

Piliation of *Lactobacillus rhamnosus* GG Promotes Adhesion, Phagocytosis, and Cytokine Modulation in Macrophages

Cynthia E. Vargas García,^{a,b} Mariya Petrova,^{a,b} Ingmar J. J. Claes,^{a,b} Ilke De Boeck,^{a,b} Tine L. A. Verhoeven,^a Ellen Dilissen,^c Ingemar von Ossowski,^d Airi Palva,^d Dominique M. Bullens,^c Jos Vanderleyden,^a Sarah Lebeer^{a,b}

KU Leuven, Centre of Microbial and Plant Genetics, Heverlee, Belgium^a; University of Antwerp, Department of Bioscience Engineering, Research Group Environmental Ecology and Applied Microbiology, Antwerp, Belgium^b; KU Leuven, Laboratory of Pediatric Immunology, Leuven, Belgium^c; University of Helsinki, Department of Veterinary Biosciences, Helsinki, Finland^d

Recently, *spaCBA*-encoded pili on the cell surface of *Lactobacillus rhamnosus* GG were identified to be key molecules for binding to human intestinal mucus and Caco-2 intestinal epithelial cells. Here, we investigated the role of the SpaCBA pilus of *L. rhamnosus* GG in the interaction with macrophages *in vitro* by comparing the wild type with surface mutants. Our results show that SpaCBA pili play a significant role in the capacity for adhesion to macrophages and also promote bacterial uptake by these phagocytic cells. Interestingly, our data suggest that SpaCBA pili also mediate anti-inflammatory effects by induction of interleukin-10 (IL-10) mRNA and reduction of interleukin-6 (IL-6) mRNA in a murine RAW 264.7 macrophage cell line. These pili appear to mediate these effects indirectly by promoting close contact with the macrophages, facilitating the exertion of anti-inflammatory effects by other surface molecules via yet unknown mechanisms. Blockage of complement receptor 3 (CR3), previously identified to be a receptor for streptococcal pili, significantly decreased the uptake of pilus-expressing strains in RAW 264.7 cells, while the expression of IL-10 and IL-6 mRNA by these macrophages was not affected by this blocking. On the other hand, blockage of Toll-like receptor 2 (TLR2) significantly reduced the expression of IL-6 mRNA irrespective of the presence of pili.

Lactobacillus bacteria belong to the lactic acid bacteria (LAB) and are applied worldwide in the production of a variety of fermented food products. Besides their role in the food industry, some *Lactobacillus* species are naturally found throughout the human gastrointestinal tract (GIT) in different proportions, with the highest numbers being found in the proximal small intestine (1), yet their most important niche in the human body is probably within the vaginal cavity (2). Several lactobacilli have a documented probiotic function by providing health-promoting effects in humans upon administration. Among them, the well-studied probiotic *Lactobacillus rhamnosus* GG has been shown to prevent atopic diseases (3–5), decrease the risk of respiratory tract infections (6–8), prevent nosocomial gastrointestinal infections (7), prevent antibiotic-associated diarrhea (9), treat acute infectious diarrhea (10, 11), and be beneficial in the treatment of rotavirus-associated diarrhea (12), although there are always responders and nonresponders. For a more targeted approach, better knowledge of the underlying mechanisms is important. Because of its wide clinical use, *L. rhamnosus* GG is an important model probiotic strain for use in such targeted approaches.

During the last decade, incredible numbers of studies have focused on the possible immunomodulatory effects of probiotics. Probiotic bacteria are thought to confer such immunomodulatory effects by the microbe-associated molecular patterns (MAMPs) that are present on their cell surface or that are secreted in the environment by them (13). These MAMPs can be highly conserved, such as certain peptidoglycan motifs, or can be more strain specific, such as unique glyco- or lipoproteins. The interaction of a MAMP with its pattern recognition receptor (PRR) on host cells, such as Toll-like receptors (TLRs), results in the induction of a signaling cascade that modulates the expression of various response genes, such as cytokines and defensins (14). Increasing numbers of MAMPs that participate in the host interaction have been identified on the surface of probiotic lactobacilli (13, 15, 16).

For instance, lipoteichoic acid (LTA) from *L. rhamnosus* GG was demonstrated to interact with the PRRs TLR2/6 (17), the S-layer protein SlpA in *Lactobacillus acidophilus* NCFM interacts with the dendritic cell (DC) receptor DC-SIGN (18), and specific peptidoglycan muropeptides in *Lactobacillus salivarius* Ls33 are recognized by NOD2 (19).

Recently, pili or fimbriae (i.e., proteinaceous heteropolymeric extracellular appendages) were discovered at the cell surface of *L. rhamnosus* GG (20). These pili are encoded by the *spaCBA* gene cluster (LGG_00441 to LGG_00444). The SpaC subunit, located at the tip and along the backbone structure, was shown to be responsible for the high mucus-binding capacity (21). A comparison of the retention times of exogenously applied piliated *L. rhamnosus* GG and the related nonpiliated strain *L. rhamnosus* Lc705 also suggests a key role for the pili in promoting residence in the human colon, since piliated *L. rhamnosus* GG could be detected for 7 days longer in fecal samples from healthy volunteers (21). By mutant analysis of the *spaCBA* genes, we could show that the loss of pili results in the loss of adhesion to the microvillus-containing Caco-2 intestinal epithelial cell (IEC) line and increased induction

Received 3 December 2014 Accepted 5 January 2015

Accepted manuscript posted online 9 January 2015

Citation Vargas García CE, Petrova M, Claes IJ, De Boeck I, Verhoeven TLA, Dilissen E, von Ossowski I, Palva A, Bullens DM, Vanderleyden J, Lebeer S. 2015. Piliation of *Lactobacillus rhamnosus* GG promotes adhesion, phagocytosis, and cytokine modulation in macrophages. *Appl Environ Microbiol* 81:2050–2062. doi:10.1128/AEM.03949-14.

Editor: G. T. Macfarlane

Address correspondence to Sarah Lebeer, sarah.lebeer@uantwerpen.be.

Copyright © 2015, American Society for Microbiology. All Rights Reserved.

doi:10.1128/AEM.03949-14

TABLE 1 Bacterial strains and plasmids

Strain or plasmid	Relevant genotype and/or phenotype	Reference or source
Strains		
<i>Lactobacillus rhamnosus</i>		
GG (ATCC 53103)	Wild type; human isolate	21
CMPG5357	$\Delta spaCBA::Tc^r$; mutant of <i>L. rhamnosus</i> GG without <i>spaCBA</i> -encoded pili	22
CMPG5351	$\Delta welE::Tc^r$; mutant of <i>L. rhamnosus</i> GG without long galactose-rich exopolysaccharides but with increased exposure of pili	20
CMPG1908	CMPG5357 with chromosomal insertion of pMEC10 in the <i>attB</i> sequence of phage mv4; Ery ^r	This work
CMPG1907	CMPG5351 with chromosomal insertion of pMEC10 in the <i>attB</i> sequence of phage mv4; Ery ^r	This work
FAJ1906	<i>L. rhamnosus</i> GG with chromosomal insertion of pFAJ1934 in the <i>attB</i> sequence of phage A2; Ery ^r	33
<i>Lactococcus lactis</i> subsp. <i>cremoris</i>		
MG1363	Mesophilic lactic acid bacterium isolated from green plants	30
GRS1185	<i>L. lactis</i> NZ9000 containing the nisin-inducible vector pKTH5080, which in turn contains the coding region of the <i>spaCBA</i> pilus operon; Cm ^r	23
Plasmids		
pFAJ1934	2,634-bp PCR fragment (flanking EcoRI sites) carrying the <i>nisRK</i> genes of pMEC10 cloned into the EcoRI site of pEM40 downstream of the <i>ery</i> gene in the opposite orientation; Ery ^r Amp ^r	33
pMEC10	Integration plasmid (<i>attB</i> located at the 3' end of the tRNA ^{Ser} locus) containing the <i>int-attP</i> cassette (of phage mv4) of pMC1 cloned into pNZ9500 (a pUC19 derivative carrying a 2.7-kb chromosomal DNA fragment of <i>L. lactis</i> NZ9700 containing the 3' end of <i>nisP</i> and <i>nisRK</i>); expression driven by <i>ery</i> read-through; Ery ^r	34
pMEC45	<i>L. lactis</i> pSH71 replicon; pNZ8037 derivative with <i>gfp_{uv}</i> ^a cloned downstream of the <i>nisA</i> promoter from <i>L. lactis</i> NZ9800; Cm ^r	36

^a *gfp_{uv}*, synthetic GFP gene (with improved codon usage) that was subsequently subjected to recurrent cycles of DNA shuffling, thereby selecting for bright fluorescence that can be observed under UV light.

of the proinflammatory marker interleukin-8 (IL-8) in these nonphagocytic cells (22). Using recombinant lactococcal constructs, von Ossowski et al. (23) demonstrated that the SpaCBA pilus is a contributory factor in the activation of TLR2-dependent signaling in HEK cells by the recombinant lactococci as well as in the modulation of pro- and anti-inflammatory cytokine (tumor necrosis factor alpha, IL-6, IL-10, and IL-12) production in human monocyte-derived DCs (moDCs). However, a possible immunomodulatory role of the native pili present on the surface of live *L. rhamnosus* GG has not yet been explored. Interestingly, in the case of the pathogen *Streptococcus pneumoniae*, interaction of the pilus-associated RrgA adhesin with the complement receptor 3 (CR3), also named integrin CD11b/CD18 or Mac-1, was shown to enhance uptake of the bacteria by murine and human macrophages, on the basis of the results of experiments with a blocking antibody on the wild type and the isogenic mutant lacking RrgA (24). In addition, TLR2 was shown to mediate inflammatory responses to oligomerized RrgA (25).

In this study, we aimed to investigate the role of SpaCBA pili from the probiotic strain *L. rhamnosus* GG in the interaction with phagocytic innate immune cells. Intestine-resident DCs and macrophages are eager phagocytes capable of sampling bacteria in the intestinal lumen through the extension of transepithelial dendrites (26–28) and are important host cells for probiotic immunomodulatory applications. Here, we chose a simplified model with a murine macrophage-like (RAW 264.7) cell line. Cells of this cell line were coinoculated with the *L. rhamnosus* GG wild-type strain and different pilus mutants of *L. rhamnosus* GG, so that the role of pili in their natural conformation and organization on live cells could be studied. Such *in situ* cellular experiments are important because host cellular responses against bacteria are always an integrated sum of the interactions of different MAMPs with dif-

ferent PRRs (13). In addition, we previously showed the importance of the zipper-like and nanospring properties of the pili in promoting close contacts of whole *L. rhamnosus* GG cells with substrates (29), which cannot be mimicked with isolated pili without their cellular anchor.

MATERIALS AND METHODS

Bacterial strains and growth conditions. Wild-type *L. rhamnosus* GG and its mutant derivatives (Table 1) were routinely grown under non-shaking conditions in Man-Rogosa-Sharpe (MRS) medium (catalog number 288130; Difco, Becton, Dickinson, Erembodegem, Belgium) for 24 h at 37°C, and 1% of this preculture was inoculated into fresh MRS broth and grown overnight at 37°C. Bacteria were washed once with 1× phosphate-buffered saline (PBS; catalog number 14040-091; Invitrogen, Life Technologies, Ghent, Belgium), pH 7.4, and brought to a final concentration of 5 × 10⁷ CFU/ml prior to incubation with macrophages. This concentration was selected on the basis of findings of preliminary tests in which different dose-response relationships (1:1, 1:10, 1:50, and 1:100 cells/bacteria) were evaluated.

Wild-type strain *Lactococcus lactis* subsp. *cremoris* MG1363 (30) and the recombinant *L. lactis* subsp. *cremoris* clone GRS1185, itself having a nisin-inducible vector that in turn contains the coding region of the *spaCBA* pilus operon, were grown overnight in M17 broth (catalog number 218561; Becton, Dickinson, Erembodegem, Belgium) supplemented with 0.5% glucose at 30°C. A 1% aliquot taken from this preculture was used to further inoculate fresh M17 broth, and cells were kept overnight at 30°C. In the case of the recombinant lactococcal construct GRS1185, a similar procedure was followed, and the media were supplemented with 7.5 µg/ml chloramphenicol (catalog number C0378-25G; Sigma-Aldrich, St. Louis, MO, USA). After inoculation with 100 µl of preculture, the cultures were allowed to grow until the optical density at 600 nm (OD₆₀₀) was 0.5. Subsequently, 3 ng/ml of nisin was added to the cultures to induce pilus expression. Nisin (catalog number N5764; Sigma-Aldrich, St. Louis, MO, USA) was prepared as previously described (31). Briefly, a stock

solution of nisin (10 mg/ml) was made by suspending 100 mg of nisin per ml in 0.05% acetic acid, followed by 10-fold dilution with dimethyl sulfoxide.

Detection of SpaCBA pili on the *L. rhamnosus* GG wild type and mutants by immunofluorescence microscopy. To phenotypically characterize the derivative mutants of *L. rhamnosus* GG, i.e., *spaCBA* mutant CMPG5357 and the exopolysaccharide (EPS)-deficient mutant CMPG5351, immunofluorescence assays were performed as described previously (32). Briefly, cells were grown to exponential phase and fixed for 1 h in 2.5% formaldehyde (catalog number VELC1374.1000; VWR International, Radnor, PA, USA) and 0.05% glutaraldehyde (catalog number G5882-50ML; Sigma-Aldrich, St. Louis, MO, USA). Cells were then washed and incubated in blocking buffer (0.5%; Boehringer Mannheim, Mannheim, Germany) for 10 min. The samples were further incubated in blocking buffer containing anti-SpaC rabbit polyclonal antiserum (21) or preimmune serum (1:100 and 1:400 dilutions, respectively) for 30 min and subsequently incubated in blocking buffer containing goat anti-rabbit IgG (1:200 dilution) conjugated to Alexa Fluor 488 (catalog number A-11070; Invitrogen, Life Technologies, Ghent, Belgium) for another 30 min. Samples were visualized with a Zeiss Axio Imager Z1 microscope equipped with an AxioCam MRm Rev 3 monochrome digital camera.

GFP labeling of *L. rhamnosus* GG pilus mutants. The *nisRK* genes with their own promoter were integrated into the chromosome of the *L. rhamnosus* GG mutants, as previously described for the wild type (33). Site-specific integration was accomplished by electroporating the suicide plasmid pMEC10 (34) into the *L. rhamnosus* GG mutants. Integration of pMEC10 at the *tRNA^{Ser}* locus was confirmed by PCR as previously described (35). The following primers were used: PRO-567 (5'-GTCGACA CAGGATTTGAACC-3'; which corresponds to primer T2 [35]) and PRO-570 (5'-CAAGCCAACAGACGTGCAAGCA-3'; which binds to the 3' end of the *int* gene [35]). Subsequently, pMEC45 (36), the expression vector containing *gfp* under the control of the *nisA* promoter, was electroporated into CMPG5357/pMEC10 (CMPG1908) and CMPG5351/pMEC10 (CMPG1907). Transformants were verified by PCR analysis. Finally, the expression of green fluorescent protein (GFP) driven by the *nisA* promoter (pMEC45) in the *L. rhamnosus* GG derivatives was optimally induced as described previously (33). Briefly, overnight cultures of *L. rhamnosus* GG FAJ1906/pMEC45, CMPG1908/pMEC45, and CMPG1907/pMEC45 were used to inoculate fresh prewarmed MRS medium diluted 1:50. After 30 min of incubation at 37°C without shaking, 500 ng/ml nisin was added to the cultures, which were then incubated for 2.5 to 3 h. Nisin was used to induce the transcription of the genes under the control of the *nisA* promoter.

Maintenance and culturing of cell lines. The murine macrophage cell line RAW 264.7 (ATCC TIB-71) was purchased from the American Type Culture Collection (ATCC; Manassas, VA, USA). RAW 264.7 cells were maintained in Dulbecco's modified Eagle medium (DMEM; catalog number 41966-029; Life Technologies, Ghent, Belgium) containing 10% fetal bovine serum (FBS; catalog number SH30071.03; HyClone, USA) and 50 µg/ml gentamicin (catalog number G1397; Sigma-Aldrich, St. Louis, MO, USA) in a 5% CO₂ humidified incubator at 37°C. Every 2 or 3 days, the cells were split at a subcultivation ratio of 1:3. For adhesion, phagocytosis, and immunomodulation experiments, RAW 264.7 cells were grown overnight in 12-well culture plates (catalog number 665180; Greiner Bio-One, Ltd., Solingen, Germany) at a concentration of 1×10^6 cells/ml.

Adhesion and phagocytosis assays with murine macrophages. Experiments to assess the adhesion of various *L. rhamnosus* GG strains to RAW 264.7 cells were carried out as follows on the basis of the methods of Lebeer et al. (22). A 1-ml volume of *L. rhamnosus* GG and the corresponding mutants (5×10^7 CFU/ml) was added to tissue culture plate wells containing RAW 264.7 cells. The bacteria were allowed to adhere to the cells for 1 h at 37°C. The cells were then rinsed three times with $1 \times$ PBS prewarmed to 37°C and resuspended in 1 ml of $1 \times$ PBS, pH 7.4. Finally, the cells were detached using a cell scraper, and serial dilutions of bacterial

suspensions were prepared and plated out on solid MRS medium for *Lactobacillus* strains and M17 medium for *Lactococcus* strains. The percentage of bacterial adhesion was calculated by comparing the total number of bacterial colonies counted after adhesion to the number of cells in the bacterial suspension originally added to the tissue culture plate wells. In addition, the total amount of macrophage-associated bacteria was determined. To this point the aforementioned protocol for adhesion had been followed, but macrophages were then lysed with 0.2% Triton X-100 (catalog number T8787; Sigma-Aldrich, St. Louis, MO, USA) for 15 min prior to preparation of the dilution series in order to liberate internalized bacteria. The percentage of total associated bacteria was then calculated by comparing the total number of bacterial colonies counted after the macrophages were lysed to the number of bacteria originally added. Furthermore, to determine the number of internalized bacteria, phagocytosis assays were performed as described previously (37), with slight modifications. Briefly, RAW 264.7 cells (1×10^6 cells) were incubated for 1 h at 37°C with *L. rhamnosus* GG or *L. lactis* subsp. *cremoris* pilus variants as described above. Bacterial internalization was stopped with the addition of ice-cold $1 \times$ PBS, pH 7.4, and macrophages were washed three times to remove nonadhered bacteria. The remaining extracellular bacteria were killed by addition of streptomycin (100 µg/ml) and penicillin (100 U/ml) for another hour. The cells were then washed three times with $1 \times$ PBS and lysed with 0.2% Triton X-100 for 15 min. Serial dilutions of bacterial suspensions were prepared and plated out on solid media. The percentage of internalized bacteria was determined by comparing the total number of bacterial colonies counted after the cells were lysed to the number of bacteria originally added. All experiments were repeated at least three times, and all strains were tested in triplicate.

Analysis of bacterium-macrophage interaction by epifluorescence microscopy and flow cytometry. Epifluorescence microscopy was used to visualize the pattern of adhesion of the *L. rhamnosus* GG strains to RAW 264.7 cells. Macrophages were seeded onto coverslips in 6-well tissue culture plates (catalog number 657160; Greiner Bio-One, Ltd., Solingen, Germany) at a concentration of 1×10^6 cells per well and were grown overnight. An overnight culture of GFP-expressing (GFP⁺) *L. rhamnosus* GG (nisin-induced cultures of FAJ1906/pMEC45, CMPG1908/pMEC45, and CMPG1907/pMEC45) was used to inoculate fresh, prewarmed MRS medium diluted 1:50. After 30 min of incubation at 37°C without shaking, 500 ng/ml nisin was added to the culture, which was then incubated for 2.5 to 3 h. Fluorescently labeled *L. rhamnosus* GG strains were subsequently added to the macrophages at a ratio of 50 to 1 CFU/cell. After 2 h of incubation at 37°C in 5% CO₂, the cells were washed three times with $1 \times$ PBS, pH 7.4, to remove nonadhered bacteria, and the interaction between the bacteria and the macrophages was examined by epifluorescence microscopy using a Zeiss Axio Imager Z1 microscope equipped with an AxioCam MRm Rev 3 monochrome digital camera (Carl Zeiss Microimaging GmbH, Germany). This assay was repeated at least five times.

For flow cytometric analysis, GFP-expressing *L. rhamnosus* GG strains were incubated with macrophages for 2 h at 37°C in 5% CO₂. The cells were then fixed with 3% paraformaldehyde and resuspended in $1 \times$ PBS, and the fluorescence in the macrophages was examined by fluorescence-activated cell sorter (FACS) analysis with a BD Influx cell sorter (Becton, Dickinson, Erembodegem, Belgium). Data were analyzed using FlowJo software, version 7.6.4. The experiment was repeated at least three times, and all strains were tested in triplicate.

Induction of cytokine gene expression in RAW 264.7 cells. Wild-type and mutant *L. rhamnosus* GG cells were grown overnight in MRS medium and then centrifuged at $2,000 \times g$ for 15 min at 4°C. After washing with $1 \times$ PBS, pH 7.4, cells were resuspended in DMEM without FBS and adjusted to a final concentration of 5×10^7 CFU/ml. One milliliter of the bacterial suspension was added to the wells containing RAW 264.7 cells and incubated for 3 h at 37°C in 5% CO₂ at 90% humidity. Then, the cells were rinsed three times with prewarmed $1 \times$ PBS, pH 7.4. Here, lipopolysaccharide (LPS; 1 µg/ml) from *Escherichia coli* O111:B4 (catalog number L4391; Sigma-Aldrich, St. Louis, MO, USA) was included as a positive

TABLE 2 Primers and probes sequences used in this study for mRNA measurements^a

Target mRNA	Primer or probe	Sequence (5'–3')
TBP	FW	TTGTACCGCAGCTTCAAAATATTGT
	RV	GCAGCAAATCGCTTGGGAT
	TP	FAM-TTGACCTAAAGACCATTGCACTTCGTGCA-TAMRA
B2M	FW	CCACTGAGACTGATACATACGCCT
	RV	GATCACATGTCTCGATCCAGTAG
	TP	FAM-TAAGCATGCCAGTATGGCCGAGCC-TAMRA
SDHA	FW	ACATCAGAACTACGCCTAAACATGC
	RV	AAACCATTCCCCTGTGCAATG
	TP	FAM-AACCATGTGTCAGTGTCCCGTGTGG-TAMRA
RPL13A	FW	GCGCCTCAAGGTGTTGGAT
	RV	CCCAGGTAAGCAAACCTTTCTGG
	TP	FAM-TGGTCCCTGCTGCTCTCAAGGTTGTT-TAMRA
HPRT	FW	TTATCAGACTGAAGAGCTACTGTAATGATC
	RV	TTACCAGTGTCAATTATATCTTCAACAATC
	TP	FAM-TGAGAGATCATCTCCACCAATAACTTTTATGTCCC-TAMRA
PPIA	FW	GCCGCGTCTCCTTCGAG
	RV	GTAAAGTCAACCACCTGGCAC
	TP	FAM-TTGCAGACAAAGTTCCAAAGACAGCAGAAA-TAMRA
β-Actin	FW	AGAGGGAAATCGTGCGTGAC
	RV	CAATAGTGATGACCTGGCCGT
	TP	FAM-CACTGCCGCATCCTCTTCTCCC-TAMRA
GAPDH	FW	TCACCACCATGGAGAAGGC
	RV	GCTAAGCAGTTGGTGGTGCA
	TP	FAM-ATGCCCCCATGTTTGTGATGGGTGT-TAMRA
IL-6	FW	GAGGATACCACTCCCAACAGACC
	RV	AAGTGCATCATCGTTGTCATACA
	TP	FAM-CAGAATTGCCATTGCACAACCTCTTTTCTCA-TAMRA
IL-10	FW	GGTTGCCAAGCCTTATCGGA
	RV	ACCTGCTCCAATGCCTTGCT
	TP	FAM-TGAGGCGCTGTCATCGATTCTCCC-TAMRA

^a TBP, TATAA box binding protein; B2M, β₂-microglobulin; SDHA, succinate dehydrogenase complex subunit A; RPL13A, ribosomal protein L13a; HPRT, hypoxanthine phosphoribosyltransferase; PPIA, peptidyl-prolyl *cis-trans*-isomerase A; GAPDH, glyceraldehyde-3-phosphate dehydrogenase; IL-6, interleukin-6; IL-10, interleukin-10; FW, forward primer; RV, reverse primer; TP, hydrolysis probe; FAM, 6-carboxyfluorescein; TAMRA, 6-carboxytetramethylrhodamine.

control, and DMEM unsupplemented with FBS was used as a negative control. Additionally, nonpiliated *L. lactis* subsp. *cremoris* MG1363 and the recombinant SpaCBA-piliated lactococcal construct (GRS1185) were also included as controls. RNA was extracted from the RAW 264.7 cells by using an RNeasy minikit (Qiagen, MD, USA), following the manufacturer's protocol. The concentration was determined by spectrophotometric measurement of the OD₂₆₀ (ND-1000; NanoDrop Technologies). In addition, OD₂₆₀/OD₂₈₀ and OD₂₆₀/OD₂₃₀ ratios were measured and found to range from 2.0 to 2.2, and the RNA quality and integrity were confirmed with an Experion RNA analysis kit (Bio-Rad Laboratories, Hercules, CA, USA). Values of the RNA integrity number (RIN) of between 9.8 and 10 were obtained, with a value of 10 indicating intact RNA (data not shown). Measurement of cytokine gene expression was determined by quantitative real-time PCR (qPCR) as described below. The experiment was repeated at least three times, and all strains were tested in triplicate. To determine whether direct cell-to-cell contact is required for cytokine induction, RAW 264.7 cells were incubated with the *L. rhamnosus* GG strains in a Transwell culture system (catalog number CLS3401-48EA; Sigma-Aldrich, St. Louis, MO, USA). RAW 264.7 cells were located in the lower chamber, and *L. rhamnosus* GG cells were located in the upper chamber. The cells were separated by a 0.4-μm-pore-size permeable filter membrane support, thereby avoiding any direct contact between bacterial cells and macrophages.

qPCR analysis. Isolated total RNA (1 μg) was transcribed to cDNA with SuperScript II reverse transcriptase (catalog number 18064-071; Life Technologies, Ghent, Belgium) and oligo(dT) primers (catalog number N8080128; Life Technologies, Ghent, Belgium) according to the manufacturer's protocol. Afterward, nuclease-free water (catalog number

AM9935; Life Technologies, Ghent, Belgium) was added to a volume of 100 μl. For each sample, 5 μl cDNA (containing 50 ng of transcribed RNA) was amplified in duplicate with the TaqMan universal PCR master mix (catalog number 4304437; Life Technologies, Ghent, Belgium) in a total reaction volume of 25 μl. We chose to evaluate eight reference genes commonly used as internal controls in expression studies, namely, TATAA box binding protein (TBP), succinate dehydrogenase complex subunit A (SDHA), glyceraldehyde-3-phosphate dehydrogenase (GAPDH), β₂-microglobulin (B2M), peptidyl-prolyl *cis-trans*-isomerase A (PPIA), ribosomal protein L13a (RPL13A), hypoxanthine phosphoribosyltransferase (HPRT), and β-actin. Interestingly, the TBP, SDHA, and GAPDH genes gave comparable results on the basis of RefFinder analyses (<http://www.leonxie.com/referencegene.php>) (data not shown). The GAPDH gene was then chosen as a reference gene, so that all results were normalized to those for the GAPDH gene to compensate for differences in the amount of starting material. qPCR was performed for IL-10, IL-6, and GAPDH in a StepOnePlus real-time PCR system (Applied Biosystems, Lennik, Belgium). cDNA plasmid standards, consisting of purified plasmid DNA specific for each cytokine, were used to quantify the target gene in the unknown samples, as described previously (38). For each cytokine, at least one primer or probe was designed to span an intron region of the matching cytokine gene. All primers and probes were designed on the basis of published sequences (39, 40) and chemically synthesized by Integrated DNA Technologies (IDT; Coralville, IA, USA) (Table 2). Each qPCR amplification was performed in duplicate wells in clear 96-well reaction plates (catalog number 4346906; Life Technologies, Ghent, Belgium) under the following conditions: 95°C for 10 min, followed by 40 cycles at 95°C for 15 s and 60°C for 1 min. qPCR data are presented as a

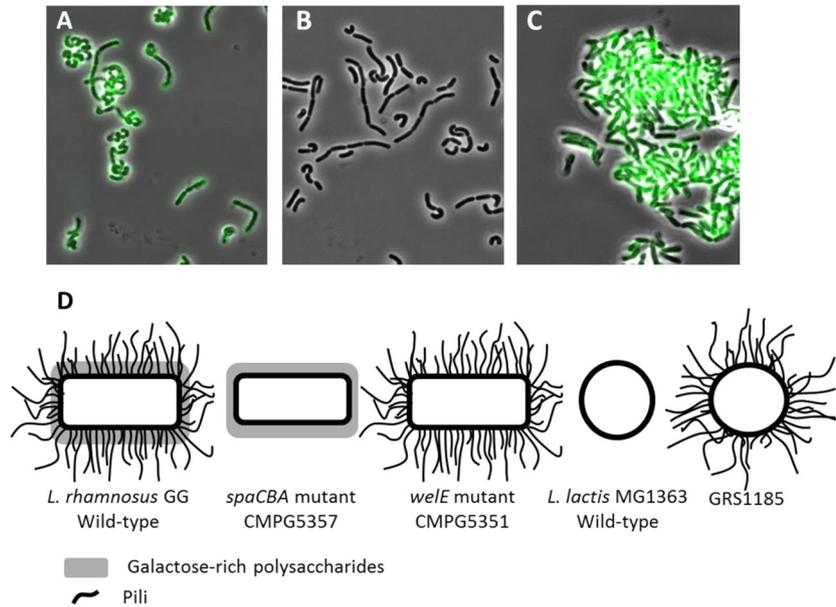


FIG 1 Presence of pili in *L. rhamnosus* GG and its derivative mutants. (A to C) Indirect immunofluorescence microscopy. Anti-SpaC rabbit antisera (1:100 dilution) were used on the *L. rhamnosus* GG wild type (A), pilus-deficient *spaCBA* mutant CMPG5357 (B), and EPS-deficient mutant CMPG5351 (C). SpaC-containing pili were then identified by anti-rabbit IgG antibodies (1:200 dilution) conjugated with Alexa Fluor 488. (D) Schematic representation of the bacterial strains used. The *L. rhamnosus* GG wild type, its derivative mutant strains, the *L. lactis* subsp. *cremoris* MG1363 wild type, and its recombinant SpaCBA-piliated lactococcal construct (GRS1185) were used in this study. The *L. rhamnosus* GG wild type has galactose-rich polysaccharides and pili on its surface. The *spaCBA* mutant CMPG5357 has galactose-rich polysaccharides but lacks the pili. The *welE* mutant CMPG5351 lacks the long galactose-rich polysaccharides but has increased exposure of pili and other surface molecules (20–23, 64–66).

ratio of the amount of cytokine mRNA to the amount of GAPDH mRNA. Nontemplate controls were included for each run.

TLR2 and CD11b blocking assay. The blocking monoclonal anti-human/mouse CD282 TLR2 purified antibody (clones T2.5 and 6C2) and monoclonal anti-mouse CD11b purified antibody (clone M1/70) were obtained from eBioscience (San Diego, CA, USA). RAW 264.7 cells (1×10^6 cells/ml) were incubated with TLR2 (5 to 10 $\mu\text{g/ml}$) or CD11b antibodies (5 $\mu\text{g/ml}$) for 30 min at 37°C in 5% CO₂. Cells were washed three times with serum-free culture medium to wash off any free antibody. Subsequently, cells were cocultured with bacteria (5×10^7 CFU/ml). After 3 h of incubation, the RAW 264.7 cells were rinsed three times with prewarmed 1× PBS, pH 7.4. RNA was extracted, and measurement of cytokine gene expression was determined by qPCR as described above. Bacterial adhesion to and uptake by macrophages pretreated with CD11b antibodies were also analyzed as described above.

Statistical analysis. Significant differences in the data between wild-type *L. rhamnosus* GG and its mutant derivatives were evaluated by the unpaired Student's *t* test. A *P* value of ≤ 0.05 was considered statistically significant. All data were also analyzed using SPSS Statistics (version 22) software. The normality of the distribution was assessed using the Shapiro-Wilk test in which significant values greater than 0.05 represented a normal distribution. Graphical tools were also used to test normality by analyzing histograms and quantile-quantile (Q-Q) plots of the sample data.

RESULTS

Detection of pili on the *L. rhamnosus* GG wild type and isogenic mutants by immunofluorescence microscopy. In addition to the *L. rhamnosus* GG wild type, the isogenic *spaCBA* mutant CMPG5357, which lacks the SpaCBA pili (22), and the *welE* mutant CMPG5351, which is deficient in the production of EPS but shows increased exposure of pili and glucose-rich polysaccharides (20), were the main pilus-expressing variants studied in this work

(Fig. 1D). To phenotypically confirm the presence of pili in these derivative mutants of *L. rhamnosus* GG, indirect immunofluorescence microscopy with anti-SpaC serum was performed. Figure 1B confirms that the pilus mutant CMPG5357 does not contain SpaC monomeric adhesins, as no fluorescence was detected on the mutant cells stained with anti-SpaC serum. In contrast, we were able to visualize SpaC-containing pili on the *L. rhamnosus* GG wild type (Fig. 1A) and even observed an increased signal with the SpaC antiserum in the EPS-deficient mutant CMPG5351 (Fig. 1C). Staining with preimmune serum as a negative control showed no fluorescence in the wild type or mutants (data not shown).

SpaCBA pili promote *L. rhamnosus* GG adhesion to RAW 264.7 cells and modulate its internalization. Since intestinal epithelial cells and monocytic cells have different cell surface properties, we subsequently aimed to investigate the role of pili in adherence to macrophages. Compared to the adhesion of the *L. rhamnosus* GG wild type, the pilus *spaCBA*-knockout mutant showed a ca. 2.3-fold reduction in adhesion to murine macrophage RAW 264.7 cells, while the EPS-deficient mutant CMPG5351 showed a ca. 2-fold increased adhesion capacity (Fig. 2). Likewise, an increase in the level of adhesion to RAW 264.7 cells of ca. 10-fold was observed for the nisin-induced GRS1185 lactococcal construct producing SpaCBA pili compared to the level observed for the nonpilated *L. lactis* subsp. *cremoris* wild type. To specifically assess the role of SpaCBA pili on macrophage binding, the total amount of macrophage-associated bacteria was also determined, and when RAW 264.7 cells were stimulated with the SpaCBA pilus-deficient mutant CMPG5357, a reduction in the total amount of bacteria of ca. 2.3-fold compared to the total

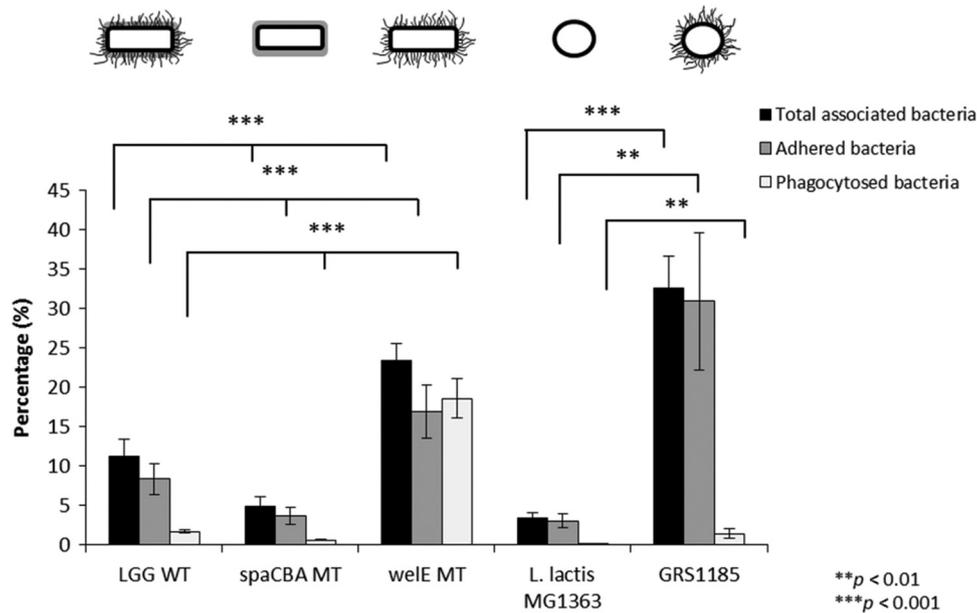


FIG 2 Adhesion to RAW 264.7 cells and phagocytosis of wild-type (WT) *L. rhamnosus* GG and its knockout mutant (MT) derivatives by RAW 264.7 cells. For adhesion assays, *L. rhamnosus* GG (5×10^7 CFU/ml) was coincubated for 1 h with 1×10^6 RAW 264.7 cells, and the percentage of adherent bacteria was determined. The total amount of macrophage-associated bacteria was also evaluated by lysis of the RAW 264.7 cells prior to preparation of the dilution series in order to liberate internalized bacteria. For phagocytosis assays, *L. rhamnosus* GG and its knockout mutant derivatives were added to RAW 264.7 cells at a ratio of 50:1 for 1 h. The remaining extracellular bacteria were killed by addition of streptomycin (100 μ g/ml) and penicillin (100 U/ml) for an additional hour, and the percentage of internalized bacteria was then determined. Nonpiliated *L. lactis* subsp. *cremoris* MG1363 and the recombinant SpaCBA-piliated lactococcal construct (GRS1185) were included as controls. These experiments were done in triplicate, and error bars represent standard deviations. Asterisks represent statistically significant differences compared to the results for the *L. rhamnosus* GG wild type or *L. lactis* subsp. *cremoris* MG1363.

amount found when they were stimulated with *L. rhamnosus* GG was found. Conversely, the EPS-deficient mutant CMPG5351 showed an increase in the total amount of associated bacteria of ca. 2.1-fold (Fig. 2). Similarly, we could observe an increase in the total number of macrophage-associated bacteria of ca. 9.6-fold for the recombinant SpaCBA-piliated lactococcal cells compared to that for the nonpiliated *L. lactis* subsp. *cremoris* wild type.

Furthermore, to examine whether SpaCBA pilus-mediated adhesion affects the uptake of *L. rhamnosus* GG by macrophages by phagocytosis and related uptake mechanisms, the *L. rhamnosus* GG wild type and its mutant derivatives were incubated with macrophages, and the number of internalized bacteria was counted. External bacteria were killed by antibiotic treatment consisting of a combination of streptomycin (100 μ g/ml) and penicillin (100 U/ml). Previous experiments showed that this treatment resulted in a more than 99% decrease of bacterial survival after 1 h of incubation, so that external bacteria do not significantly bias the results (data not shown). We subsequently observed that the SpaCBA-deficient mutant CMPG5357 was internalized to a ca. 2.7-fold lower degree than the *L. rhamnosus* GG wild type in RAW 264.7 cells. Conversely, the EPS-deficient mutant CMPG5351 showed a ca. 10.8-fold increased uptake compared to the wild type (Fig. 2). Similarly, the recombinant SpaCBA-piliated lactococcal construct GRS1185 was internalized by RAW264.7 cells ca. 452-fold more than the nonpiliated *L. lactis* subsp. *cremoris* wild type.

To further verify our findings, we also performed fluorescence microscopy analyses. First, the pilus mutant CMPG5357 and the EPS-deficient mutant CMPG5351 were transformed with a GFP-expressing plasmid as described in Materials and Methods. RAW 264.7 cells were then incubated with the GFP-expressing bacteria

for 2 h, after which the interaction between bacteria and macrophages was analyzed. Notably, in five independent experiments we could barely detect the pilus-knockout mutant (CMPG1908/pMEC45) in the sample (Fig. 3B), while the EPS-deficient mutant (CMPG1907/pMEC45) showed a stronger capacity to interact with RAW 264.7 cells (Fig. 3C) than the *L. rhamnosus* GG wild type (Fig. 3A). Similarly, our flow cytometric data showed that the *spaCBA* pilus mutant (CMPG1908/pMEC45) had a lower capacity to interact with macrophages than the wild type, while the EPS-deficient mutant with an increased exposure of pili (CMPG1907/pMEC45) interacted with RAW 264.7 cells more than the *L. rhamnosus* GG wild type, showing a biphasic interaction curve indicating that the interaction is not synchronized (Fig. 3D). Collectively our data indicate that SpaCBA pili indeed play a crucial role in the adhesion of *L. rhamnosus* GG to murine macrophages and are also involved in its internalization process, while galactose-rich EPS molecules seem to prevent the adhesion capacity and further phagocytosis of *L. rhamnosus* GG by macrophages.

Cytokine mRNA induction in RAW 264.7 cells is modulated by the presence of SpaCBA pili on *L. rhamnosus* GG. Subsequently, we investigated whether the SpaCBA pilus-mediated adhesion influences the induction of the pro- and anti-inflammatory cytokines IL-6 and IL-10, respectively, in RAW 264.7 cells. Macrophages were coincubated with bacteria for 3 h, on the basis of the findings of previous experiments in which different incubation times were tested, and 3 h was found to be the earliest time point with optimal expression of the mRNA of the cytokines under study (data not shown). Our qPCR analysis showed a ca. 4-fold increase in the expression of the anti-inflammatory cytokine IL-10 mRNA in macrophages stimulated with the EPS-defi-

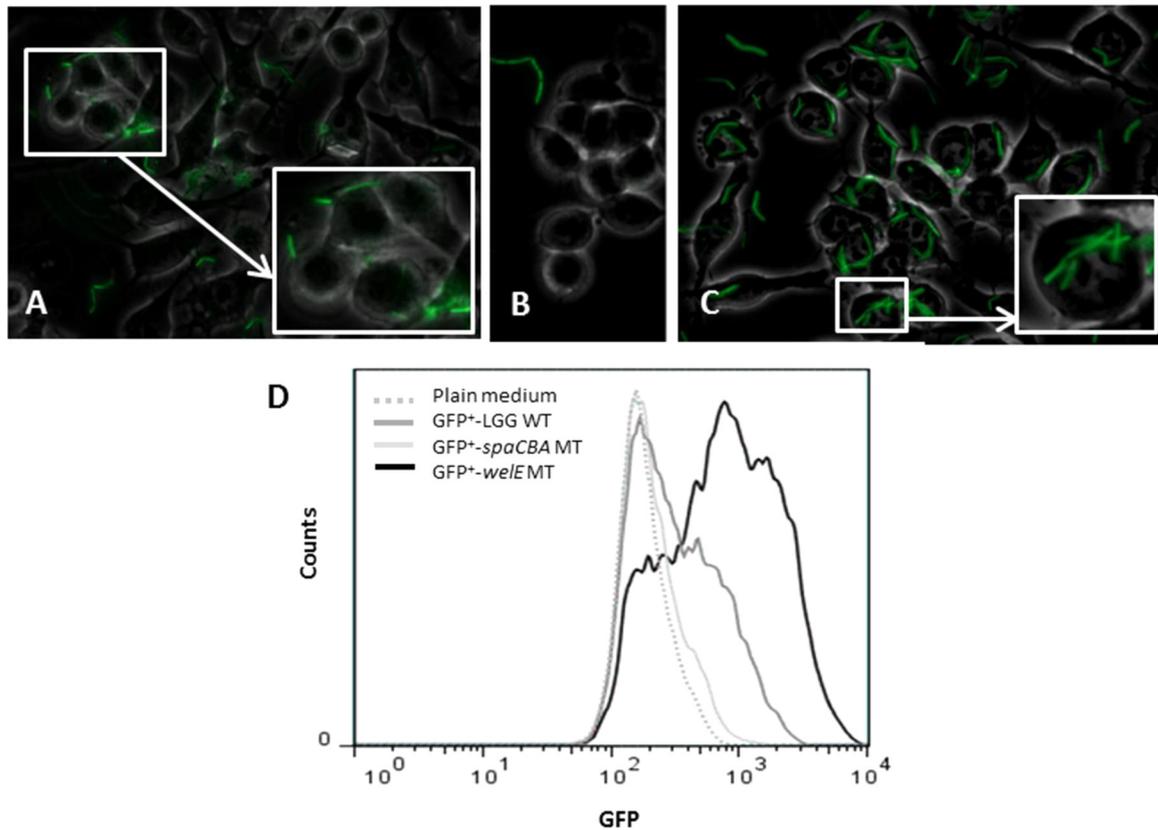


FIG 3 Interaction of GFP-expressing *L. rhamnosus* GG WT and its mutant derivatives with macrophages visualized by fluorescence microscopy and flow cytometry. (A to C) For fluorescence microscopy assays, RAW 264.7 cells were incubated with *L. rhamnosus* GG FAJ1906/pMEC45 (A), CMPG1908/pMEC45 (GFP⁺ *spaCBA* mutant) (B), and CMPG1907/pMEC45 (GFP⁺ *welE* mutant) (C) for 2 h. Fluorescence micrographs are representative of those from 5 independent experiments. (D) FACS analysis. RAW 264.7 cells were incubated with the GFP-expressing *L. rhamnosus* GG wild type and mutants for 2 h at 37°C. The graph depicts the signal intensity of GFP (previously calibrated with fluorescently labeled beads [rainbow beads, catalog number RCP-30-5A, 8 peaks; Spherotech]) (x axis) versus the number of cells (y axis). The results represent the fluorescence intensity of 5×10^5 cells in one representative experiment.

cient mutant CMPG5351 compared to that in macrophages stimulated with the *L. rhamnosus* GG wild type, whereas the pilus mutant CMPG5357 induced a similar level of IL-10 mRNA (Fig. 4A, left). We also analyzed the role of pili in a recombinantly engineered *L. lactis* subsp. *cremoris* strain that produces SpaCBA pili. Interestingly, GRS1185 cells producing SpaCBA pili induced the expression of IL-10 mRNA to levels ca. 4.5-fold greater than those for the nonpilated *L. lactis* subsp. *cremoris* wild type. Taken these findings together, the presence of pili seems to moderately promote IL-10 mRNA induction by *L. rhamnosus* GG and *L. lactis* subsp. *cremoris*, but comparisons of the results for the mutants show that the pili *per se* are not required for IL-10 mRNA induction. It is also noteworthy that the level of induction of the expression of IL-10 by *L. rhamnosus* GG or the *L. lactis* subsp. *cremoris* wild type is comparable to that by the control, LPS (1 μ g/ml) (Fig. 4A, left).

We also tested the expression of IL-6 mRNA as a proinflammatory cytokine, considering its relation with inflammation. As shown in Fig. 4A (right), the expression of IL-6 mRNA was slightly increased by ca. 1.2-fold when macrophages were stimulated with the *spaCBA* pilus mutant CMPG5357, while the enhanced exposure of pili did not appear to impact IL-6 mRNA induction significantly when the results obtained with EPS-deficient mutant CMPG5351 and the *L. rhamnosus* GG wild type were compared

($P < 0.05$). However, stimulation with GRS1185 producing SpaCBA pili resulted in a decreased induction of IL-6 mRNA in RAW 264.7 cells by ca. 2-fold compared to that achieved with stimulation by the nonpilated *L. lactis* subsp. *cremoris* wild type. Taken together, the presence of pili seems to moderately reduce IL-6 mRNA induction by *L. rhamnosus* GG and *L. lactis* subsp. *cremoris*. Importantly, LPS (1 μ g/ml) induced IL-6 mRNA to much higher levels than all strains tested.

Subsequently, in order to investigate the role of cell contact in *L. rhamnosus* GG-mediated cytokine modulation in macrophages, Transwell experiments were performed to prevent direct contact between macrophages and bacteria. IL-10 mRNA expression was examined after 3 h using qPCR. Separation of macrophages from bacteria by a 0.4- μ m-pore-size membrane indeed completely suppressed the induction of IL-10 mRNA (Fig. 4B, left). Similar results were found while investigating the expression of IL-6 mRNA (Fig. 4B, right).

Role of the host receptors CR3 and TLR2 in the interaction of *L. rhamnosus* GG with macrophages. To identify putative receptors on macrophages that are involved in the recognition of *L. rhamnosus* GG, possibly via the SpaCBA pili, we used previously validated blocking antibodies (24, 41) against complement receptor 3 (CR3; consisting of the two subunits CD11b and CD18) and Toll-like receptor 2 (TLR2). First, we evaluated the effect of CR3 in

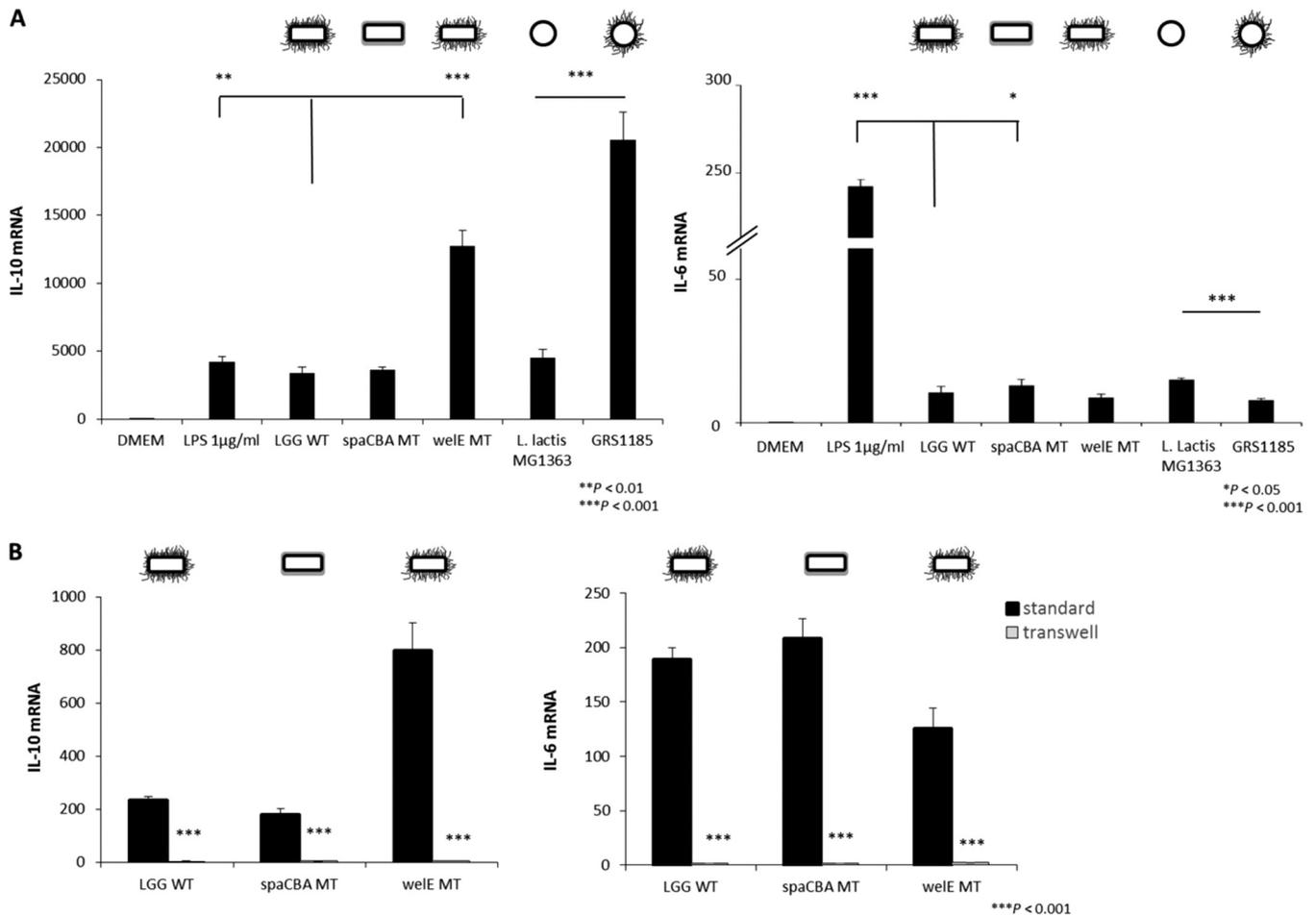


FIG 4 Interleukin-10 mRNA and interleukin-6 mRNA response of RAW 264.7 macrophages. (A) Cells were stimulated with *L. rhamnosus* GG strains for 3 h. DMEM served as a negative control, and LPS at 1 µg/ml served as a positive control. Nonpiliated *L. lactis* subsp. *cremoris* MG1363 and the recombinant SpaCBA-piliated lactococcal construct (GRS1185) were included as additional controls. Bacteria were administered at a ratio of 1:50 cells to bacteria. Results were quantified by the use of a cDNA plasmid standard and expressed as the ratio of the cDNA copy numbers for IL-10 (left) or IL-6 (right) divided by the cDNA copy numbers for GAPDH multiplied by 10^4 . Data are mean values \pm standard deviations of triplicate values and represent those from one of three independent experiments. Asterisks represent statistically significant differences compared to the results for the *L. rhamnosus* GG wild type or *L. lactis* subsp. *cremoris* MG1363. (B) The role of direct cell-cell contact on *L. rhamnosus* GG-induced expression of IL-10 (left) and IL-6 (right) mRNA by RAW 264.7 cells was measured by qPCR after 3 h of coinubation with the different bacterial strains. In a Transwell system, direct cell-cell contact was prevented, and macrophages were grown in the basolateral compartment with apical addition of the respective bacterial strains. Data are mean values \pm standard deviations of triplicate values and represent those from one of three independent experiments. Controls (no treatment) did not induce cytokine secretion.

SpaCBA pilus-mediated adhesion to RAW 264.7 cells by blocking its CD11b component. We observed no significant differences ($P < 0.05$) in the adhesion of the *L. rhamnosus* GG wild type, the pilus mutant CMPG5357 and the EPS-deficient mutant CMPG5351 to murine macrophages that had previously been treated with anti-CD11b antibodies (Fig. 5A, left). Even though the difference was not significant, a minor inhibition of the adhesion of the EPS-deficient mutant was found when RAW 264.7 cells were pretreated with the antibody. Furthermore, the level of macrophage ingestion of pilus-expressing strains was slightly but significantly inhibited by ca. 1.2-fold ($P < 0.01$) by anti-CD11b treatment (Fig. 5A, right) of both the *L. rhamnosus* GG wild type and the EPS-deficient mutant. However, when analyzing the role of CD11b blocking in the cytokine modulatory capacity of *L. rhamnosus* GG, we observed that the expression of IL-10 and IL-6 mRNA was not significantly affected by treatment with anti-CD11b antibodies (data not shown). In contrast, antibody block-

age of TLR2 showed a significant decrease in the expression of IL-6 mRNA by macrophages after stimulation with all the *L. rhamnosus* GG strains, while for IL-10 mRNA, this decrease was significant only for the EPS-deficient mutant ($P < 0.05$) (Fig. 5B).

DISCUSSION

Adherence is the first step by which components of the bacterial cell surface interact with receptors present on the host cells. Subsequently, surface-bound components and molecules secreted by the bacteria can trigger particular signaling pathways leading to the production of specific host proteins, resulting in specific immune responses. Thus, investigation of the molecular interactions between components on the bacterial cell surface and host cells is of great importance to unravel new mechanisms of regulation of the host immune responses (13). However, the role of bacterial adhesion in immunomodulation, especially in probiotic lactobacilli, remains unclear. Previous studies by our group and others

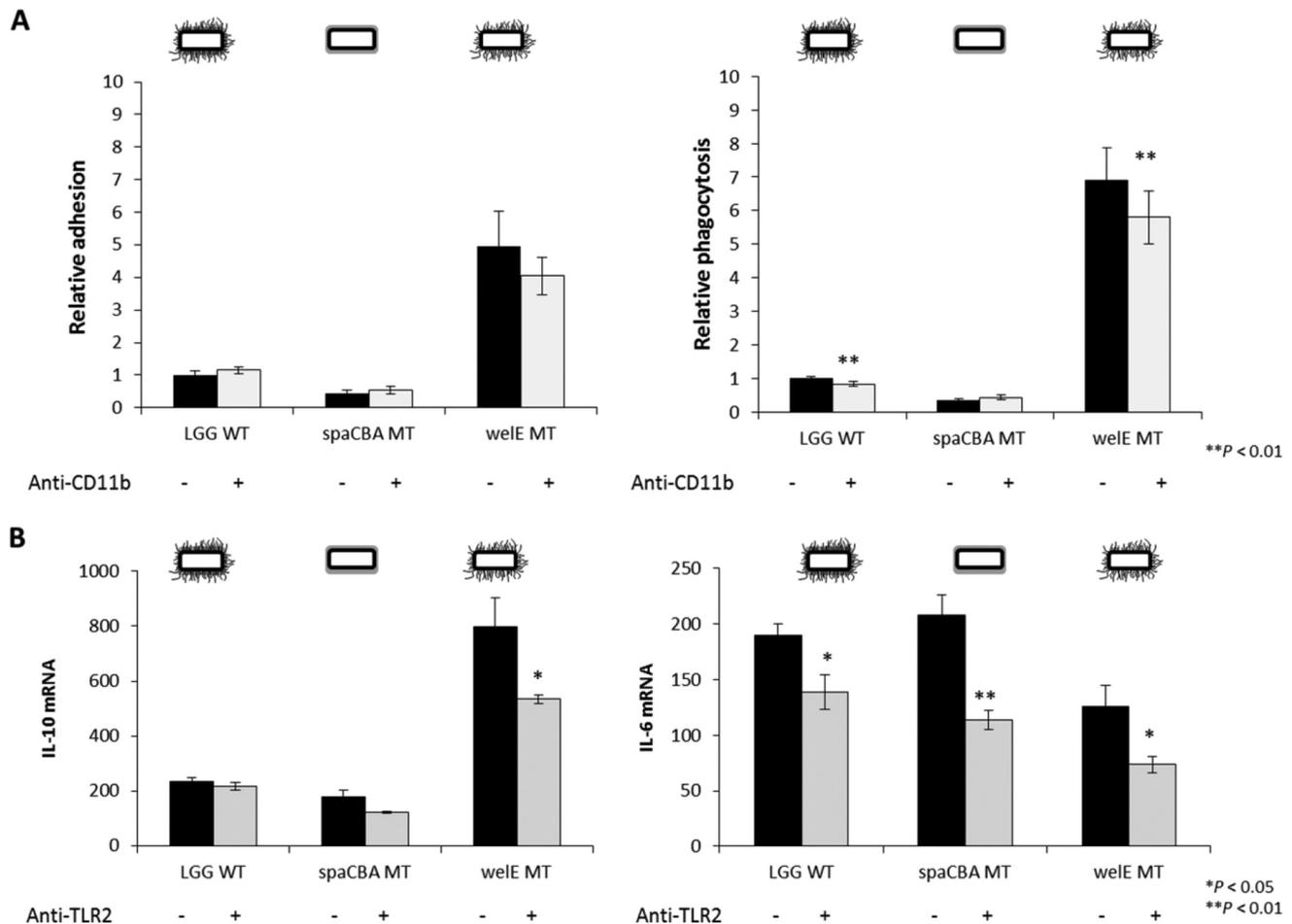


FIG 5 Role of CD11b in adhesion and uptake of *L. rhamnosus* GG by RAW 264.7 cells and effect of TLR2 in the subsequent immunomodulatory response. (A) Role of CD11b in adhesion of the mutants to macrophages (left) and phagocytosis of the mutants by macrophages (right) relative to the results for the wild-type strain, which were set equal to 1. Bacterial strains (5×10^7 CFU/ml) were coincubated for 1 h with 1×10^6 RAW 264.7 cells that had previously been treated with anti-mouse CD11b monoclonal antibody (5 μ g/ml) for 30 min. The data are presented as the mean relative adhesion or phagocytosis \pm standard deviation. These experiments were done in triplicate. Asterisks represent statistically significant differences compared to the results for the controls (no antibody treatment). (B) The effect of TLR2 on the relative levels of IL-10 and IL-6 mRNA expression by RAW 264.7 macrophages was analyzed by pretreatment of macrophages with antibodies against TLR2 (5 to 10 μ g/ml) for 30 min and then coincubation with *L. rhamnosus* GG strains for 3 h. Bacteria were administered at a ratio of 1:50 of cells to bacteria. The results are mean values \pm standard deviations from three separate experiments. The values are normalized against those for GAPDH. Asterisks denote statistically significant differences from the results for the control (no antibody treatment).

have indicated that there are large differences in the adherence capacities of lactobacilli (reviewed in, e.g., reference 42). We recently showed that the long flexible pili of *L. rhamnosus* GG promote a strong interaction with substrates by a zipper-like mechanism involving SpaC adhesin subunits present at the tip but also along the pilus fibers (29). In addition, these SpaCBA pili seem to show a remarkable elasticity imparting nanospring properties that might help them to retract closer to substrates even under conditions of shear flow or peristalsis (29). Thus, given its high adherence capacity mediated by pili, *L. rhamnosus* GG is an interesting model strain with which to investigate the interplay between adhesion and immunomodulation. Here, we aimed to explore the importance of the presence of these pili on native *L. rhamnosus* GG cells in the interaction with phagocytic innate immune cells, i.e., macrophages. Interest in macrophages as important modulators of the interaction with gut microbiota and probiotics has been renewed, since it has been shown that they can also pass their dendrites through intestinal epithelial cells (26, 43). Moreover,

macrophages can differentiate into classical (M1) or alternative (M2) subsets, with the first group promoting T helper 1 (Th1) immune responses and the second one being related to immunosuppression by having an anti-inflammatory phenotype (44, 45).

In this work, we demonstrated that SpaCBA pili of *L. rhamnosus* GG are crucial for its capacity to adhere to a murine macrophage (RAW 264.7) cell line and that this interaction stimulates bacterial uptake. Additionally, we observed that the pilus-mediated adhesion and enhanced cell-to-cell contact are coupled to an anti-inflammatory response, as monitored by the induction of IL-10 mRNA, since EPS-deficient mutant CMPG5351 with enhanced exposure of the pili and the *L. lactis* subsp. *cremoris* mutant with overexpression of the SpaCBA pili showed a markedly increased ability to induce this anti-inflammatory cytokine in RAW 264.7 cells compared to that of their respective wild-type strains. However, we could not observe any significant differences in the expression of IL-10 mRNA when the cells were stimulated with the nonpilated *spaCBA* mutant CMPG5357 strain and the wild-

type *L. rhamnosus* GG strain, suggesting that the pili *per se* are not required for the induction of IL-10 mRNA. IL-10 was selected as a biomarker for immunological activities due to its crucial role in the suppression and prevention of inflammatory responses (46, 47). This cytokine is produced by several immune cells, such as regulatory CD4⁺ T cells, which are key players in anti-inflammatory immune responses (48), and M2 macrophages, which express an IL-10^{high} phenotype (49, 50). Interestingly, the level of IL-10 mRNA induced was similar to or even greater than that induced by LPS (1 µg/ml), a previously documented bacterial ligand shown by different groups to activate IL-10 gene expression in macrophages (51–53). Our findings after heterologous expression of the SpaCBA pili in *L. lactis* subsp. *cremoris* for IL-10 induction in murine macrophages are in agreement with those of previous work of von Ossowski and coworkers using moDCs (23). Given its zipper-like adhesion role, it is highly likely that the pili do not directly induce large amounts of IL-10 mRNA but, rather, mediate close interaction with the macrophages, resulting in an enhanced delivery of anti-inflammatory molecules. The exact IL-10-inducing molecules in *L. rhamnosus* GG thus remain to be determined, but our Transwell experiments suggest that they are located on the bacterial cell wall. Documented *Lactobacillus* molecules with anti-inflammatory properties include polysaccharide chains of rhamnose-rich EPSs produced by *L. rhamnosus* RW-9595M that induce immunosuppression by the production of anti-inflammatory IL-10 in macrophages (54), but they do not seem to be present in *L. rhamnosus* GG (20). Bernardo and coworkers identified a peptide (STp) secreted by *L. plantarum* BMC12 that could induce intracellular IL-10 in human blood and intestinal DCs (67). Likewise, the S-layer protein A (SlpA) from *L. acidophilus* NCFM was found to mediate the interaction of bacteria with DCs by specifically binding their C-type lectin PRR DC-SIGN, resulting in the induction of IL-10 in the presence of LPS, but IL-10 was not as highly induced by LPS or SlpA alone (18). Furthermore, teichoic acids (TAs) from *L. plantarum* ATCC 14917 were found to be recognized by TLR2, activate the extracellular signal-regulated kinase pathway, and act synergistically with *L. casei* Shirota YIT9029 to induce IL-10 production in mouse peritoneal macrophages (55). However, other studies showed that expression of IL-10 was almost entirely unaffected in THP-1 cells after treatment with LTA isolated from *L. plantarum* KCTC10887BP (56) and that LTA from *L. plantarum* NCIM8826 promotes IL-12 rather than IL-10 induction (57). Clearly, multiple *Lactobacillus* cell wall or secreted molecules could be directly involved in IL-10 mRNA induction.

Furthermore, our results indicate that the SpaCBA pili of *L. rhamnosus* GG do not directly influence the expression of proinflammatory cytokine IL-6 mRNA. We could observe an increased induction for the SpaCBA pilus mutant CMPG5357 compared to that for the wild type, whereas EPS-deficient mutant CMPG5351, showing an enhanced exposure of the pili, had the same levels of expression of IL-6 mRNA as the wild type. In support of the above-mentioned results, further experiments indicated that the SpaCBA-piliated lactococcus GRS1185 also reduced the level of IL-6 mRNA induction in RAW 264.7 cells compared to that achieved with nonpiliated *L. lactis* subsp. *cremoris*. However, these data are not in agreement with those from previous work using human moDCs (23), where the SpaCBA-piliated lactococcal constructs had increased levels of IL-6 production. It is worthy of mention that in the previous study (23), IL-6 production was measured according to the protein levels, while in the present

work, gene expression at the mRNA level was evaluated. Moreover, different antigen-presenting cells (i.e., human monocyte-derived DCs and murine macrophages) were used in both studies, so that a variation in the pattern of cytokine production can be expected due to differential receptor expression (58). Taken together, our data suggest that the presence of pili probably indirectly prevents the induction of IL-6 mRNA by other surface molecules. Interestingly, a former study has demonstrated a role for pathogenic pneumococcal pili in the induction of proinflammatory cytokines, as higher levels of IL-6 were found in the serum of mice previously challenged with *S. pneumoniae* strain T4, which expresses pili, than in the serum of mice treated with its isogenic nonpiliated deletion mutant (59). Although the results from this *in vivo* study cannot be easily compared with those of our molecular interaction studies with cell lines, it is intriguing that pili from pathogenic bacteria induce proinflammatory immune responses in the host, while our data for the pili of the probiotic strain under study showed instead an indirect antipathogenic role. Nevertheless, future studies will have to investigate the role of pili on probiotic lactobacilli in modulating other anti- and proinflammatory biomarkers in different cell types and hosts with different disease states and genetic backgrounds.

To gain a better insight into the molecular interaction between pilated *L. rhamnosus* GG and macrophages, we also investigated the role of two PRRs, i.e., CR3 (CD11b/CD18) and TLR2. CR3 is a receptor present on the surface of most immune cells, including monocytes/macrophages, DCs, neutrophils, and natural killer (NK) cells, and plays an essential role in different immunological processes, including cell activation, chemotaxis, cytotoxicity, and phagocytosis (60) and tolerance induction (61). Previous studies demonstrated that pilus-associated adhesin RrgA from *S. pneumoniae* binds to CR3 and promotes bacterial uptake by murine and human macrophages (24). Using a similar approach with the same anti-CD11b antibodies, our data show that SpaCBA pili of *L. rhamnosus* GG promote bacterial phagocytosis by RAW 264.7 cells partially through CR3. However, the induction of IL-10 and IL-6 mRNA in these macrophages was not affected by the treatment with anti-CD11b antibodies, suggesting that *L. rhamnosus* GG-mediated cytokine modulation occurs in a CR3-independent manner. TLR2 has been found to be involved in the recognition of various MAMPs of Gram-positive bacteria. For instance, we recently demonstrated that LTA from *L. rhamnosus* GG stimulates TLR2/6-dependent activation of NF-κβ signaling in the HEK293T cell line and stimulates proinflammatory cytokines, at least under certain conditions (17). Interestingly, our present results show that induction of IL-6 mRNA by *L. rhamnosus* GG in macrophages is significantly influenced by the interaction with TLR2. This *L. rhamnosus* GG-TLR2 interaction is at least partially mediated by *L. rhamnosus* GG LTA, being a MAMP for TLR2/6, as determined on the basis of our previous work (17). Nevertheless, it is important to emphasize that LPS was still able to induce much higher levels of IL-6 mRNA than all *L. rhamnosus* GG strains tested, indicating that the induction of IL-6 by *L. rhamnosus* GG is subtler than that by the more immunostimulatory LPS of *E. coli*. Blockage of TLR2 also significantly affected the expression of IL-10 mRNA, but only when cells were treated with the EPS-deficient mutant, suggesting that molecules other than SpaCBA pili that are overexposed in this EPS-deficient mutant are signaling through TLR2 and that TLR2 could thus also modulate the anti-inflammatory responses. Interestingly, recent work in

Staphylococcus aureus has also shown that its cell surface contains both proinflammatory and IL-10-inducing immunomodulatory molecules. These peptidoglycan-embedded IL-10-inducing molecules also bind to TLR2 on host antigen-presenting cells, activating the PI3K/Akt pathway, resulting in high levels of production of IL-10, which in turn downregulates the adaptive T cell response (62, 63). Thus, the exact receptor(s) by which the SpaCBA pili could mediate cytokine induction remains to be identified. For such studies, sufficient amounts of highly pure pili need to be obtained and preferentially studied in their native configuration. Such experiments are ongoing in our lab but are complicated.

Taken together, our data indicate that *L. rhamnosus* GG pili play a role in the host interaction with macrophages, mainly by promoting adhesion and internalization. As a result, other MAMPs, both pro- and anti-inflammatory molecules of *L. rhamnosus* GG, can interact more closely with their respective PRRs. Nevertheless, it remains to be substantiated in more complex models that the pilus-mediated high levels of adhesion of *L. rhamnosus* GG to host immune cells combined with a low inflammatory potential would be beneficial to humans.

ACKNOWLEDGMENTS

This work was supported in part through BOF program financing (spokesman, Jan Balzarini; PF/10/018) at KU Leuven and in part through the Fund for Scientific Research, Flanders (FWO; grants KaN 150711N and 1520114N), and IOF-SBO financing at the Universiteit Antwerpen. Dominique Bullens is a recipient of a senior researcher fellowship from the Fund for Scientific Research, Flanders (FWO). I.V.O. and A.P. acknowledge project financing from the Academy of Finland-funded Center of Excellence in Microbial Food Safety (CoE-MiFoSa) research program (grant no. 118602 and 141140) and an Academy of Finland general research grant (no. 118165).

Marijke Segers and Hanne Tytgat are acknowledged for many insightful discussions and suggestions. Lieve Coorevits is gratefully thanked for her invaluable help with the culture of RAW 264.7 cells. We also thank Stefanie Roberfroid, Louis Deforche, and David De Coster for their technical assistance with the fluorescence microscope, as well as Geert Schoofs for his assistance with the setup of the flow cytometry experiments.

REFERENCES

- Bongaerts GPA, Severijnen RSV. 2001. The beneficial, antimicrobial effect of probiotics. *Med Hypotheses* 56:174–177. <http://dx.doi.org/10.1054/mehy.2000.1135>.
- Petrova MI, van den Broek M, Balzarini J, Vanderleyden J, Lebeer S. 2013. Vaginal microbiota and its role in HIV transmission and infection. *FEMS Microbiol Rev* 37:762–792. <http://dx.doi.org/10.1111/1574-6976.12029>.
- Kalliomaki M, Salminen S, Arvilommi H, Kero P, Koskinen P, Isolauri E. 2001. Probiotics in primary prevention of atopic disease: a randomised placebo-controlled trial. *Lancet* 357:1076–1079. [http://dx.doi.org/10.1016/S0140-6736\(00\)04259-8](http://dx.doi.org/10.1016/S0140-6736(00)04259-8).
- Kalliomaki M, Salminen S, Poussa T, Arvilommi H, Isolauri E. 2003. Probiotics and prevention of atopic disease: 4-year follow-up of a randomised placebo-controlled trial. *Lancet* 361:1869–1871. [http://dx.doi.org/10.1016/S0140-6736\(03\)13490-3](http://dx.doi.org/10.1016/S0140-6736(03)13490-3).
- Kalliomaki M, Salminen S, Poussa T, Isolauri E. 2007. Probiotics during the first 7 years of life: a cumulative risk reduction of eczema in a randomized, placebo-controlled trial. *J Allergy Clin Immunol* 119:1019–1021. <http://dx.doi.org/10.1016/j.jaci.2006.12.608>.
- Hatakka K, Savilahti E, Ponka A, Meurman JH, Poussa T, Nase L, Saxelin M, Korpela R. 2001. Effect of long term consumption of probiotic milk on infections in children attending day care centres: double blind, randomised trial. *Br Med J* 322:1327–1329. <http://dx.doi.org/10.1136/bmj.322.7298.1327>.
- Hojsak I, Abdovic S, Szajewska H, Milosevic M, Krzrnaric Z, Kolacek S. 2010. Lactobacillus GG in the prevention of nosocomial gastrointestinal and respiratory tract infections. *Pediatrics* 125:e1171–e1177. <http://dx.doi.org/10.1542/peds.2009-2568>.
- Hojsak I, Snovak N, Abdovic S, Szajewska H, Misak Z, Kolacek S. 2010. Lactobacillus GG in the prevention of gastrointestinal and respiratory tract infections in children who attend day care centers: a randomized, double-blind, placebo-controlled trial. *Clin Nutr* 29:312–316. <http://dx.doi.org/10.1016/j.clnu.2009.09.008>.
- Vanderhoof JA, Whitney DB, Antonson DL, Hanner TL, Lupo JV, Young RJ. 1999. Lactobacillus GG in the prevention of antibiotic-associated diarrhea in children. *J Pediatr* 135:564–568. [http://dx.doi.org/10.1016/S0022-3476\(99\)70053-3](http://dx.doi.org/10.1016/S0022-3476(99)70053-3).
- Guandalini S, Pensabene L, Abu Zikri M, Dias JA, Casali LG, Hoekstra H, Kolacek S, Massar K, Micetic-Turk D, Papadopoulou A, de Sousa JS, Sandhu B, Szajewska H, Weizman Z. 2000. Lactobacillus GG administered in oral rehydration solution to children with acute diarrhea: a multicenter European trial. *J Pediatr Gastroenterol Nutr* 30:54–60. <http://dx.doi.org/10.1097/00005176-200001000-00018>.
- Szajewska H, Skorka A, Rusczyński M, Gieruszczak-Bialek D. 2013. Meta-analysis: Lactobacillus GG for treating acute gastroenteritis in children—updated analysis of randomised controlled trials. *Aliment Pharmacol Ther* 38:467–476. <http://dx.doi.org/10.1111/apt.12403>.
- Guarino A, Canani RB, Spagnuolo MI, Albano F, DiBenedetto L. 1997. Oral bacterial therapy reduces the duration of symptoms and of viral excretion in children with mild diarrhea. *J Pediatr Gastroenterol Nutr* 25:516–519. <http://dx.doi.org/10.1097/00005176-199711000-00005>.
- Lebeer S, Vanderleyden J, De Keersmaecker SCJ. 2010. Host interactions of probiotic bacterial surface molecules: comparison with commensals and pathogens. *Nat Rev Microbiol* 8:171–184. <http://dx.doi.org/10.1038/nrmicro2297>.
- Schlee M, Harder J, Koten B, Stange EF, Wehkamp J, Fellermann K. 2008. Probiotic lactobacilli and VSL#3 induce enterocyte beta-defensin 2. *Clin Exp Immunol* 151:528–535. <http://dx.doi.org/10.1111/j.1365-2249.2007.03587.x>.
- Kleerebezem M, Hols P, Bernard E, Rolain T, Zhou M, Siezen RJ, Bron PA. 2010. The extracellular biology of the lactobacilli. *FEMS Microbiol Rev* 34:199–230. <http://dx.doi.org/10.1111/j.1574-6976.2009.00208.x>.
- Sengupta R, Altermann E, Anderson RC, McNabb WC, Moughan PJ, Roy NC. 2013. The role of cell surface architecture of lactobacilli in host-microbe interactions in the gastrointestinal tract. *Mediators Inflamm* 2013:237921. <http://dx.doi.org/10.1155/2013/237921>.
- Claes IJJ, Segers ME, Verhoeven TLA, Dusselier M, Sels BF, De Keersmaecker SCJ, Vanderleyden J, Lebeer S. 2012. Lipoteichoic acid is an important microbe-associated molecular pattern of Lactobacillus rhamnosus GG. *Microb Cell Fact* 11:161. <http://dx.doi.org/10.1186/1475-2859-11-161>.
- Konstantinov SR, Smidt H, de Vos WM, Bruijns SC, Singh SK, Valence F, Molle D, Lortal S, Altermann E, Kleenhammer TR, van Kooyk Y. 2008. S layer protein A of Lactobacillus acidophilus NCFM regulates immature dendritic cell and T cell functions. *Proc Natl Acad Sci U S A* 105:19474–19479. <http://dx.doi.org/10.1073/pnas.0810305105>.
- Macho Fernandez EM, Valenti V, Rockel C, Hermann C, Pot B, Boneca IG, Grangette C. 2011. Anti-inflammatory capacity of selected lactobacilli in experimental colitis is driven by NOD2-mediated recognition of a specific peptidoglycan-derived muropeptide. *Gut* 60:1050–1059. <http://dx.doi.org/10.1136/gut.2010.232918>.
- Lebeer S, Verhoeven TLA, Francius G, Schoofs G, Lambrichts I, Dufrene Y, Vanderleyden J, De Keersmaecker SCJ. 2009. Identification of a gene cluster for the biosynthesis of a long, galactose-rich exopolysaccharide in Lactobacillus rhamnosus GG and functional analysis of the priming glycosyltransferase. *Appl Environ Microbiol* 75:3554–3563. <http://dx.doi.org/10.1128/AEM.02919-08>.
- Kankainen M, Paulin L, Tynkkynen S, von Ossowski I, Reunanen J, Partanen P, Satokari R, Vesterlund S, Hendrickx APA, Lebeer S, De Keersmaecker SCJ, Vanderleyden J, Hamalainen T, Laukkanen S, Salovuori N, Ritari J, Alatalo E, Korpela R, Mattila-Sandholm T, Lassig A, Hatakka K, Kinnunen KT, Karjalainen H, Saxelin M, Laakso K, Surakka A, Palva A, Salusjarvi T, Auvinen P, de Vos WM. 2009. Comparative genomic analysis of Lactobacillus rhamnosus GG reveals pili containing a human-mucus binding protein. *Proc Natl Acad Sci U S A* 106:17193–17198. <http://dx.doi.org/10.1073/pnas.0908876106>.
- Lebeer S, Claes I, Tytgat HLP, Verhoeven TLA, Marien E, von Ossowski I, Reunanen J, Palva A, de Vos WM, De Keersmaecker SCJ, Vanderleyden J. 2012. Functional analysis of Lactobacillus rhamnosus GG pili in

- relation to adhesion and immunomodulatory interactions with intestinal epithelial cells. *Appl Environ Microbiol* 78:185–193. <http://dx.doi.org/10.1128/AEM.06192-11>.
23. von Ossowski I, Pietila TE, Rintahaka J, Nummenmaa E, Makinen VM, Reunanen J, Satokari R, de Vos WM, Palva I, Palva A. 2013. Using recombinant lactococci as an approach to dissect the immunomodulating capacity of surface piliation in probiotic *Lactobacillus rhamnosus* GG. *PLoS One* 8:e64416. <http://dx.doi.org/10.1371/journal.pone.0064416>.
 24. Orrskog S, Rounioja S, Spadafina T, Gallotta M, Norman M, Hentrich K, Falker S, Ygberg-Eriksson S, Hasenberger M, Johansson B, Uotila LM, Gahmberg CG, Barocchi M, Gunzer M, Normark S, Henriques-Normark B. 2012. Pilus adhesin RrgA interacts with complement receptor 3, thereby affecting macrophage function and systemic pneumococcal disease. *mBio* 4(1):e00535-12. <http://dx.doi.org/10.1128/mBio.00535-12>.
 25. Basset A, Zhang F, Benes C, Sayeed S, Herd M, Thompson C, Golenbock DT, Camilli A, Malley R. 2013. Toll-like receptor (TLR) 2 mediates inflammatory responses to oligomerized RrgA pneumococcal pilus type 1 protein. *J Biol Chem* 288:2665–2675. <http://dx.doi.org/10.1074/jbc.M112.398875>.
 26. Medina-Contreras O, Geem D, Laur O, Williams IR, Lira SA, Nusrat A, Parkos CA, Denning TL. 2011. CX3CR1 regulates intestinal macrophage homeostasis, bacterial translocation, and colitogenic Th17 responses in mice. *J Clin Invest* 121:4787–4795. <http://dx.doi.org/10.1172/JCI59150>.
 27. Niess JH, Brand S, Gu X, Landsman L, Jung S, McCormick BA, Vyas JM, Boes M, Ploegh HL, Fox JG, Littman DR, Reinecker HC. 2005. CX3CR1-mediated dendritic cell access to the intestinal lumen and bacterial clearance. *Science* 307:254–258. <http://dx.doi.org/10.1126/science.1102901>.
 28. Rescigno M, Urbano M, Valzasina B, Francolini M, Rotta G, Bonasio R, Granucci F, Kraehenbuhl JP, Ricciardi-Castagnoli P. 2001. Dendritic cells express tight junction proteins and penetrate gut epithelial monolayers to sample bacteria. *Nat Immunol* 2:361–367. <http://dx.doi.org/10.1038/86373>.
 29. Tripathi P, Beaussart A, Alsteens D, Dupres V, Claes I, von Ossowski I, de Vos WM, Palva A, Lebeer S, Vanderleyden J, Dufrene YF. 2013. Adhesion and nanomechanics of pili from the probiotic *Lactobacillus rhamnosus* GG. *ACS Nano* 7:3685–3697. <http://dx.doi.org/10.1021/nn400705u>.
 30. Wegmann U, O'Connell-Motherway M, Zomer A, Buist G, Shearman C, Canchaya C, Ventura M, Goesmann A, Gasson MJ, Kuipers OP, van Sinderen D, Kok J. 2007. Complete genome sequence of the prototype lactic acid bacterium *Lactococcus lactis* subsp. *cremoris* MG1363. *J Bacteriol* 189:3256–3270. <http://dx.doi.org/10.1128/JB.01768-06>.
 31. Eichenbaum Z, Federle MJ, Marra D, de Vos WM, Kuipers OP, Kleerebezem M, Scott JR. 1998. Use of the lactococcal *nisA* promoter to regulate gene expression in gram-positive bacteria: comparison of induction level and promoter strength. *Appl Environ Microbiol* 64:2763–2769.
 32. Claes IJ, Schoofs G, Regulski K, Courtin P, Chapot-Chartier MP, Rolain T, Hols P, von Ossowski I, Reunanen J, de Vos WM, Palva A, Vanderleyden J, De Keersmaecker SC, Lebeer S. 2012. Genetic and biochemical characterization of the cell wall hydrolase activity of the major secreted protein of *Lactobacillus rhamnosus* GG. *PLoS One* 7:e31588. <http://dx.doi.org/10.1371/journal.pone.0031588>.
 33. De Keersmaecker SC, Braeken K, Verhoeven TL, Perea Velez M, Lebeer S, Vanderleyden J, Hols P. 2006. Flow cytometric testing of green fluorescent protein-tagged *Lactobacillus rhamnosus* GG for response to defensins. *Appl Environ Microbiol* 72:4923–4930. <http://dx.doi.org/10.1128/AEM.02605-05>.
 34. Pavan S, Hols P, Delcour J, Geoffroy MC, Grangette C, Kleerebezem M, Mercenier A. 2000. Adaptation of the nisin-controlled expression system in *Lactobacillus plantarum*: a tool to study in vivo biological effects. *Appl Environ Microbiol* 66:4427–4432. <http://dx.doi.org/10.1128/AEM.66.10.4427-4432.2000>.
 35. Dupont L, Boizet-Bonhoure B, Coddeville M, Auvray F, Ritzenthaler P. 1995. Characterization of genetic elements required for site-specific integration of *Lactobacillus delbrueckii* subsp. *bulgaricus* bacteriophage mv4 and construction of an integration-proficient vector for *Lactobacillus plantarum*. *J Bacteriol* 177:586–595.
 36. Geoffroy MC, Guyard C, Quatannens B, Pavan S, Lange M, Mercenier A. 2000. Use of green fluorescent protein to tag lactic acid bacterium strains under development as live vaccine vectors. *Appl Environ Microbiol* 66:383–391. <http://dx.doi.org/10.1128/AEM.66.1.383-391.2000>.
 37. Chapot-Chartier MP, Vinogradov E, Sadovskaya I, Andre G, Mistou MY, Trieu-Cuot P, Furlan S, Bidnenko E, Courtin P, Pechoux C, Hols P, Dufrene YF, Kulakauskas S. 2010. Cell surface of *Lactococcus lactis* is covered by a protective polysaccharide pellicle. *J Biol Chem* 285:10464–10471. <http://dx.doi.org/10.1074/jbc.M109.082958>.
 38. Giulietti A, Overbergh L, Valckx D, Decallonne B, Bouillon R, Mathieu C. 2001. An overview of real-time quantitative PCR: applications to quantify cytokine gene expression. *Methods* 25:386–401. <http://dx.doi.org/10.1006/meth.2001.1261>.
 39. Hafstad AD, Khalid AM, Hagve M, Lund T, Larsen TS, Severson DL, Clarke K, Berge RK, Aasum E. 2009. Cardiac peroxisome proliferator-activated receptor- α activation causes increased fatty acid oxidation, reducing efficiency and post-ischaemic functional loss. *Cardiovasc Res* 83:519–526. <http://dx.doi.org/10.1093/cvr/cvp132>.
 40. Overbergh L, Giulietti A, Valckx D, Decallonne R, Bouillon R, Mathieu C. 2003. The use of real-time reverse transcriptase PCR for the quantification of cytokine gene expression. *J Biomol Tech* 14:33–43.
 41. Kanaya S, Nemoto E, Ogawa T, Shimauchi H. 2009. Porphyromonas gingivalis fimbriae induce unique dendritic cell subsets via Toll-like receptor 2. *J Periodontol Res* 44:543–549. <http://dx.doi.org/10.1111/j.1600-0765.2008.01149.x>.
 42. Lebeer S, Vanderleyden J, De Keersmaecker SC. 2008. Genes and molecules of lactobacilli supporting probiotic action. *Microbiol Mol Biol Rev* 72:728–764. <http://dx.doi.org/10.1128/MMBR.00017-08>.
 43. Peterson LW, Artis D. 2014. Intestinal epithelial cells: regulators of barrier function and immune homeostasis. *Nat Rev Immunol* 14:141–153. <http://dx.doi.org/10.1038/nri3608>.
 44. Barros MH, Hauck F, Dreyer JH, Kempkes B, Niedobitek G. 2013. Macrophage polarisation: an immunohistochemical approach for identifying M1 and M2 macrophages. *PLoS One* 8:e80908. <http://dx.doi.org/10.1371/journal.pone.0080908>.
 45. Sica A, Mantovani A. 2012. Macrophage plasticity and polarization: in vivo veritas. *J Clin Invest* 122:787–795. <http://dx.doi.org/10.1172/JCI59643>.
 46. Saraiva M, O'Garra A. 2010. The regulation of IL-10 production by immune cells. *Nat Rev Immunol* 10:170–181. <http://dx.doi.org/10.1038/nri2711>.
 47. Zhang Y, Kim HJ, Yamamoto S, Kang XY, Ma XJ. 2010. Regulation of interleukin-10 gene expression in macrophages engulfing apoptotic cells. *J Interferon Cytokine Res* 30:113–121. <http://dx.doi.org/10.1089/jir.2010.0004>.
 48. O'Garra A, Vieira PL, Vieira P, Goldfeld AE. 2004. IL-10-producing and naturally occurring CD4⁺ Tregs: limiting collateral damage. *J Clin Invest* 114:1372–1378. <http://dx.doi.org/10.1172/JCI200423215>.
 49. Martinez FO, Gordon S. 2014. The M1 and M2 paradigm of macrophage activation: time for reassessment. *F1000prime Rep* 6:13. <http://dx.doi.org/10.12703/P6-13>.
 50. Tatano Y, Shimizu T, Tomioka H. 2014. Unique macrophages different from M1/M2 macrophages inhibit T cell mitogenesis while upregulating Th17 polarization. *Sci Rep* 4:4146. <http://dx.doi.org/10.1038/srep04146>.
 51. Brightbill HD, Plevy SE, Modlin RL, Smale ST. 2000. A prominent role for Sp1 during lipopolysaccharide-mediated induction of the IL-10 promoter in macrophages. *J Immunol* 164:1940–1951. <http://dx.doi.org/10.4049/jimmunol.164.4.1940>.
 52. Iyer SS, Ghaffari AA, Cheng G. 2010. Lipopolysaccharide-mediated IL-10 transcriptional regulation requires sequential induction of type I IFNs and IL-27 in macrophages. *J Immunol* 185:6599–6607. <http://dx.doi.org/10.4049/jimmunol.1002041>.
 53. Liu YW, Chen CC, Tseng HP, Chang WC. 2006. Lipopolysaccharide-induced transcriptional activation of interleukin-10 is mediated by MAPK- and NF- κ B-induced CCAAT/enhancer-binding protein delta in mouse macrophages. *Cell Signal* 18:1492–1500. <http://dx.doi.org/10.1016/j.cellsig.2005.12.001>.
 54. Bleau C, Monges A, Rashidan K, Laverdure JP, Lacroix M, Van Calsteren MR, Millette M, Savard R, Lamontagne L. 2010. Intermediate chains of exopolysaccharides from *Lactobacillus rhamnosus* RW-9595M increase IL-10 production by macrophages. *J Appl Microbiol* 108:666–675. <http://dx.doi.org/10.1111/j.1365-2672.2009.04450.x>.
 55. Kaji R, Kiyoshima-Shibata J, Nagaoka M, Nanno M, Shida K. 2010. Bacterial teichoic acids reverse predominant IL-12 production induced by certain *Lactobacillus* strains into predominant IL-10 production via TLR2-dependent ERK activation in macrophages. *J Immunol* 184:3505–3513. <http://dx.doi.org/10.4049/jimmunol.0901569>.
 56. Kim HG, Gim MG, Kim JY, Hwang HJ, Ham MS, Lee JM, Hartung T, Park JW, Han SH, Chung DK. 2007. Lipoteichoic acid from *Lactobacil-*

- lus plantarum elicits both the production of interleukin-23p19 and suppression of pathogen-mediated interleukin-10 in THP-1 cells. *FEMS Immunol Med Microbiol* 49:205–214. <http://dx.doi.org/10.1111/j.1574-695X.2006.00175.x>.
57. Grangette C, Nutten S, Palumbo E, Morath S, Hermann C, Dewulf J, Pot B, Hartung T, Hols P, Mercenier A. 2005. Enhanced anti-inflammatory capacity of a *Lactobacillus plantarum* mutant synthesizing modified teichoic acids. *Proc Natl Acad Sci U S A* 102:10321–10326. <http://dx.doi.org/10.1073/pnas.0504084102>.
 58. Liu BS, Janssen HL, Boonstra A. 2011. IL-29 and IFN α differ in their ability to modulate IL-12 production by TLR-activated human macrophages and exhibit differential regulation of the IFN γ receptor expression. *Blood* 117:2385–2395. <http://dx.doi.org/10.1182/blood-2010-07-298976>.
 59. Barocchi MA, Ries J, Zogaj X, Hemsley C, Albiger B, Kanth A, Dahlberg S, Fernebro J, Moschioni M, Massignani V, Hultenby K, Taddei AR, Beiter K, Wartha F, von Euler A, Covacci A, Holden DW, Normark S, Rappuoli R, Henriques-Normark B. 2006. A pneumococcal pilus influences virulence and host inflammatory responses. *Proc Natl Acad Sci U S A* 103:2857–2862. <http://dx.doi.org/10.1073/pnas.0511017103>.
 60. Springer TA. 1990. Adhesion receptors of the immune-system. *Nature* 346:425–434. <http://dx.doi.org/10.1038/346425a0>.
 61. Ehrchiou D, Xiong Y, Xu GW, Chen WJ, Shi YF, Zhang L. 2007. CD11b facilitates the development of peripheral tolerance by suppressing Th17 differentiation. *J Exp Med* 204:1519–1524. <http://dx.doi.org/10.1084/jem.20062292>.
 62. Chau TA, McCully ML, Brintnell W, An G, Kasper KJ, Vines ED, Kubes P, Haeryfar SM, McCormick JK, Cairns E, Heinrichs DE, Madrenas J. 2009. Toll-like receptor 2 ligands on the staphylococcal cell wall down-regulate superantigen-induced T cell activation and prevent toxic shock syndrome. *Nat Med* 15:641–648. <http://dx.doi.org/10.1038/nm.1965>.
 63. Peres AG, Madrenas J. 2013. The broad landscape of immune interactions with *Staphylococcus aureus*: from commensalism to lethal infections. *Burns* 39:380–388. <http://dx.doi.org/10.1016/j.burns.2012.12.008>.
 64. Francius G, Lebeer S, Alsteens D, Wildling L, Gruber HJ, Hols P, De Keersmaecker S, Vanderleyden J, Dufrene YF. 2008. Detection, localization, and conformational analysis of single polysaccharide molecules on live bacteria. *ACS Nano* 2:1921–1929. <http://dx.doi.org/10.1021/nm800341b>.
 65. Reunanen J, von Ossowski I, Hendrickx AP, Palva A, de Vos WM. 2012. Characterization of the SpaCBA pilus fibers in the probiotic *Lactobacillus rhamnosus* GG. *Appl Environ Microbiol* 78:2337–2344. <http://dx.doi.org/10.1128/AEM.07047-11>.
 66. Tripathi P, Dupres V, Beaussart A, Lebeer S, Claes JJ, Vanderleyden J, Dufrene YF. 2012. Deciphering the nanometer-scale organization and assembly of *Lactobacillus rhamnosus* GG pili using atomic force microscopy. *Langmuir* 28:2211–2216. <http://dx.doi.org/10.1021/la203834d>.
 67. Bernardo D, Sánchez B, Al-Hassi HO, Mann ER, Urdaci MC, Knight SC, Margolles A. 2012. Microbiota/host crosstalk biomarkers: regulatory response of human intestinal dendritic cells exposed to *Lactobacillus* extracellular encrypted peptide. *PLoS One* 7(5):e36262. <http://dx.doi.org/10.1371/journal.pone.0036262>.