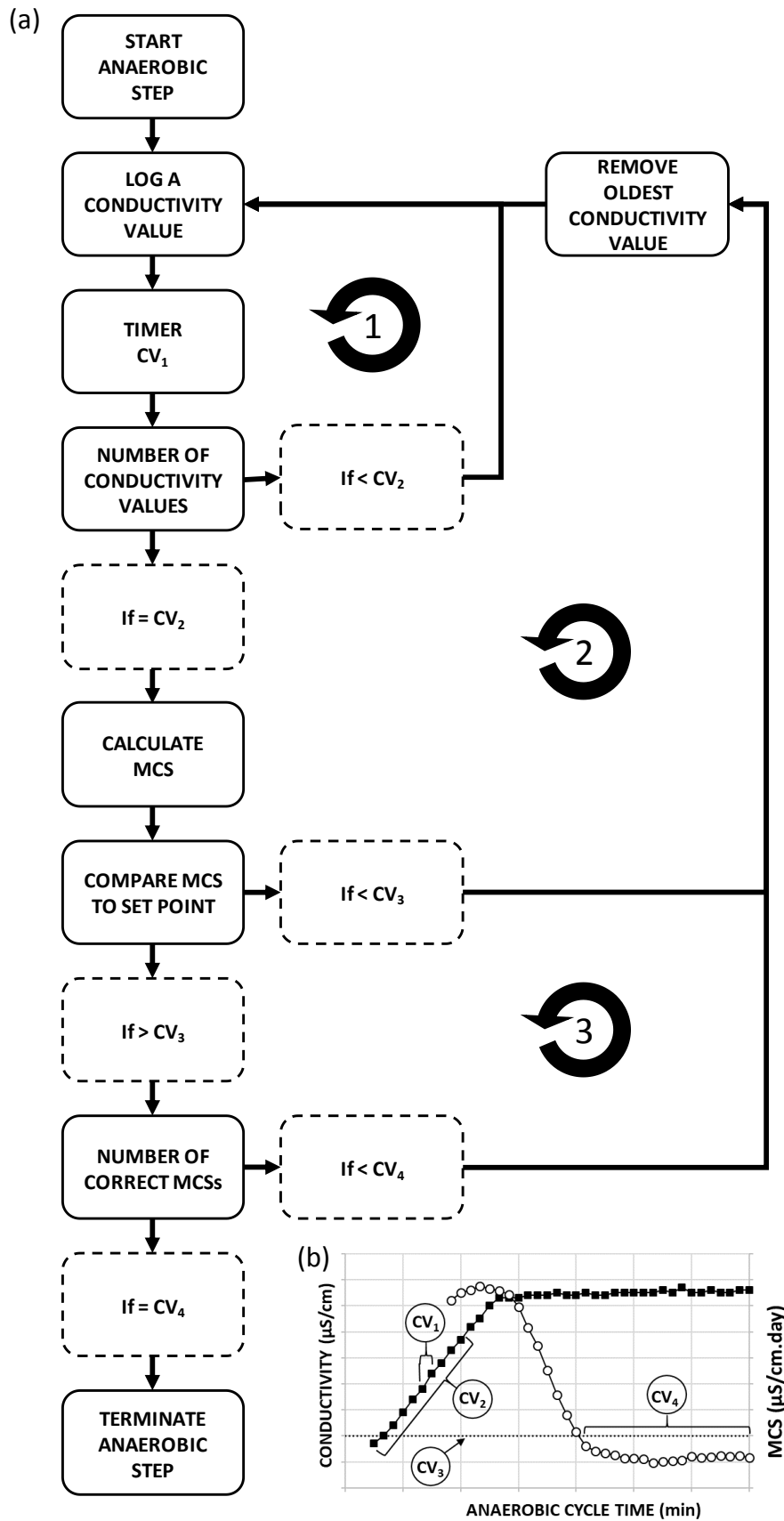
560 Figure 4: qPCR and microbial community composition quantification of  
561 *Candidatus Accumulibacter Phosphatis*.

562

563 Figure 5: Data phase 2, reactor B; (a) Influent COD and PO<sub>4</sub>-P concentration  
564 during the different IVSs; (b) Anaerobic and aerobic OLR during the different  
565 IVSs; (c) Anaerobic TOC uptake and PO<sub>4</sub>-P release efficiency at grab sample  
566 point SPb and SPc; (d) Anaerobic and aerobic time during the different IVSs.

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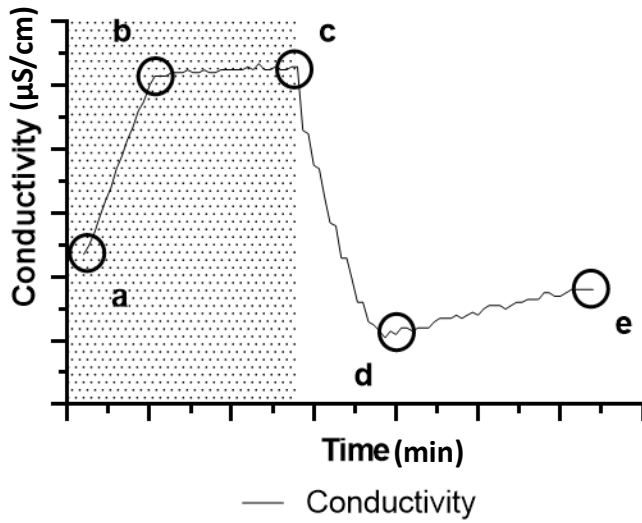
569 FIGURE 1



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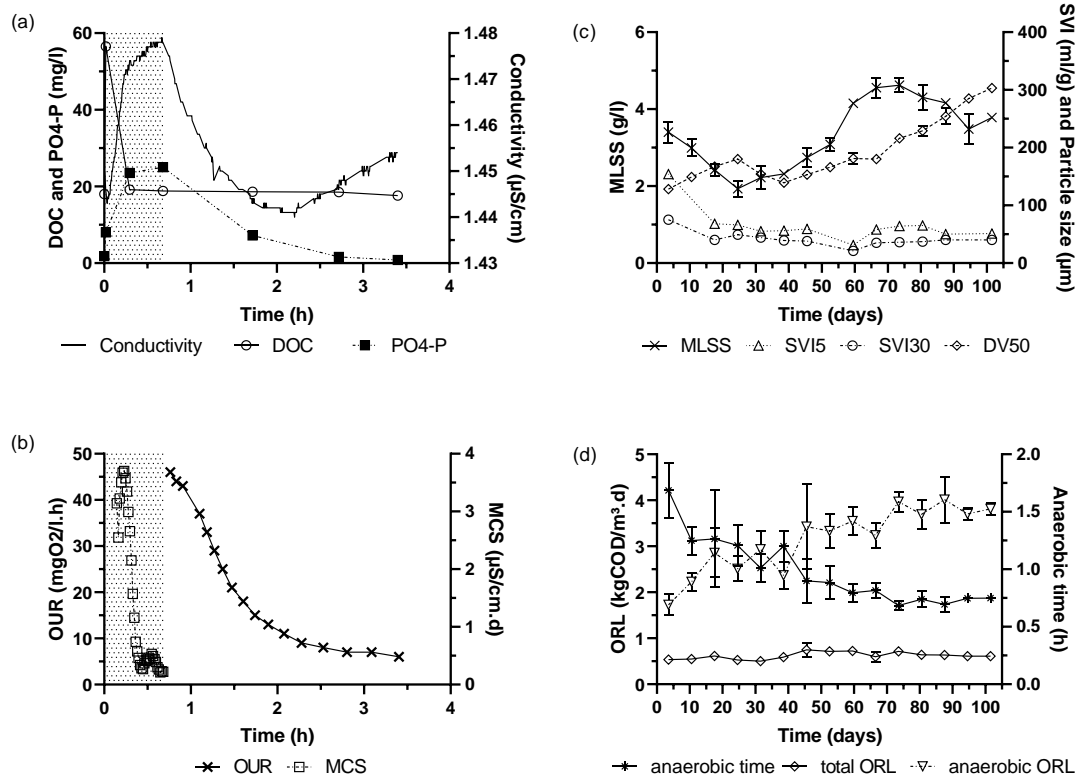
571 FIGURE 2

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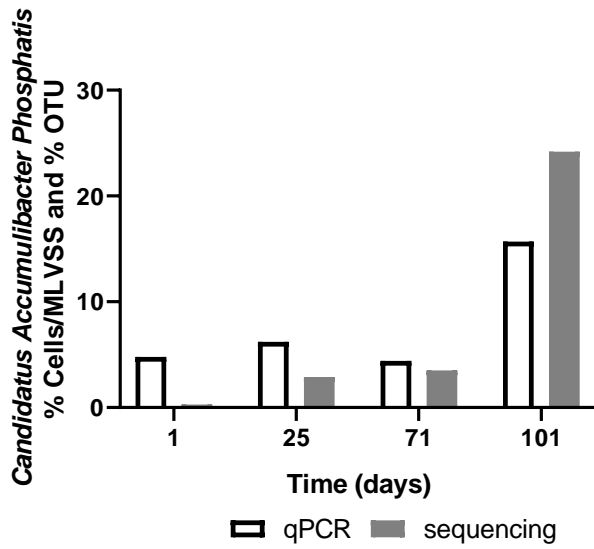
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574 FIGURE 3



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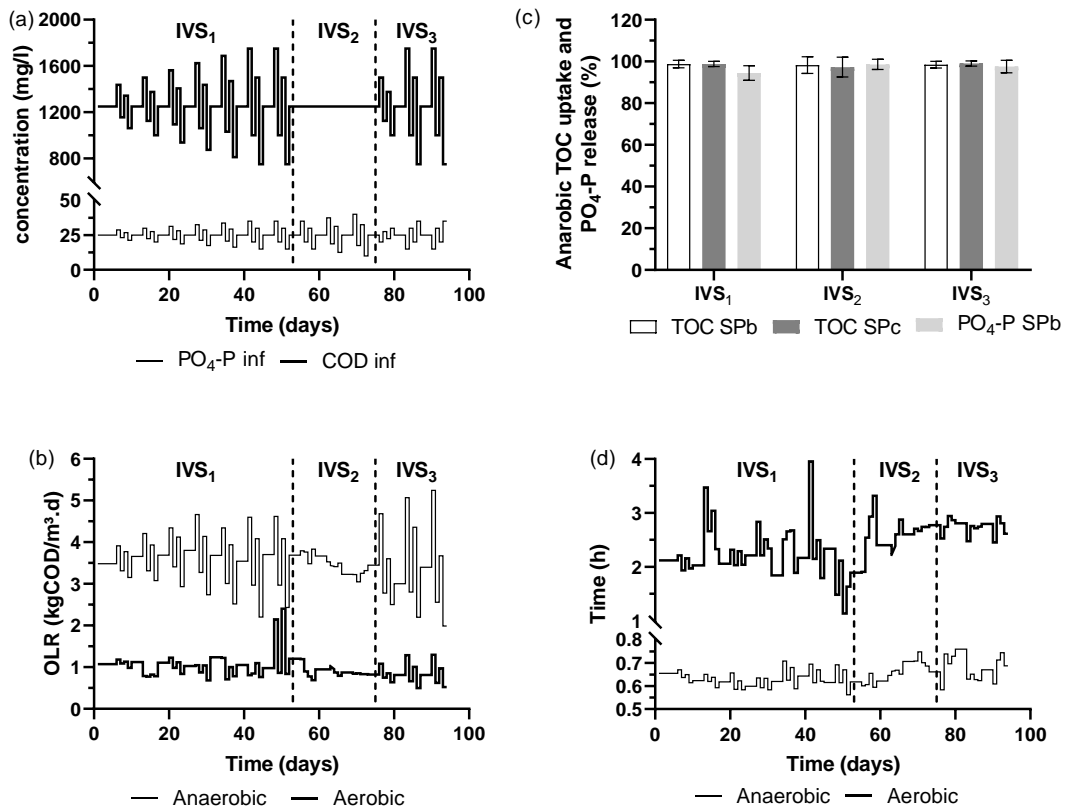
577 FIGURE 4



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580 FIGURE 5



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**Supplementary Interactive Plot Data (CSV)**

[Click here to download Supplementary Interactive Plot Data \(CSV\): SUPPLEMENTARY DATA.docx](#)