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Two sub-Antarctic and Northern Europe distributed diatom species found in a middle-mountain lake in France

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ABSTRACT

During a survey of the diatom flora of Longemer Lake (Konrupt-Longemer, Vosges, France), two species with an unusual biogeographical distribution were observed. *Psammothidium abundans*, previously considered being a sub-Antarctic endemic but today also found in rivers in Sweden, Ireland and the UK, and *Achnanthidium sublineare*, up to now confined to northern Europe, were identified. These are the first records of these rare diatom species in France (*A. sublineare*) and in a European lake (*P. abundans*). Since there are only a few illustrations of these two species in the literature, both populations are illustrated using light and scanning electron microscopy in order to provide a detailed morphological account. Details on their actual biogeographical distribution are added.

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Achnanthidium sublineare; *Psammothidium abundans*; diatoms; France; lake; Vosges

Introduction

Our understanding of the biogeography of freshwater diatoms has undergone drastic changes during the past 15 years. For a long time, the biogeography of diatoms was only poorly known (Mann 1999), leading to a large number of hypotheses and speculations (Finlay 2002; Martiny et al. 2006) concerning their cosmopolitan distribution. The endemic or ubiquitous feature of their distribution was often difficult to ascertain (Vanormelingen, Verleyen, and Vyverman 2008), frequently influenced by an incorrect assessment of their diversity due to taxonomic drift and force-fitting (Tyler 1996). Moreover, the species concept within the Bacillariophyta is complex to delineate (Mann 1999). Taxa are often split based on small differences in the morphology of their frustules, lacking a full understanding of phenotypical variations (Cox 1995). Furthermore, the diatom life cycle following asexual reproduction leads to a broad size range, often inducing pronounced morphological variation (Round, Crawford, and Mann 1990).

Uniformity and identification accuracy across a national territory, a continent, or even worldwide is linked to the operators' capacity to access relevant literature for the area under survey. Typically, the visibility of European floristic guidebooks across the globe enhanced the idea that a lot of (occasionally locally endemic) taxa can be considered being widespread by “force-fitting” these taxa into European or North American names (Tyler 1996; Kociolek and Spaulding 2000). The development of diatom-based methods for

monitoring water quality increases the sampling effort, especially in previously overlooked waterbodies: this tends to widen the species known distribution to new environments (Van de Vijver et al. 2008b). Moreover, the human-mediated introductions of exotic species in a growing number of areas complicates our understanding of the geographical distribution and ecology of certain taxa (Coste and Ector 2000).

Sellaphora archibaldii (J.C.Taylor & Lange-Bertalot) Ács, C.E.Wetzel & Ector can be considered as a good example of a purported endemic species (Ács et al. 2017). Some other taxa were overlooked due to a too wide species concept (Mann and Droop 1996) and are now distinguished as separate (often newly described) species. For example, *Achnanthidium hoffmannii* Van de Vijver, Ector, A.Mertens & Jarlman, was originally described from a Swedish river (Van de Vijver et al. 2011b), but concealed for a long time in the *Achnanthidium minutissimum* (Kützing) Czarnecki complex and is now regularly observed in several siliceous bedrock rivers across France (Ector et al. 2015; Peeters and Ector 2018). Finally, it is still difficult to ascertain the distribution of a species by bibliographical means as most checklists do not provide illustrations of the taxa (Mann and Droop 1996) and researchers not infrequently apply the same name to different species, but also occasionally different names to the same species (Mann and Vanormelingen 2013).

The purpose of this paper is to illustrate two species, originally thought to be restricted either to sub-Antarctic or Northern Europe habitats, which were

identified during a floristic survey of Longemer Lake (Xonrupt-Longemer, Vosges): *Psammothidium abundans* (Manguin) Bukhtiyarova & Round and *Achnanthidium sublineare* Van de Vijver, Jarlman & Ector. Based on the available literature, these observations seems to represent their first published record in France and in a European lake.

Material and methods

Study area

The Longemer Lake is located in the Vosges Mountains at 736 m above sea level. It was formed by morainic blocks originating from the Honeck glacier. The lake, with a length of about 1950 m and a width of 550 m, is crossed by the Vologne River. It has a surface area of 76 ha, a maximum depth of around 30 m and a mean depth of 14.2 m. It has a capacity of 10^7 m^3 and a mean retention time of eight months (Centre Technique du Génie Rural des Eaux et Forêts 1979; Collas 2011, 2016). It is the second lake of natural origin in the Vosges Mountains in terms of size and capacity. The substratum is essentially constituted of the impermeable crystalline (granite) and cristalphylian (gneiss) rocks. Because of the chemical nature of the bedrocks, the water is circumneutral to slightly acidic. It is an oligotrophic lake, with low temperature and well oxygenated water. There are several activities on the lake such as fishing, swimming and non-motorised nautical activities (sailing or canoeing). Maps showing the location and hydrosystem of Longemer Lake can be found in Heudre et al. (2017, fig. 1, 2018, fig. 1).

Processing and observations

The samples were collected according to the French survey protocol for benthic diatoms in lakes (Institut national de recherche en sciences et technologies 2013). They are collected at the sampling sites following AFNOR (2010). In the Longemer Lake there are three sites ($6^{\circ}57'15''$ E $48^{\circ}4'14''$ N; $6^{\circ}57'28''$ E $48^{\circ}3'54''$ N; $6^{\circ}56'47''$ E $48^{\circ}4'23''$ N), which were inspected on

12 July 2017. As no suitable macrophytes were available on those sites, diatoms were sampled only from hard mineral substrates.

Diatom slides were made following the French diatom protocol (AFNOR 2016). The samples were treated by oxidation with hot 37% hydrogen peroxide (H_2O_2) and hydrochloric acid (HCl), and rinsed three times with deionised water. For light microscopy (LM) observations, cleaned diatoms were mounted with Naphrax®. LM was performed with an OLYMPUS BX53 microscope using a $\times 100$ oil immersion objective and a Jenoptik ProgRes Speed XT3 camera. For scanning electron microscopy (SEM), parts of the oxidised suspensions were filtered with additional deionised water through a 3- μm Isopore™ polycarbonate membrane filter (Merck Millipore). Filters were mounted on aluminium stubs and coated with platinum using a Modular High Vacuum Coating System BAL-TEC MED 020 (BAL-TEC AG, Balzers, Liechtenstein). An ultra-high-resolution analytical field emission (FE) scanning electron microscope, Hitachi SU-70 (Hitachi High-Technologies Corporation, Japan), operated at 5 kV and 10 mm distance, was used for the analysis. SEM images were taken using the lower (SE-L) detector signal and a tilting of up to 28° . Micrographs were digitally manipulated and plates containing light and scanning electron microscopy images were created using CorelDraw X8®.

Physical and chemical data (Table 1) were taken from the “Système d’Information sur l’Eau Rhin-Meuse (S.I.E.R.M.)” (<http://www.rhin-meuse.eau-france.fr/>). The presented data are related to samples taken in the euphotic zone at the deepest point of the Longemer Lake on 27 July 2017.

Results

Division Bacillariophyta
 Class Bacillariophyceae
 Subclass Bacillariophycidae
 Order Achnanthales
 Family Achnanthidiaceae
 Genus *Psammothidium*

Table 1. Values of physical and chemical parameters from Longemer Lake on 27 July 2017. DOC: Dissolved Organic Carbon; COD: Chemical Oxygen Demand; BOD₅: Biochemical Oxygen Demand after five days.

	Longemer Lake	Longemer Lake	
NH ₄ ⁺ (mg.l ⁻¹)	0.01	Pheopigments (µg.l ⁻¹)	1
NO ₂ ⁻ (mg.l ⁻¹)	0.01	Ca ²⁺ (mg.l ⁻¹)	2.9
NO ₃ ⁻ (mg.l ⁻¹)	0.6	Cl ⁻ (mg.l ⁻¹)	18
Kjeldhal Nitrogen (mg.l ⁻¹)	0.5	Mg ²⁺ (mg.l ⁻¹)	0.88
Total Nitrogen (mg.l ⁻¹)	0.13	K ⁺ (mg.l ⁻¹)	0.3
PO ₄ ³⁻ (mg.l ⁻¹)	0.01	SiO ₂ (mg.l ⁻¹)	2.7
Total phosphorus (mg.l ⁻¹)	0.01	Na ⁺ (mg.l ⁻¹)	12.5
SO ₄ ²⁻ (mg.l ⁻¹)	2.8	Conductivity (µS.cm ⁻¹)	88
DOC (mg.l ⁻¹)	1.7	Suspended Matter (mg.l ⁻¹)	2
COD (mg.l ⁻¹)	20	pH	7.1
BOD ₅ (mg.l ⁻¹)	0.9	Dissolved Oxygen (mg.l ⁻¹)	9.9
Chlorophyll a (µg.l ⁻¹)	3	Dissolved Oxygen saturation (%)	88

Psammothidium abundans (Figures 1–16) has been found in two samples from Longemer Lake occurring in low relative abundance (maximum of 4% of the total valve count).

Morphology

Valves linear, usually slightly swollen in the middle part, with broadly rounded, only weakly protracted apices. Length 10.0–12.5 µm, width 3.5–4.0 µm. Raphe valve with a very narrow, linear axial area. Raphe filiform (Figure 13). Central area rectangular, bordered by shortened striae, the latter occasionally absent (Figures 2 and 12). Rapheless valve with a slightly lanceolate axial area, widening to an enlarged central area (Figures 6–8, 14 and 16). Transapical striae radiate throughout, 28–32 in 10 µm, a little more spaced in the valve middle. Areolae not visible in LM. Areolae rounded, often becoming transapically elongated to slit-like near the valve margin and near the central area (Figures 13–16). Small areolae terminating the striae on the valve face eventually present (Figure 14).

Genus *Achnanthidium*

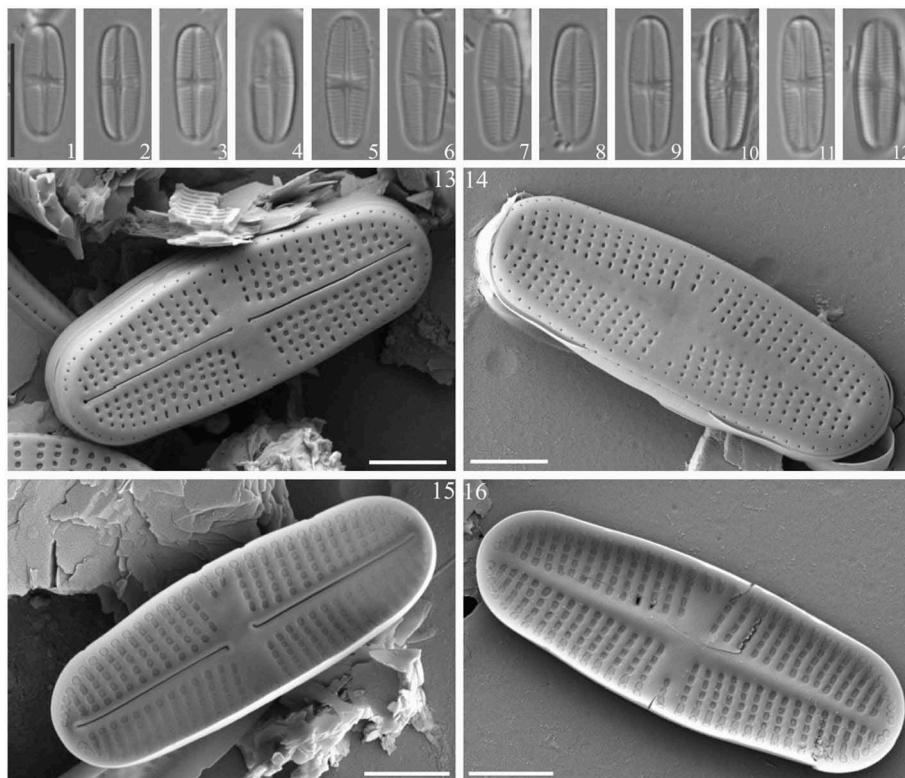
Achnanthidium sublineare (Figures 17–50) was present in all three samples, with a relatively high abundance in two of them (with a maximum of 25.5%).

Morphology

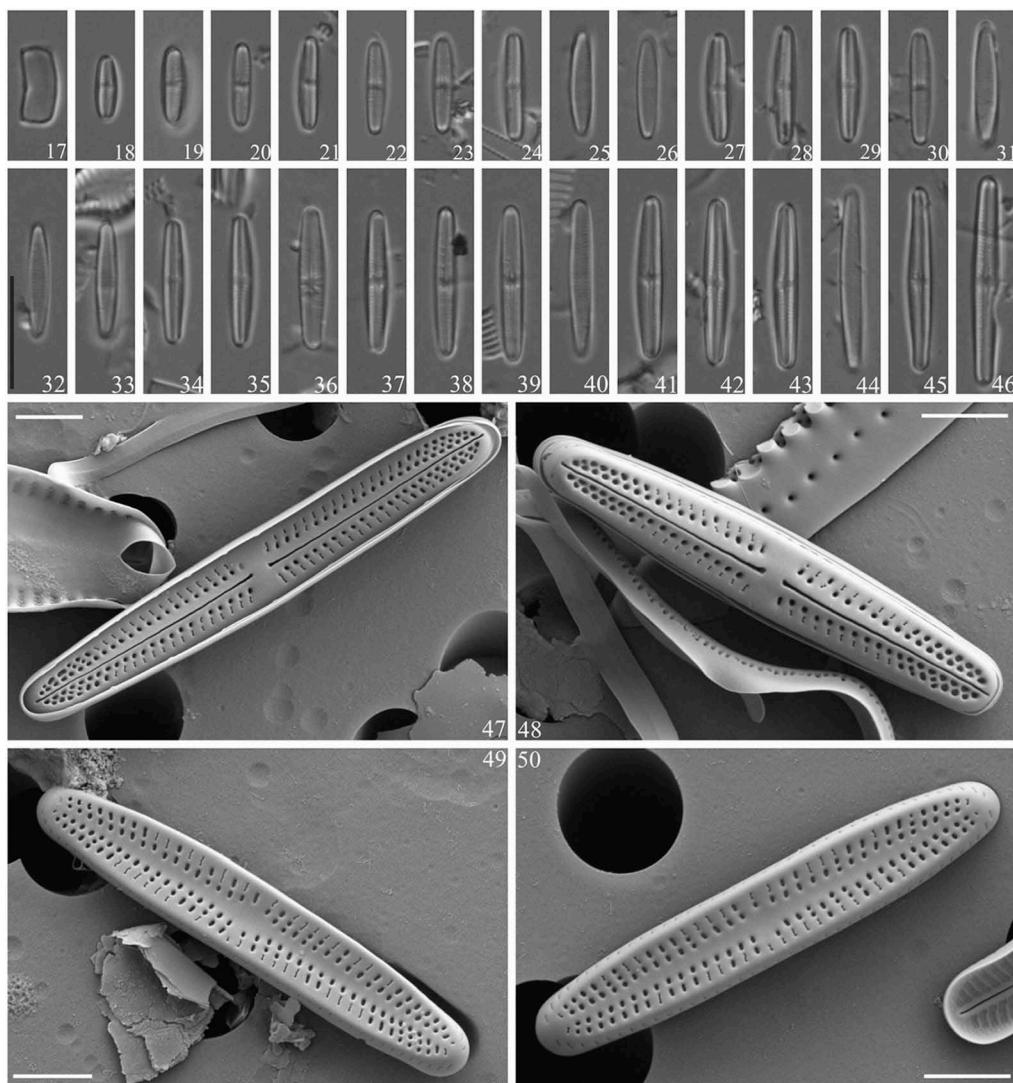
Raphe valves strictly linear. Rapheless valves narrowly lanceolate. Apices acutely rounded, never rostrate, protracted nor capitate. Length 6.0–19.0 µm, width 1.3–2.0 µm. Axial area narrows near the apices, lanceolate, widening towards the central area (Figures 47–50). Central area of the raphe valves formed by a rectangular fascia (Figures 27–30 and 45–48) whereas central area almost absent in the rapheless valves (Figures 31–33, 49 and 50). Transapical striae radiate throughout the entire valve, more strongly radiate and less spaced towards the apices, 32–33 in 10 µm. One, rarely two, rounded areolae present near the raphe and one areola slit-like near the valve margin. At the apices, all areolae rounded (Figures 47–50).

Discussion

Achnanthes abundans Manguin was originally described together with its var. *elliptica* from a sample collected on Iles Kerguelen, located in the southern Indian Ocean (Bourrelly and Manguin 1954, 19). Larson (1974), Le Cohu and Maillard (1983), Van de Vijver et al. (1998) and Gremmen et al. (2007) observed *P. abundans* on different localities on Iles Kerguelen. Le Cohu and Maillard (1983) amended the diagnosis of the species, including the variety *elliptica* within the nominate form



Figures 1–16. *Psammothidium abundans* (material from Longemer Lake at Xonrupt-Longemer). 1–12. LM valve views. Scale bar = 10 µm. 13–16. SEM views. 13. Raphe valve in external view. 14. Rapheless valve in external view. 15. Raphe valve in internal view. 16. Rapheless valve in internal view. Scale bar = 2 µm.



Figures 17–50. *Achnanthidium sublineare* (material from Longemer Lake at Xonrupt-Longemer). 17–46. LM girdle (17) and valve (18–46) views. Scale bar = 10 µm. 47–50. SEM views. 47–48. Raphe valve in external view. 49–50. Rapheless valve in external view.

as the separation was only based on valve outline lacking a thorough analysis of the entire cell cycle. In 1996, Bukhtiyarova and Round transferred *A. abundans* to the genus *Psammothidium*.

Psammothidium abundans is widespread in the Antarctic region as can be seen by the numerous records in Kellogg and Kellogg (2002). Most records were reported from the sub-Antarctic region (McBride, Selkirk, and Adamson 1999; Van de Vijver and Beyens 1999; Van de Vijver, Frenot, and Beyens 2002; Van de Vijver et al. 2004; Van de Vijver and Gremmen 2006; Van de Vijver, Gremmen, and Smith 2008a; Saunders, Hodgson, and McMinn 2009; Chattová, Lebouvier, and Van de Vijver 2014) but there are also records from the Maritime and Continental Antarctic regions (Oppenheim 1990, 1994; Schmidt, Mausbacher, and Müller 1990; Roberts and McMinn 1999; Hodgson, Vyverman, and Sabbe 2001; Sabbe et al. 2003, 2004; Hodgson et al. 2005, 2006; Roberts et al. 2006; Kopalová and Van de Vijver 2013; Sterken et al. 2015; Zidarova,

Kopalová, and Van de Vijver 2016; John 2018). Besides these numerous Antarctic records, the species was found in some neighbouring localities such as Easter Island, South Africa, southern Australia and southern Argentina (Cocquyt 1991; Dumont et al. 1998; Maidana 2000; Sonneman et al. 2001; Flower 2005; Newall and Walsh 2005; Van de Vijver et al. 2008b; Borromei et al. 2010; Harding and Taylor 2014; García and Maidana 2015). In the Northern Hemisphere only a few records exist from rivers and brooks in Sweden, the United Kingdom and Ireland (Lange-Bertalot and Krammer 1989; Bukhtiyarova and Round 1996; Van de Vijver et al. 2008b). Figure 51 shows a world distribution map based on all previously quoted references. *Psammothidium abundans* is mainly found in unpolluted environments such as acidic freshwater lakes and ponds with low conductivity, but also in wet mosses and soils (Le Cohu 2005). It is probably more widespread, as our knowledge of its distribution was considerably widened during the last few decades.

The population of *Psammothidium abundans* from Longemer Lake fits into the valve dimensions and characteristics of the type population as shown in Bourrelly and Manguin (1954) and Van de Vijver et al. (2008b). However, large and clearly rostro-capitulated valves as shown in the Kerguelen type population (Van de Vijver et al. 2008b, figs 2, 3, 16–18) have not been observed. Moreover, the observed French specimens show a high similarity to all other known European populations (Van de Vijver et al. 2008b). The small areolae terminating the striae on the valve face shown in Bukhtiyarova and Round (1996) are present (Figures 13 and 14). The ecology seems similar to that found in the Antarctic Region: oligotrophic lakes with slightly acidic to circumneutral, cold water. Van de Vijver et al. (2008b) discussed the possible origin of this presumably Antarctic-endemic species in European waterbodies. Since morphology and ecology do not separate populations from both hemispheres, molecular-phylogenetic studies may be useful in the future to investigate possible cryptic diversity.

Achnanthidium sublineare Van de Vijver, Jarlman & Ector was described from a sample collected in a Swedish river, Semlan, Sweden (Van de Vijver et al. 2011a). Although published already more than seven years ago, additional records of this species are lacking in the literature. Figure 51 shows the two occurrences of this species known so far.

The morphology of the Longemer population corresponds almost entirely with the original diagnosis. *Achnanthidium sublineare* is quite similar to *Achnanthidium lineare* W. Smith in light microscopy since they are only separated based on valve width,

respectively 1.5–2.1 µm (*A. sublineare*) and 2.2–2.8 µm (*A. lineare*) (Van de Vijver et al. 2011a). However, most of the rapheless valves in the Longemer population possess clearly non-protracted apices, contrary to *A. lineare* that shows clearly rostrate apices and the maximum valve length differs also clearly. In scanning electronic microscopy, differences are also not very obvious. The axial area of the rapheless valve of *A. sublineare* (Figures 49 and 50) is narrower than observed in *A. lineare* (Krahn et al. 2018, figs 88–94) and the stria density differs slightly (33–35 striae per 10 µm for *A. sublineare* vs. 28–32 for *A. lineare*). The striae in *Achnanthidium sublineare* are mainly composed of one rounded and one slit-like areola near the central area, and by usually two (Figures 48 and 49) or rarely three (Figures 47 and 50) rounded areolae towards the apices. The striae in *Achnanthidium lineare* on the contrary are usually composed of three areolae: two rounded and a third rounded or slit-like near the valve margins (Van de Vijver et al. 2011a). It is nevertheless possible that some individuals present striae formed by two areolae as well (Krahn et al. 2018, figs 89, 93).

Achnanthidium sublineare might be more frequent than expected due to undersampling and under-reporting in oligotrophic environments, or due to confusion with the widespread and more common *A. lineare*.

Studies on lakes and other lentic habitats have proven to be a valuable addition to the research on diatom biogeography and biodiversity which resulted in the description of a large number of freshwater diatoms species new to science or to a territory (e.g. Kulikovskiy et al. 2012; Potapova

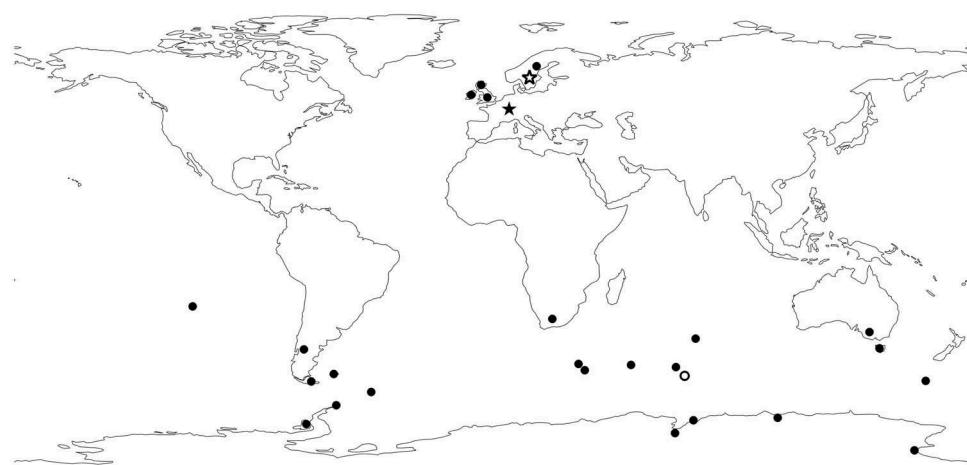


Figure 51. World geographic distribution of *Psammothidium abundans* and *Achnanthidium sublineare*. Unfilled circle indicates the type locality of *Psammothidium abundans* (Kerguelen Islands). Filled circles indicate bibliographical citations of *Psammothidium abundans* (see references in the text). White star indicates type locality of *Achnanthidium sublineare* (Semlan, Sweden). Black star indicates Longemer Lake (Xonrupt-Longemer, Vosges, France) where both *Achnanthidium sublineare* and *Psammothidium abundans* were found on July 2017.

2014; Kulikovskiy, Lange-Bertalot, and Kuznetsova 2015; Rondón and Catalan 2017). This is also true in north-eastern France (Heudre et al. 2015, 2017, 2018, 2019). Increasing survey of those environments will most likely reveal their remaining unexplored biodiversity potential.

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David Heudre is a hydrobiologist at the Direction Régionale de l'Environnement, de l'Aménagement et du Logement Grand Est. He has written several articles and has been working on diatoms, aquatic plants and algae in rivers and lakes. He has been teaching on bioassessment using diatoms. *Contribution:* Sample collection, LM photos, plates and writing of the article.

Carlos E. Wetzel is a botanist and a researcher at the Luxembourg Institute of Science and Technology. He has written over 90 articles and he has been working on diatoms in rivers, lakes and soils. Carlos E. Wetzel has been teaching numerous training courses on diatom ecology and taxonomy designed for biologists, technicians and ecologists, contributing to the continuous improvement in the Water Framework Directive implementation in Europe. *Contribution:* SEM photos, discussion of results and development of the manuscript.

Bart Van de Vijver is a full-time researcher at the Meise Botanic Garden, Belgium, and a part-time professor at the University of Antwerp, Belgium. His research focuses mainly on the taxonomy, morphology and biogeography of Antarctic freshwater and terrestrial diatoms. He has been studying non-marine diatoms in various parts of the Antarctic region for more than 20 years. He has described almost 400 new taxa and revised an additional 250 taxa. *Contribution:* identification of diatom species, discussion of results and assistance with the writing and editing of the manuscript.

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Luc Ector is a botanist and senior researcher at the Luxembourg Institute of Science and Technology. He has written over 185 articles and has been working on diatoms

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