

This item is the archived peer-reviewed author-version of:

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Reference:

Fu Yongshuo, Piao Shilong, Delpierre Nicolas, Hao Fanghua, Hänninen Heikki, Liu Yongjie, Sun Wenchao, Janssens Ivan, Campioli Matteo.- Larger temperature response of autumn leaf senescence than spring leaf-out phenology
Global change biology - ISSN 1354-1013 - 24:5(2018), p. 2159-2168
Full text (Publisher's DOI): <https://doi.org/10.1111/GCB.14021>
To cite this reference: <https://hdl.handle.net/10067/1479800151162165141>

1 **Larger temperature response of autumn leaf senescence than spring**
2 **leaf-out phenology**

3
4 **Running title:** temperature response of autumn leaf senescence
5

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21 **Keywords:** climate change, summer and autumn warming, leaf phenology, leaf senescence, leaf-
22 out, temperature sensitivity

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26 **Paper type:** Primary Research Articles
27

28 Manuscript for *Global Change Biology*

29 **Abstract**

30 Climate warming is substantially shifting the leaf phenological events of plants, and thereby
31 impacting on their individual fitness and also on the structure and functioning of ecosystems.
32 Previous studies have largely focused on the climate impact on spring phenology, and to date the
33 processes underlying leaf senescence and their associated environmental drivers remain poorly
34 understood. In this study, experiments with temperature gradients imposed during the summer
35 and autumn were conducted on saplings of European beech to explore the temperature responses
36 of leaf senescence. An additional warming experiment during winter enabled us to assess the
37 differences in temperature responses of spring leaf-out and leaf senescence. We found that
38 warming significantly delayed the dates of leaf senescence both during summer and autumn
39 warming, with similar temperature sensitivities (6 - 8 days delay per °C warming), suggesting
40 that, in the absence of water and nutrient limitation, temperature may be a dominant factor
41 controlling the leaf senescence in European beech. Interestingly, we found a significantly larger
42 temperature response of autumn leaf senescence than of spring leaf-out. This suggests a possible
43 larger contribution of delays in autumn senescence, than of the advancement in spring leaf-out,
44 to extending the growing season under future warmer conditions.

45 **Introduction**

46 Plant phenology is the study of periodic plant life cycle events, and how these are influenced by
47 seasonal and interannual variations in climate (Lieth, 2013). Plant phenology is one of the most
48 reliable biological indicators of anthropogenic climate change (Parmesan & Yohe, 2003, Root *et*
49 *al.*, 2003, Walther *et al.*, 2002), and changes in plant phenology impact on individual fitness,
50 species distribution, interspecific interactions, ecosystem structure and function, as well as on
51 feedbacks to the climate system (Chuine *et al.*, 2010, Peñuelas & Filella, 2009, Piao *et al.*, 2007,
52 Thackeray *et al.*, 2016, Zeng *et al.*, 2017). Therefore, understanding the processes underlying
53 plant phenology is essential to improve our understanding of plant and ecosystem responses to
54 the ongoing climate change.

55

56 Plant phenology research has grown tremendously over the past four decades (Hänninen, 2016,
57 IPCC, 2014, Wolkovich & Ettinger, 2014). Most studies, however, have addressed spring
58 phenological events, such as budburst, leaf-out and flowering (Fu *et al.*, 2015, Richardson *et al.*,
59 2013), while autumn phenological events, such as leaf senescence, have been paid comparatively
60 less attention (Gallinat *et al.*, 2015, Panchen *et al.*, 2015). One probable reason for this is the
61 difficulty to accurately acquire leaf senescence observations in natural conditions. However, as
62 the final stage of the leaf's life cycle and as adaptive strategy to unfavorable environmental
63 conditions of temperate and boreal plant species (Chabot & Hicks, 1982), leaf senescence is
64 critical to plants' fitness as well as to ecosystem functions (Estiarte & Peñuelas, 2015, Piao *et al.*,
65 2008, Rohde & Bhalerao, 2007).

66

67 The main function of autumn leaf senescence is to recycle nutrients from senescing leaves and
68 transport them to other plant tissues to support growth during the following spring (Chapin III,

69 1980, Killingbeck, 1996, Maillard *et al.*, 2015). This nutrient-conservation mechanism increases
70 the fitness of individual plants, especially in nutrient poor environments (Chapin III, 1980, May
71 & Killingbeck, 1992). Generally, more than half of the leaf macro-nutrients, such as N and P,
72 are being resorbed during the leaf senescence process, although the nutrient resorption efficiency
73 varies widely among species and elements (Aerts, 1996, Freschet *et al.*, 2010, Wright &
74 Westoby, 2003). Apart from its influence on nutrient cycles, the timing of leaf senescence
75 influences the ecosystem carbon balance by modulating the length of the photosynthetically
76 active period (Myneni *et al.*, 1997, Richardson *et al.*, 2010). Leaf senescence may even play a
77 more critical role than spring phenology in determining the length of the photosynthetically
78 active period (Garonna *et al.*, 2014, Wu *et al.*, 2013). Understanding the response of leaf
79 senescence to climate change is therefore important. However, to date, the processes underlying
80 autumn leaf senescence, their associated environmental controls and the response of leaf
81 senescence to climate change are still poorly understood.

82

83 Photoperiod has generally been proposed as the primary driver of leaf senescence, with a critical
84 photoperiodic threshold, i.e., a critical day length below which leaf senescence is triggered
85 (Lagercrantz, 2009, Wareing, 1956, Way & Montgomery, 2015, Welling & Palva, 2006). For
86 example, the autumnal senescence in mature European aspen occurs every year on almost the
87 same date (Fracheboud *et al.*, 2009). However, photoperiod is not consistently important across
88 species and even sites. For example, a recent study reported that leaf senescence is not triggered
89 by photoperiod across 116 European aspen natural genotypes (Michelson *et al.*, 2017),
90 suggesting that other environmental factors must be involved in driving the leaf senescence
91 process. In line with this finding, many studies have suggested that temperature interacts with

92 photoperiod to control the leaf senescence process (Chung *et al.*, 2013, Hänninen & Tanino,
93 2011, Heide & Prestrud, 2005, Liu *et al.*, 2016a, Tanino *et al.*, 2010), and that temperature can
94 be even the main controlling factor of leaf senescence (Chmielewski & Rötzer, 2001, Estiarte &
95 Peñuelas, 2015, Heide & Prestrud, 2005, Xie *et al.*, 2015), in particular autumn temperature
96 (Delpierre *et al.*, 2009). However, these results are not conclusive because other studies have
97 shown that autumnal senescence is only weakly affected by air temperature (Čufar *et al.*, 2012,
98 Olsson & Jönsson, 2015, Sparks & Menzel, 2002). In addition to photoperiod and temperature,
99 also other environmental factors have been reported to influence the leaf senescence process.
100 These include light intensity (Liu *et al.*, 2016b), precipitation and soil water conditions (see
101 review in Estiarte and Peñuelas (2015)) and soil nutrient status (Sigurdsson, 2001, Weih, 2009).
102 Furthermore, a positive correlation was recently reported between spring leaf-out dates and
103 autumn leaf senescence dates, suggesting that the factors regulating the leaf-out days are carried
104 over to affect leaf senescence in the following autumn (Fu *et al.*, 2014, Signarbieux *et al.*, 2017).
105 Thus, the literature remains inconsistent about the determinants of autumn leaf senescence
106 (Estiarte & Peñuelas, 2015), so that well-designed experiments are needed to investigate and
107 better understand the leaf senescence process.

108
109 Current studies of leaf senescence are generally based on either species-specific long-term in situ
110 observations (Menzel *et al.*, 2006, Panchen *et al.*, 2015), or on remote-sensing based
111 observations (Garonna *et al.*, 2014, Julien & Sobrino, 2009, Liu *et al.*, 2016b, Shen *et al.*, 2015,
112 Xie *et al.*, 2015, Jeong *et al.*, 2011). While manipulation experiments have been conducted, only
113 few have studied the autumn phase in relation to climate change, as opposed to spring
114 (Wolkovich *et al.*, 2012). Furthermore, these few-experimental autumn phenology studies were

115 designed with only one or two warming treatments (Gunderson *et al.*, 2012, Marchin *et al.*,
116 2015, Morin *et al.*, 2010, Norby *et al.*, 2003). How leaf senescence responds to a temperature
117 gradient, whether summer and autumn warming influence leaf senescence differently, and
118 whether autumn phenology has the same temperature sensitivity as spring phenology, to our
119 knowledge, has not yet been experimentally investigated.

120

121 In the present study, we therefore carried out gradient-warming/cooling manipulation
122 experiments using two-year old and one-meter-high saplings of *Fagus sylvatica* L. (European
123 beech), a widespread deciduous forest tree species in temperate Europe. Specifically, we exposed
124 the saplings to either summer or autumn warming. The primary objectives of this study were (1)
125 to quantify the temperature sensitivity of leaf senescence date (St, changes in days per °C
126 warming); (2) to investigate the effect of summer versus autumn warming on leaf senescence
127 dates, and (3) to compare St of autumn senescence with that of spring leaf-out.

128

129 **Materials and methods**

130 **Study site and climate chambers**

131 The experiment was conducted in 12 climate-controlled transparent chambers at the Drie Eiken
132 campus of the University of Antwerp (Belgium, 51°19'N, 4°21'E). The long-term mean annual
133 air temperature is 9.6 °C, and mean monthly air temperatures vary from 2.2 °C in January to 17.0
134 °C in July. Annual precipitation averages 780 mm, being uniformly distributed throughout the
135 year (Campioli *et al.*, 2012). The chambers could be artificially warmed/cooled by a centralized
136 heating/cooling system ensuring different levels of continuous (day and night) warming or
137 cooling above/below the fluctuating ambient air temperature (Fu *et al.*, 2016). Each chamber

138 could accommodate 12 saplings. Temperature sensors (Siemens, type QFA66, Berlin, Germany)
139 were used to continuously monitor the air temperature inside each chamber, logging every 30
140 minutes and storing hourly data. Here, we combine the results from three different experiments
141 performed in the climate chambers using beech saplings of the same provenance and size.

142

143 **Experimental design and leaf senescence measurements**

144 *Tree material.* The experiments used 2-year-old and one-meter-high saplings of *Fagus sylvatica*
145 L. grown from seeds of the same origin and cultivated in the same field for one year at a
146 commercial nursery. We transplanted the saplings into plastic pots (diameter 25 cm, depth 40
147 cm). The pots were moved into the climate-controlled chambers during early summer, late
148 summer or winter (see below). The pots were filled with a substrate that was created by
149 combining potting soil and Lommel sand (grain size <1 mm diameter), bought from commercial
150 suppliers (Van den Broeck and Jos Meeussen & Zoon bvba) in Belgium. In experiment 1 and 2,
151 slow release fertilizer (100 g·m⁻², 13–10–20 for N, P, and K, respectively, all in percentage) was
152 added in end of May to each sapling. Over the growing period, the saplings were watered at least
153 three times per week to ensure no water limitation.

154

155 *Experiment 1 (temperature treatment during summer, targeting leaf senescence)* (Fig. 1). In this
156 experiment, one control treatment (+0°C) and three temperature treatments were applied: +1°C,
157 +3°C and +4°C. Two to four replicate chambers were used, except for the +4°C treatment, for
158 which data from only one chamber were available because one chamber failed. At summer
159 solstice (21 June 2016), four saplings were placed in each chamber. The saplings were moved

160 out at the “end of summer”, i.e., on 15 August 2016. In total, 9 chambers and 36 saplings were
161 used in this experiment.

162

163 *Experiment 2 (temperature treatment during autumn, targeting leaf senescence)* (Fig. 1). In this
164 experiment, we applied one control treatment (+0°C), one cooling treatment (-1°C) and one
165 warming treatment (+1°C), using three replicated chambers for each treatment. Four saplings
166 were exposed to the treatment in each chamber during the autumn period, i.e. from 15 Aug 2016
167 to leaf senescence. In total, 9 chambers and 36 saplings were used in this experiment.

168

169 *Experiment 3 (temperature treatment during winter-spring, targeting leaf-out)*. In winter-spring
170 2016, we conducted a separate warming experiment on the spring leaf-out phenology. Four
171 saplings were moved into each of the twelve climate-controlled chambers on 1 January 2016.
172 During the experiment, five warming temperature treatments, i.e. +1°C, +2°C, +3°C, +4°C and
173 +5°C (two chambers per treatment) and a control treatment (+0°C) were applied. In total, 12
174 chambers and 48 saplings were used in this experiment.

175

176 The warming/cooling provided was generally stable for the experiments 1 and 3 (actual warming
177 was on average $\pm 10\%$ of the prescribed value; see below for description of the experiments).
178 However, for experiment 2 (see below) the warming during autumn was less stable (within \pm
179 20%). This inaccuracy was not considered crucial, because our main analyses were based on the
180 actually realized warming/cooling (e.g. regression analysis, see below), not the envisaged
181 warming.

182

183 **Observation of leaf senescence and leaf-out**

184 Leaf senescence was monitored following Vitasse *et al.* (2009). In detail, the number of leaves
185 with autumn colour was determined visually and the percentage of them, out of the total number
186 of leaves, was calculated. The senescence date was defined as the date when 50% of the leaves
187 had autumn colour. Leaf-out date was defined as the day when the entire leaf blade and the leaf
188 stalk were visible on the terminal buds (Fu *et al.*, 2016).

189

190 **Cooling degree hours**

191 To evaluate the relationships between leaf senescence and air temperature, we calculated cooling
192 degree hours (CDH) as the sum of hourly temperature below a temperature threshold (T_{base})
193 during the study period, i.e., from summer solstice (21 June) to the day of leaf senescence (LS)
194 (Dufrêne *et al.*, 2005). The T_{base} was set at 25°C, according to a previous study on *Fagus*
195 *sylvatica* (Delpierre *et al.*, 2009):

196
$$CDH = \sum_{start}^{LS} (T_{base} - T_{hourly}) \text{ if } T_{hourly} < T_{base}$$

197 We also tested the use of a higher T_{base} , i.e. 30°C, but found very similar results, and therefore
198 only reported the results based on 25°C in the main text.

199 **Data analysis**

200 The temperature responses of leaf senescence and leaf-out were evaluated using linear regression
201 based on the average dates obtained from the four saplings per chamber. The temperature
202 sensitivity of leaf senescence and leaf-out were defined as the slopes of the linear regression
203 between dates and the actual temperature change in the chambers. Independent samples *t*-tests
204 were used to evaluate the difference between leaf senescence, or leaf-out dates, as well as the

205 accumulated CDH, among different temperature treatments. Differences in the temperature
206 sensitivity of leaf senescence between autumn cooling and autumn warming, and between
207 summer and autumn, as well as in the temperature sensitivity between leaf senescence and leaf-
208 out, were tested using ANCOVA, i.e. testing the slopes and intercepts. All statistical analyses
209 were conducted using SPSS 16.0 (SPSS Inc., Chicago, IL, USA).

210

211 **Results**

212 **Leaf senescence response to experimental warming and cooling**

213 Leaf senescence dates were significantly delayed by the warming treatments, but were
214 significantly advanced by cooling (both $P < 0.05$), as compared to the control treatment (Fig. 1a).
215 Although both summer and autumn warming significantly delayed the leaf senescence dates,
216 compared to the control, a larger delay (14 days on average) was found under summer warming
217 treatments than under autumn warming treatments (11 days on average) (Fig. 1a). As opposed to
218 the warming treatments, cooling significantly advanced the dates of leaf senescence, by 3 days
219 on average ($F=9.8$; $P=0.005$; Fig. 1a).

220

221 **Temperature sensitivity of leaf senescence and leaf-out**

222 In the previous paragraphs the phenology responses were given, independent of the intensity of
223 the warming or cooling. This paragraph aims to render these treatments more comparable by
224 expressing all phenology changes on a per °C basis. By using a relative variable, we aim to
225 removing the influence of different periods (summer vs. autumn) and exposure times to warming.
226 On average, a rise of air temperature by one degree delayed the leaf senescence date by 6.4 ± 1.1
227 days, and the difference in the temperature sensitivity of leaf senescence (St, delay in days per °C

228 warming) between autumn warming treatment (St_{autumn} , 8.3 ± 1.1 days $^{\circ}\text{C}^{-1}$) and summer
229 warming treatment (St_{summer} , 6.1 ± 0.8 days $^{\circ}\text{C}^{-1}$) was not statistically significant ($P=0.75$,
230 Fig. 1b). The absolute St values of leaf senescence during autumn warming and autumn cooling
231 (-6.7 ± 1.0 days $^{\circ}\text{C}^{-1}$ for cooling treatments) also did not differ statistically significantly (Fig. 2).
232
233 Warming significantly advanced the date of leaf-out in spring, with 4.5 ± 0.5 days advancement
234 per degree Celsius warming (Fig. 3a). Compared to the autumn leaf senescence (delay of $8.3 \pm$
235 1.1 days $^{\circ}\text{C}^{-1}$), the temperature sensitivity of leaf-out was thus significantly lower (Fig. 3b),
236 suggesting a larger effect of climate warming on autumn leaf senescence than on spring leaf-out
237 phenology.

238

239 **Correlation between leaf senescence and cooling degree hours**

240 No significant correlation was found between leaf senescence dates and CDH that were
241 accumulated from 21 June to the day of leaf senescence across the temperature treatments
242 ($R^2=0.09$, $P=0.12$, Fig. 4). In addition, the CDH requirement was not significantly different
243 between the two autumn treatments, i.e., 14677K and 13067K for autumn warming treatment
244 and autumn cooling treatment, respectively. However, the CDH requirement of saplings in the
245 summer warming treatment was statistically significantly lower than in the autumn warming and
246 autumn cooling treatments ($P<0.05$).

247

248 **Discussion**

249 Previous studies have highlighted the ambiguous nature of the warming response of leaf
250 senescence (Gunderson *et al.*, 2012, Heide & Prestrud, 2005, Menzel *et al.*, 2006), and

251 attributed this to the limited availability of long-term datasets, the difficulty of quantifying the
252 exact date of leaf senescence under natural conditions, and the lack of focused experimental
253 studies designed to understand the leaf senescence response to temperature. For temperate trees
254 under favorable conditions, i.e., without water or nutrient stress, it is generally assumed that the
255 leaf senescence process is mainly triggered by photoperiod and temperature (Lieth, 2013, Way &
256 Montgomery, 2015). Some studies reported a delayed trend of leaf senescence with climate
257 warming, based on in situ observations (Delpierre *et al.*, 2009, Vitasse *et al.*, 2011), remote
258 sensing observations (Liu *et al.*, 2016b, Reed *et al.*, 2009, Stöckli & Vidale, 2004), as well as
259 open top chamber-based field warming experiments (Gunderson *et al.*, 2012). In contrast, other
260 studies reported insignificant responses or even advanced senescence with climate warming
261 (Norby *et al.*, 2003, Xie *et al.*, 2015), which may be related to warming-induced drought stress
262 (Xie *et al.*, 2015). Based on gradient warming experiments, in which drought was excluded, our
263 study clearly revealed that warming significantly delays the timing of leaf senescence in
264 European beech saplings in both summer and autumn warming, with even more than 30 days
265 delay under the +4°C treatment (4°C higher than ambient). This suggested that, under sufficient
266 water and nutrient conditions, temperature may be more important than photoperiod in
267 controlling the leaf senescence process under temperate latitudes. In fact, if there was a
268 photoperiod threshold, this would have been overpassed by up to 30 days. Note that we found a
269 larger delay (14 days on average) under summer warming treatments than under autumn
270 warming treatments, this was, however, maybe largely attributed to the more intensive warming
271 treatment that was applied during summer (warmed up to ca. 4°C) than during autumn (warmed
272 up to ca. 1°C) (see Fig. 1b). We also did not find a statistical difference of the temperature
273 sensitivity of leaf senescence between autumn warming and autumn cooling treatments. This

274 may be due to the limited sample size in this study, i.e. only one cooling treatment and one
275 warming treatment during autumn, and therefore this conclusion needs to be further studied. In
276 addition, we found a larger temperature sensitivity of leaf senescence, with 6-8 days delay per
277 degree Celsius warming, as opposed to 2-7 days delay per °C warming found in earlier
278 experimental studies (Gunderson *et al.*, 2012, Han *et al.*, 2014, Nakamura *et al.*, 2010). This
279 difference might be related to species differences, to differences in the local environment, as well
280 as to the different experimental designs, i.e., only one or two warming treatments in the previous
281 studies versus gradient warming/cooling in the present study.

282

283 Surprisingly, there were no statistically significant differences in the sensitivity of the leaf
284 senescence process to summer and autumn warming. This implies that the positive impacts of
285 warming on leaf physiology, such as delayed chlorophyll degradation (Fracheboud *et al.*, 2009),
286 leading to delayed leaf senescence at the end of the growing period, does not depend on the
287 seasonal timing of the temperature elevation. Nonetheless, warming may affect different
288 processes during summer (e.g. predominantly cell division and expansion) than during autumn
289 (cell maturation and lignification). Furthermore, warming might affect different phases of the
290 leaf senescence process when applied in summer versus autumn. During summer, warming
291 might delay the leaf senescence onset, whereas autumn warming might slow down the
292 progression rate of the leaf senescence (Fracheboud *et al.*, 2009). These different aspects (delay
293 in leaf senescence onset vs. slowdown of leaf senescence rate) cannot be independently assessed
294 with the coloration method we used.

295

296 The lower CDH requirement associated with the summer treatment should be related to more
297 intense warming in summer than in autumn. We do not believe that the low CDH requirement in
298 summer is related to differences in the leaf senescence date as summer warming elicited, on
299 average, later leaf senescence than autumn warming, which should have caused a larger CDH.
300 The timing of leaf senescence simulated by cooling degree days-based models has been
301 compared in earlier studies with in situ observations (Archetti *et al.*, 2013, Delpierre *et al.*, 2009,
302 Jolly *et al.*, 2005, Vitasse *et al.*, 2011). Consistent with the assumption of degree days-based
303 models, we found that the differences in the cooling degree hours (CDH) required for leaf
304 senescence between the two autumn treatments were statistically insignificant. Furthermore,
305 good model performances were found in boreal tree species (Koski & Selkäinaho, 1985,
306 Partanen, 2004, Viherä-Aarnio *et al.*, 2005). However, contrary to the results of the present study,
307 these studies found that warming during summer and autumn would advance, not delay, the
308 timing of leaf senescence. Possibly, this opposite temperature response is attributable to
309 differences among boreal and temperate-zone species. Jeong and Medvigy (2014) reported a
310 nonlinear temperature sensitivity of leaf senescence using many ground observations and
311 suggested that warmer regions may have a larger temperature sensitivity than cooler regions. In
312 addition, recent studies have reported a positive correlation between spring leaf-out and leaf
313 senescence dates in trees (Fu *et al.*, 2014, Signarbieux *et al.*, 2017), delayed senescence
314 following exceptionally late spring greening in sub-arctic grasslands (Leblans *et al.*, 2017), and
315 the performance of senescence models was substantially improved by incorporating this legacy
316 effect.
317

318 Interestingly, we found a larger temperature sensitivity (St) of autumn leaf senescence than
319 spring leaf-out using European beech saplings of the same age. Contrasting conclusions were
320 obtained from a meta-analysis of observations on mature trees from the European phenology
321 network, which reported a larger St of spring leaf-out (4.6 ± 0.07 days $^{\circ}\text{C}^{-1}$) than of autumn leaf
322 colouring (1.0 ± 0.4 days $^{\circ}\text{C}^{-1}$) across plant species (Menzel *et al.*, 2006). This difference may be
323 attributable to the species-specific differences in the phenology response to temperature
324 (Panchen *et al.*, 2015, Richardson *et al.*, 2006, Vitasse *et al.*, 2009). However, similar finding
325 was reported on *Quercus variabilis* seedlings in an open-field warming experiment (Han *et al.*,
326 2014), and on mature beech trees in an altitude gradient (Vitasse *et al.*, 2009). The difference in
327 temperature sensitivity of leaf-out and leaf senescence are likely related to the differential
328 processes between spring and autumn phenology. Concerning spring leaf-out (particularly for
329 diffuse porous species like beech), temperature impacts the end of bud dormancy and the speed
330 of leaf unfolding but no other trees organ (which are inactive before budburst) (Delpierre *et al.*,
331 2016). On the other hand, in autumn, temperature impacts both the leaf physiological status
332 (chlorophyll content, photosynthesis, pigment degradation etc) and tree growth (e.g. wood
333 lignification, fine root growth). The latter reduction of carbon sink activity at the tree scale may
334 be an additional, overlooked trigger of leaf senescence. These interactions might affect leaf
335 senescence onset, in other words not only leaf status but also (and maybe primarily) a lack of
336 sink activity might trigger leaf senescence (see hypothesis in Fu *et al.*, 2014). Therefore, it is
337 logical that temperature has a strong effect on autumn phenology, which, as showed by our data,
338 and even can be stronger than that on spring phenology.

339

340 Given the larger warming response of leaf senescence than of spring leaf-out found in the present
341 study, under future climate warming conditions we can expect a larger contribution of the delay
342 in autumn senescence dates to the extension of photosynthetic season than of the earlier spring
343 leaf-out. Thus, warming induced changes in leaf senescence could play an important role in the
344 ecosystem carbon balance (Keenan *et al.*, 2014, Piao *et al.*, 2008). However, delayed leaf
345 senescence in response to warmer summers-autumns may increase the risk of extreme events
346 such early-frost damage to leaves (Augspurger, 2013, Hänninen, 2016, Inouye, 2008), which
347 would hamper the nutrient resorption. This can lead to reduced nutrient reserves to support next
348 season's growth, and subsequently impact the ecosystem carbon and nutrient cycles (Estiarte
349 & Peñuelas, 2015, Fracheboud *et al.*, 2009). Finally, note that considering the legacy effect of
350 leaf-out on the leaf senescence dates (Fu *et al.*, 2014), the delays in leaf senescence as observed
351 in our experiments might be partially offset by the earlier spring leaf-out in response to warmer
352 winters.

353

354 The underlying physiological processes of leaf senescence and their environmental cues,
355 especially the interactive effect of temperature and photoperiod, are still unclear. Moreover,
356 warming responses of leaf senescence largely differ between natural observations and warming
357 experiments (Wolkovich *et al.*, 2012), ontogenetic differences have been reported between
358 saplings and mature trees (Mediavilla *et al.*, 2014, Vitasse, 2013), and species variability exists
359 in response to warming (Parmesan & Hanley, 2015, Primack *et al.*, 2015). Nonetheless, our
360 study provides important insights. Taking advantage of temperature manipulative experiments,
361 we found that, in the absence of water and nutrient limitation, temperature is a dominant factor
362 controlling the leaf senescence process in European beech, and warming during summer and

363 autumn both significantly delay the date of leaf senescence. Furthermore, we found a larger
364 temperature response of leaf senescence than spring leaf-out. These findings enhance our
365 understanding of leaf phenology response to the climate change, and potentially improve our
366 understanding of phenological impacts on ecosystem carbon and nutrient cycles.

367

368 **Acknowledgments**

369 This study is supported by the General program of National Nature science foundation of China
370 (No. 31770516), the National Key Research and Development Program of China
371 (2017YFA06036001) and the Thousands Talents Program for Young Professionals. Matteo
372 Campioli is a Postdoctoral Fellow of the Research Foundation–Flanders (FWO) and
373 acknowledges ERC for support through ERC-2016-StG-714916 “LEAF-FALL”. Ivan A
374 Janssens acknowledges support from the European Research Council (ERC) through Synergy
375 grant ERC-2013-SyG-610028 “P-IMBALANCE” and support from the University Of Antwerp.

376

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591

592 Figure captions

593

594 Fig 1. (a) The distribution, mean, and standard deviation (plot box) of the leaf senescence dates
595 of European beech saplings under different temperature manipulations and the control. Each grey
596 dot indicates the result for one sapling. (b) Relationship between leaf senescence dates of
597 European beech saplings and the mean temperature change in the treatments, as compared with
598 the ambient temperature. Open circles: Experiment 1, i.e. temperature treatment over the summer
599 period (from summer solstice to 15 August 2016); Grey squares: Experiment 2, temperature
600 treatment over the autumn period (from 15 August 2016 to the date of leaf senescence). The grey
601 line and shaded areas represent linear regression fits (with 95% confidence intervals) across
602 summer and autumn treatments.

603

604 Fig 2. Temperature sensitivities of leaf senescence to autumn (from 15 August 2016 to the date
605 of leaf senescence) cooling and warming. The temperature sensitivity was calculated using
606 simple linear regression. The difference in the sensitivity between autumn cooling and warming
607 is not statistically significant

608 .

609 Fig 3. (a) Experiment 3: temperature treatment during winter-spring 2016. The temperature
610 sensitivity of leaf-out, in relation to the mean temperature change in the treatments, as compared
611 with the ambient temperature. (b) A comparison of the temperature sensitivity of leaf-out and
612 leaf senescence. The asterisk indicates a statistically significant difference ($P < 0.05$).

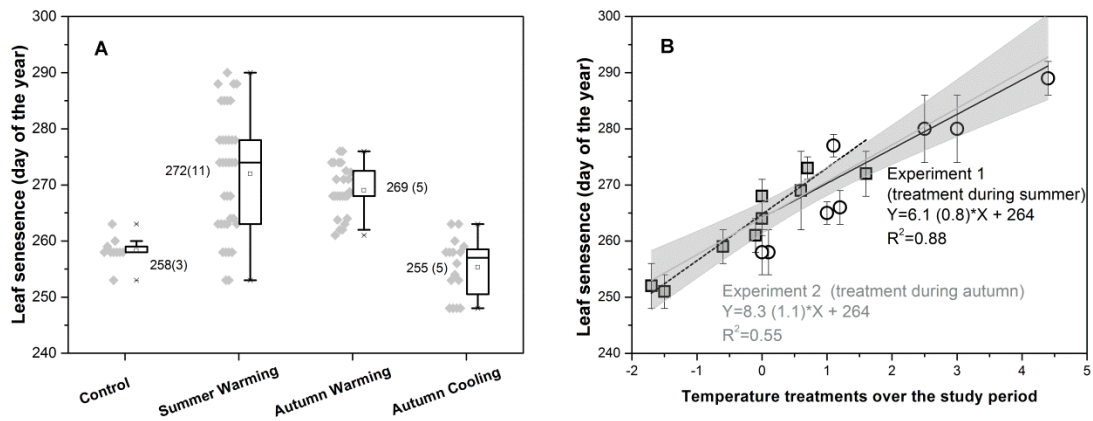
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614 Fig 4. Correlation between leaf senescence dates and cooling degree hours accumulated in the
615 different treatments. Each dot denotes one climate-controlled chamber. The color bar shows the

616 temperature anomalies in the temperature treatments (blue is control). Treatment of summer
617 warming (squares) refers to the period from summer solstice to 15 August 2016; autumn
618 warming (circles) and autumn cooling (diamonds) refer to the period from 15 August 2016 to the
619 date of leaf senescence.
620

621 FIGURE 1

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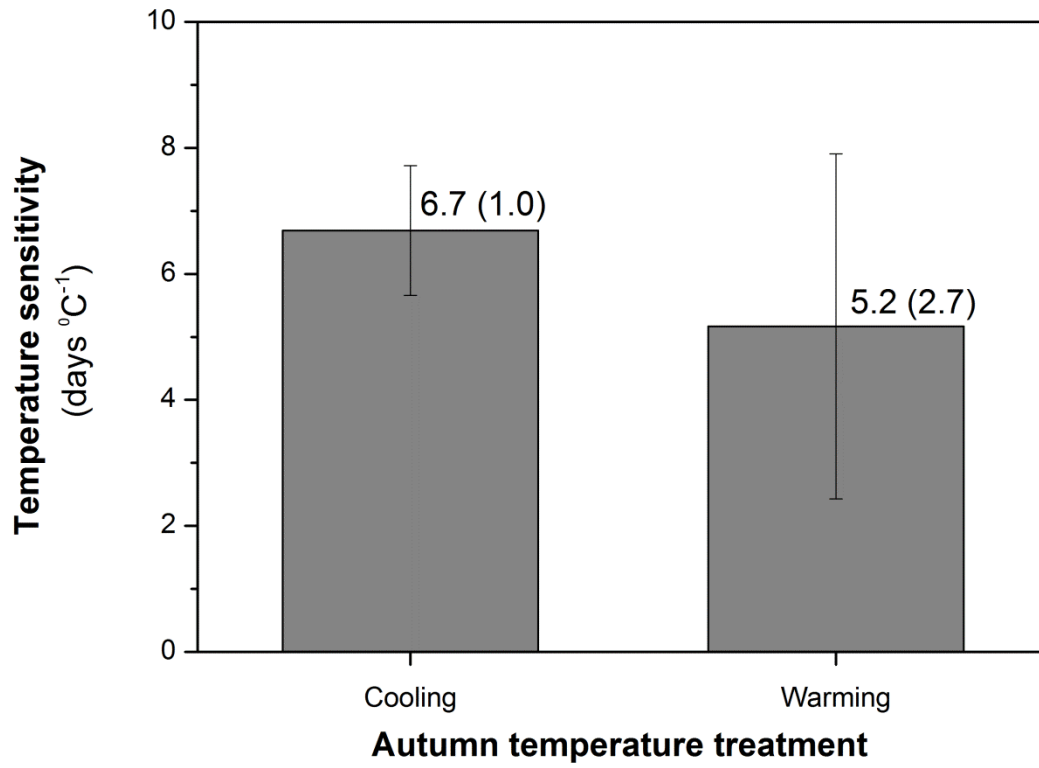
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626 FIGURE 2

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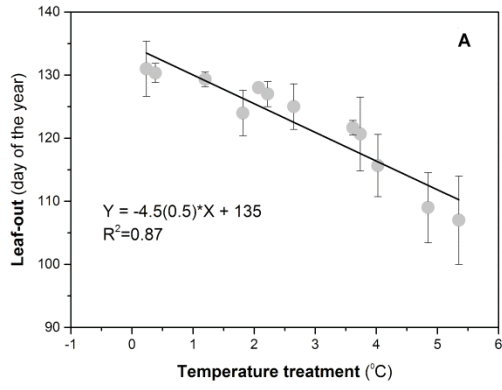


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630 FIGURE 3

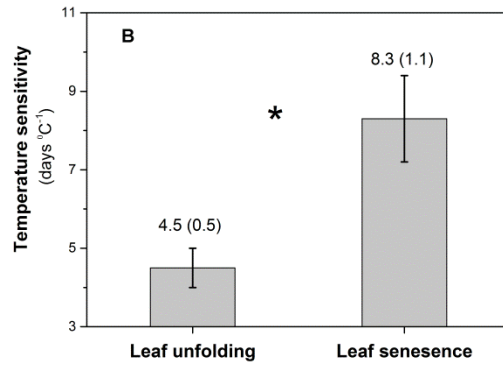
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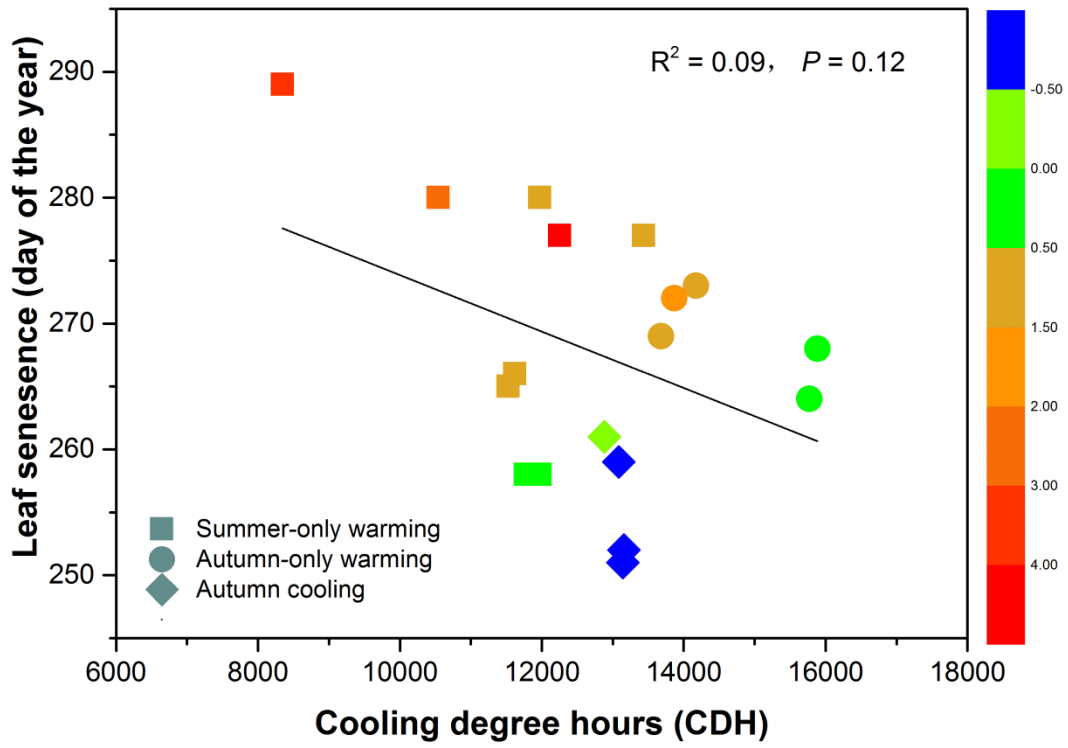
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635 FIGURE 4

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