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Opposing effects of aquatic vegetation on hydraulic functioning and transport of dissolved and organic particulate matter in a lowland river : a field experiment

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# 19 Abstract

20 The presence of instream aquatic vegetation (macrophytes) has an impact on the ecological functioning of 21 rivers through their effects on transport and retention of dissolved and particulate matter, and also on the 22 hydraulic functioning of rivers by increasing the hydraulic resistance, which results in higher water levels 23 and may induce an increased flooding risk. In order to unravel these opposing effects, two field studies 24 were conducted in 2013 and 2014 in a lowland river reach of 50 m with a high initial vegetation cover 25 (>76 %). We quantified the effects of three treatments – initial vegetation, partially mowed and vegetation 26 free - on the hydraulic functioning (hydraulic resistance) and ecological functioning (transport and 27 retention of dissolved and particulate tracers).

28 Firstly, the partially vegetated treatment (after partial vegetation removal) resulted in reduced hydraulic 29 resistance compared to the vegetated treatment and in enlarged retention of particulate matter compared to 30 the vegetation free treatments. The longitudinal dispersion and transient storage zones were similar to the 31 vegetated treatment. Moreover, the most heterogeneous flow field was also found in these partially 32 vegetated treatments. Secondly, the vegetation free treatments (after complete vegetation removal) had 33 the lowest hydraulic resistance, the highest flow velocity, the highest longitudinal dispersion coefficient, 34 the largest transient storage zone, and the lowest retention of particulate matter. Thirdly, vegetated 35 treatments had the highest hydraulic resistance, the lowest flow velocity, the lowest longitudinal dispersion coefficient, smallest transient storage zone, and the highest retention for particulate organic 36 37 matter.

We conclude that partial removal of the vegetation leads to an optimal trade-off between minimizing the flow velocity and maximizing the retention of particulate organic matter while minimizing the hydraulic resistance compared to the fully vegetated and vegetation free treatment.

42 **Keywords**: macrophytes, hydraulic resistance, longitudinal dispersion, transient storage

# 43 Highlights

- Tracer experiments matter are performed with dissolved and particulate
- The effect of vegetation covers investigated in situ at river reach scale
- Plants reduce the longitudinal dispersion coefficient and transient storage zone
- The most heterogeneous flow field is found when part of the vegetation is removed

# 48 **1. Introduction**

49 The hydraulic and ecological functioning of lowland rivers is influenced to a great extent by instream 50 aquatic vegetation (Newbold et al. 1982; Runkel 2007). The presence of macrophytes leads to reduced 51 flow conveyance, higher water levels, decreased stream velocities, and enhanced sediment deposition on 52 the river bed (Old et al. 2014). Therefore macrophytes are often mechanically removed to increase flow 53 conveyance and reduce flooding risk (Boerema et al. 2014; Lopez and Garcia 2001). The vegetation can 54 either be completely removed (Old et al. 2014) or partly (Bal et al. 2011; Vereecken et al. 2006). Changes 55 to the hydraulics directly affect the ecological functioning of lowland rivers (Hensley and Cohen 2012) 56 through its effects on the transport and retention of dissolved (Wilcock et al. 1999) and particulate matter 57 (Horvath 2004; Warren et al. 2009).

58 Nutrient cycling of dissolved matter is influenced by both hydraulic transport processes (advection, 59 dispersion, inflow, transient storage) and non-hydraulic processes (uptake rates, biomass standing stock, 60 temperature) (Runkel 2007). The hydraulic transport processes can be separated into three processes: (i) 61 advection, which is the transport by the bulk motion of the water flow; (ii) dispersion, which is the 62 combination of molecular or turbulent diffusion and of three dimensional processes, leading to shear flow 63 separation and enhancing the dispersion (Taylor 1954); and (iii) transient storage, which is the temporary 64 retention and release of molecules in certain transient storage zones within the river system (Bencala and Walters 1983; Jackman et al. 1984; Pedersen 1977; Thankston and Schnelle 1970). One or multiple 65 66 transient storage zones can be present which can be linked in serial or in parallel to the main channel 67 (Hensley and Cohen 2012). Transient storage zones are regions with low to zero flow velocity, and the exchange of dissolved matter with the main flow is driven by the concentration difference in the main 68 69 channel and within the transient storage zone (Gonzalez-Pinzon et al. 2013). In a one dimensional 70 approach, these three processes are cross-sectionally averaged and can be described by a longitudinal 71 dispersion-advection model with transient storage (Czernuszenko and Rowinski 1997).

72 The hydraulic transport processes can be quantified in river reaches through the use of conservative dissolved tracers. A dissolved conservative tracer is injected upstream of a river reach and its 73 74 concentration in function of time is recorded at the downstream end of this reach to obtain time series 75 (Das et al. 2002; Govindaraju and Das 2002). Temporal moments of these time series can be used to 76 parametrize the coefficients of the longitudinal dispersion-advection model with transient storage 77 (Czernuszenko and Rowinski 1997; Nash 1959). The first, second and third temporal moment can also be 78 used to investigate the transport and mixing properties in rivers: (i) the first temporal moment is linked 79 with the mean travel time of the tracer through the reach; (ii) the second temporal moment is the variance 80 and is associated with the longitudinal dispersion of the tracer; and (iii) the third temporal moment 81 characterizes the skewness and is related to the magnitude of the transient storage zone (Lees et al. 2000; 82 Sukhodolova et al. 2006). Multiple transport and mixing processes are acting simultaneously in rivers, so 83 the first three temporal moments are strongly linked with each other. A constant relationship between the 84 second and third normalized temporal moment was found in an extensive meta-analysis of 384 tracer 85 experiments conducted over a large range of discharges (7 orders of magnitude) and river lengths (5 86 orders of magnitude) (Gonzalez-Pinzon et al. 2013). However, the effect of instream vegetation was not 87 considered.

88 It may be expected that instream vegetation can affect each of the three aforementioned processes. First, 89 vegetation increases hydraulic resistance, hence reducing flow velocities and increasing water depth (De 90 Doncker et al. 2009b; Franklin et al. 2008), which will affect advection of dissolved matter. Lower flow 91 velocities will in turn increase the residence time which is beneficial for the water quality. For example 92 the denitrification is positively correlated with the residence time (Seitzinger et al. 2006). Second, the 93 influence of vegetation on longitudinal dispersion is less clear. Macrophytes may enhance turbulence and 94 diminish the vertical shear stress, resulting in a decreased longitudinal dispersion (Nepf et al. 1997; 95 Wilcock et al. 1999). However, the longitudinal dispersion may also increase by enhanced mechanical

96 dispersion (Nepf et al. 1997). The latter is a known phenomenon in porous media in which each particle 97 follows its own route, with a different length, through a network of pores. Third, transient storage zones 98 can be present as wake zones behind the vegetation stems (Nepf et al. 1997), within and behind dense 99 vegetation patches in the main channel (Sukhodolova et al. 2006) or riparian vegetation along the banks 100 (Wilcock et al. 1999). The net result of macrophytes on the transient storage zone is therefore difficult to 101 predict.

102 The potential effects of instream aquatic vegetation on the transport and retention of organic solid 103 particles is expected to be twofold: (i) by creating a sieve-like structure in the water column the particles 104 are physically trapped by both leaves and organisms living on the plants (Cotton et al. 2006; Pluntke and 105 Kozerski 2003), and (ii) by increasing the hydraulic resistance and reducing the flow velocity the 106 residence time and settling chance of the particles is increased (Folkard 2011). Cordova et al. (2008) 107 investigated the transport of coarse particulate organic matter (CPOM) in lowland rivers. They found that 108 approximately 50-83 % of the particle transport could be explained by particle settling, while the 109 remaining part could be explained by particle trapping on the plant surface. Besides discharge (Defina and 110 Peruzzo 2010), particle trapping depends on vegetation properties: increased submerged vegetation cover 111 increases the retention of particles (Riis and Sand-Jensen 2006), yet the configuration of the vegetation 112 does not affect the retention of particles (Defina and Peruzzo 2010). It also depends on the particle 113 properties: larger particles have a higher chance to be trapped (Ehrman and Lamberti 1992) and highly 114 buoyant particles have a higher potential travel distance (Boedeltje et al. 2004; Danvind and Nilsson 115 1997; Riis and Sand-Jensen 2006; van den Broek et al. 2005). The second process, particle settling, is 116 well studied for mineral particles (Church 2006; Wood and Armitage 1997) and is determined by the 117 settling velocity (Dietrich 1982). This velocity is proportional to the surface area of the particle, the 118 difference in density between the particle and the water, and inversely proportional to the dynamic 119 viscosity of the water (Dietrich 1982).

120 Previous studies mainly focused on either of these effects of vegetation in natural rivers or in laboratory experiments: hydraulic functioning (Bal et al. 2011; Green 2005b), solute transport (Nepf et al. 1997; 121 122 Sukhodolova et al. 2006), and particle transport (Defina and Peruzzo 2010; Horvath 2004). The majority 123 of field studies quantifying both aspects are either executed in different study sites, (e.g. Hensley and 124 Cohen 2012; Riis and Sand-Jensen 2006; Sand-Jensen et al. 1999; Sand-Jensen and Mebus 1996), or are 125 executed in one site, but at multiple moments in time with a varying discharge and stream velocity (e.g. 126 Sukhodolova et al. 2006; Wilcock et al. 1999). Since these season and site specific characteristics (such as 127 channel dimensions, bed forms, discharge etc.) also influence the transport processes (Gonzalez-Pinzon et 128 al. 2013), it is important to perform experiments in the same study reach wherein vegetation cover is 129 experimentally alerted in order to quantify the specific effects of these changes in vegetation cover.

130 The aim of this paper is to quantify the opposing effects of instream aquatic vegetation cover on the 131 drainage and transport capacity of lowland rivers. We address following research questions and 132 hypotheses:

133 1. How do changes in macrophyte cover (through partial and complete experimental vegetation 134 removal) affect the hydraulic functioning of lowland rivers, more specifically by affecting the 135 hydraulic roughness, mean flow velocity and water level? We hypothesize that vegetation cover is 136 positively correlated with the hydraulic resistance and negatively correlated with mean flow 137 velocity.

138 2. How do changes in macrophyte cover affect transport and retention of dissolved and particulate 139 matter? With decreasing vegetation cover, we hypothesize that decreased residence times of 140 dissolved organic matter (DOM) and changes in the magnitude of the dispersion coefficient and 141 transient storage zone. We also hypothesize that decreasing macrophyte cover increases the mean 142 travel distance and reduces the retention of coarse particulate organic matter (CPOM). What is the combined effect of changes in macrophyte cover on both the hydraulic functioning
and organic matter transport? We hypothesize that there are opposing effects, where macrophytes
negatively affect hydraulic functioning (through increased hydraulic roughness, decreased mean
flow velocity, and hence increasing water levels and flood risks), but positively affect water
quality (through decreased transport and increased retention of dissolved and particulate matter).

# 2. Materials and methods

### 149 **1. Study area**

150 The Brzozówka river is a lowland river (bottom slope of 0.0005 m m<sup>-1</sup>) in the North East of Poland (Fig. 151 1). A straight reach of 50 m long with a width ranging from 7 m to 10 m is selected to perform the 152 experiments (Fig. 2). This reach is outside the nature conservation area. The initial bathymetry was measured at 0.5 m intervals along cross-sectional transects which were located every 5 m along the whole 153 154 study reach. There is no lateral inflow in the study reach. The sediment on the river bed consists for 97.4 155 % of sand (diameter > 63  $\mu$ m) and for 2.6 % of silt (2-63  $\mu$ m), and the average organic matter content is 1.6 %. Concentration of the main solutes in the surface water are: 8.56 mg L<sup>-1</sup> Cl, 7.29 mg L<sup>-1</sup> Na, 0.034 156 mg L<sup>-1</sup> PO<sub>4</sub><sup>3-</sup>-P, 0.64 NO<sup>3-</sup>N mg L<sup>-1</sup>, 0.09 NH<sub>4</sub><sup>+</sup>-N. The treatments did not disturb biota nor the ecological 157 158 status of the river.



160 Figure 1: (a) Location of the study reach in the Brzozòwka river in the North East of Poland, indicated by 161 the black dot. (b) Study area in 2014 is shown with initial vegetation cover within the study reach 162 (between white lines) and a vegetation free section upstream of the study reach. Main flow direction is 163 indicated with an arrow.

### 2. Vegetation mapping

The initial species composition was recorded along cross-sectional transects with a cross-sectional resolution of 5 cm along each transect, and a longitudinal interval of 1 m between the transects (Fig. 2a and b). Seven submerged macrophyte species were found in the study reach: *Nuphar lutea* Sm., *Potamogeton crispus* L., *Potamogeton natans* L., *Potamogeton pectinatus* L., *Potamogeton perfoliatus* L., *Sparganium emersum* Rehmann, and. *Sagittaria sagittifolia* L. The emergent riparian vegetation at the river banks is all classified as riparian vegetation and not further identified.

171 The effect of vegetation cover on the hydraulic functioning and transport of dissolved and particular 172 organic matter was experimentally tested for three treatments with different vegetation covers. The first 173 treatment was with all natural initial vegetation present, referred to as *full* vegetation cover. For the 174 second treatment with a partial vegetation cover, a part of the vegetation was removed to create an 175 instream block pattern (Fig. 2c), referred to as partial vegetation cover. Finally all vegetation was 176 removed for the third treatment, which is the *no* vegetation cover. The aboveground biomass of each of 177 the seven species was determined by sampling quadrants of  $0.25 \text{ m}^2$  positioned in monotopic stands of 178 each species respectively, no replicates were taken. This was done in the study reach, when the pattern for 179 the second treatment was being cut. The vegetation samples were oven dried at 70°C for 48 h to obtain 180 the dry weight (DW). All treatments and measurements were done in June 2013 and repeated in June 181 2014, except the type of CPOM (see explanation below) varied between both years.



Figure 2: (a, b) Maps of the initial vegetation cover in the study area in 2013 and 2014 respectively. Colors are according to the dominance of the species, with *P. perfoliatus* being the most dominant species, followed by *P. pectinatus, S. sagittifolia, N. lutea, P. natans, P. crispus,* riparian vegetation and *S. emersum.* (c) Schematic overview of the location of the CDT divers ( $\bigstar$ ) and manual sampling points ( $\bigcirc$ ). The vegetation cover for three treatments is shown: exp. 1: all initial vegetation is present (called *full* vegetation cover); exp. 2: the vegetation is partly removed (*partial*); exp. 3: all vegetation is removed (*no*). The direction of the water flow is indicated with an arrow.

182

## 191 **3. Hydraulic measurements**

Each treatment started with the measurement of stream velocities with an electromagnetic flow meter (EMF, Valeport model 801, Totnes, UK) with a depth-interval of 20 cm, every meter along the crosssection at the upstream edge of the study reach. The velocity-area method was used to calculate the discharge (Bal and Meire 2009). The difference in water level is measured with a laser leveler (Spectra Precision, Coudere/geoservice, Brugge, BE). The water level slope is the difference of the upstream and
downstream water level divided by the reach length (50 m). The hydraulic resistance is expressed as a
Manning coefficient (Chow, 1959) and calculated as:

199 
$$n = \frac{A}{0} R^{2/3} S^{1/2}$$
 (Eq. 1)

with n (s m<sup>-1/3</sup>) the Manning coefficient, A (m<sup>2</sup>) the cross-sectional area, Q (m<sup>3</sup> s<sup>-1</sup>) the discharge, R (m) 200 201 the hydraulic radius which is the cross-sectional area divided by the wetted perimeter, S (m m<sup>-1</sup>) the water 202 level slope. After transport was measured with tracer experiments (see below) for the first treatment with 203 full vegetation cover, part of the vegetation was manually removed according to the scheme on Fig. 2c to 204 create the partial vegetation cover of the second treatment. The second treatment started the next day, 205 after a 20 h acclimatization period to minimize the external effects associated with the vegetation removal 206 like sediment suspension. This procedure was repeated after removal of all vegetation, i.e. the case of no 207 vegetation cover. As such, the whole procedure resulted in three similar sets of measurements for full partial and no vegetation cover in both June 2013 and June 2014. 208

209

**4. Solute transport** 

211 Two releases (with a time interval of 2h) of the dissolved tracer were performed, by adding 12 kg NaCl 212 that was dissolved in 6 buckets of 10 L river water. Only one release was done in 2013 with the partial 213 vegetation cover. Salt was used as a conservative tracer, so that no uptake or conversion to other 214 molecules is expected, within the timespan of the experiment (i.e. 20 min). One automated and two 215 manual sampling points were placed at both the upstream and downstream edge of the study reach (Fig. 216 2c). The conductivity was measured automatically with a frequency of 1 Hz (CTD divers, Eijkelkamp, 217 Geisbeek, NL). Additionally manual samples of 40 mL were taken at the upstream border, every 10 s during the first 2 min after the release, then every 5 s up to 6 min after the release and every 20 s up to 10 218

min after the release. Manual downstream sampling started after 3 min 15 s with an interval of 20 s, up to 15 min 15 s after the release, resulting in a total of 37 samples. Three additional samples were taken with an interval of 60 s, so the sampling ended after 18 min 15 s. The conductivity of the manually taken samples was measured with a multimeter (Multil 340/SET, Weilheim, GE). The conductivity is converted to NaCl concentrations with a linear calibration curve of six NaCl standards in the range of 2.2 to 610  $\mu$ S cm<sup>-1</sup>.

225 Break through curves (BTS), which show the concentration in function of time after the release, are 226 zeroed to background concentrations of the river water (Gonzalez-Pinzon et al. 2013). The fractional mass 227 recovery is the mass recovered (the area under the curve multiplied with the discharge) divided by the 228 initial mass (Gonzalez-Pinzon et al. 2013). The start time ( $T_{start}$ ) and end time ( $T_{end}$ ) are defined as the 229 start and end of the BTC, respectively. The difference between T<sub>end</sub> and T<sub>start</sub> is the duration of the signal. 230 The median residence time  $(T_{med})$  is the time needed for the passage of 50% of the salt through the study 231 reach. The peak time  $(T_{peak})$  is the time of the peak concentration  $(C_{peak})$  at the downstream edge of the 232 study reach. The incoming velocity is used as a reference velocity ( $U_{ref}$ ) and is calculated as the discharge 233 divided by the cross-sectional area. The reference time  $(T_{ref})$  is the average time that the bulk water flow 234 would need to pass the study reach without external influences and is defined as the reach length divided by U<sub>ref</sub>. All times are divided by the reference time (T<sub>ref</sub>) to correct for small differences in incoming 235 236 velocities between the treatments with different vegetation covers and in 2013 and 2014. The median 237 velocity (U<sub>med</sub>) and mean velocity (U<sub>mean</sub>) correspond to the reach length divided by T<sub>med</sub> and T<sub>peak</sub>, respectively. Similarly, these velocities, U<sub>med</sub> and U<sub>mean</sub>, are corrected for differences in incoming 238 239 velocities by dividing them by U<sub>ref</sub>.

The temporal moments of the time series are calculated to investigate solute transport (Das et al. 2002;
Govindaraju and Das 2002). The n<sup>th</sup> absolute moment is defined as:

242 
$$\mu_n = \int_0^\infty t^n C(t) dt$$
 (Eq. 2)

~ ~

244 
$$\mu_n^* = \frac{\mu_n}{\mu_0}$$
 (Eq. 3)

The normalized absolute moments are used to calculate the normalized central moments of order 1 ( $m_1$ , mean travel time), order 2 ( $m_2$ , variance on the travel time which can be due to higher dispersion) and order 3 ( $m_3$ , skewness on the travel time which can be due to larger transient storage) (Gonzalez-Pinzon et al. 2013) as:

249  $m_1 = \mu_1^*$ 

250 
$$m_2 = \mu_2^* - {\mu_1^*}^2$$
 (Eq. 4)

251 
$$m_3 = \mu_3^* - 3\mu_1^* \mu_2^* + 2 \mu_1^{*3}$$

252

253 The transient storage model (Gonzalez-Pinzon et al. 2013) is given by:

254 
$$\frac{dC}{dt} = -U \frac{dC}{dx} + D \frac{d^2C}{dx^2} - \beta \alpha (C - C_s)$$
(Eq. 5)

$$255 \qquad \frac{\mathrm{dC_s}}{\mathrm{dt}} = \alpha \left( \mathrm{C} - \mathrm{C_s} \right)$$

with C (g m<sup>-3</sup>) the concentration of the solute in the main channel, C<sub>s</sub> (g m<sup>-3</sup>) the concentration of the solute in the storage zone, U (m s<sup>-1</sup>) the flow velocity, D (m<sup>2</sup> s<sup>-1</sup>) the dispersion coefficient,  $\beta$  (-) the spatially average transient storage zone volume fraction  $\beta = A_s/A$ ,  $A_s$  (m<sup>2</sup>) the cross-sectional area of the transient storage zone, A (m<sup>2</sup>) the cross-sectional area of the main channel,  $\alpha$  (s<sup>-1</sup>) the mass-exchange rate coefficient between the main channel and the transient storage zone, t (s) the time, x (m) distance (Gonzalez-Pinzon et al. 2013).

The theoretical moments can be used to calculate the parameters of the transient storage model and are defined by (Czernuszenko and Rowinski 1997) for a general boundary condition:

265 
$$m_{1,fit} = \frac{2D}{U^2} + \frac{L}{U}(1+\beta)$$

266 
$$m_{2,fit} = \frac{8D^2}{U^4} + \frac{L}{U}\frac{2D}{U^2}(1+\beta) + \frac{2L}{U}\frac{\beta^2}{\alpha}$$
 (Eq. 6)

267 
$$m_{3,\text{fit}} = \frac{2L^2}{U^2} \frac{D}{U^2} (1+\beta)^2 \beta + \frac{64D^3}{U^6} + \frac{L}{U} \left[ \frac{12D^2}{U^4} (1+\beta)^2 + \frac{4D}{U^2} \frac{\beta^2}{\alpha} (\beta+2) + \frac{6\beta^3}{\alpha^2} \right]$$

with the same variables as explained above and L (m) the reach length, 50 m. We have three unknown variables: dispersion coefficient (D), mass-exchange rate ( $\alpha$ ) and spatially average transient storage zone volume fraction ( $\beta$ ), and three equations (Eq. 6). All BTC of each vegetation cover are used to determine the variables D,  $\alpha$ ,  $\beta$ . The flow velocity (U) is the reach length divided by moment 1 and is averaged for all experiments per vegetation cover. A pseudo randomized algorithm followed by a Levenberg-Marquardt algorithm is used to minimize the cost function (CF).

274 
$$CF = \sum_{i=1}^{3} \sum_{k=1}^{K} \left[ (m_{i,fit} - m_{i,k}) / st dv(m_{i,k}) \right]^2$$
 (Eq. 7)

275 with K the number of BTC per vegetation cover.

R (v. 3.2.0 R Core Team) was used to perform the statistical analyses. A two-way ANOVA followed by a
post-hoc Tukey test were performed to compare the parameters according to the three vegetation covers.

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279

#### 5. Particle transport and retention

The transport and retention were investigated with three coarse particulate organic matter (CPOM) types: oats, wood chips and cornflakes. The tracers were characterized in the laboratory (Tab.1). Firstly, the surface area of pictures of 70 - 150 particles was calculated using ArcGIS software (v. 10.1, ESRI Inc, 283 Redlands, USA). Secondly, the buoyancy in standing and gently stirred water was measured by placing a 284 fixed amount of each tracer in a circular aquarium (80 g oats, 20 g wood chips and 20 g cornflakes). The 285 aquarium had a water depth of 0.15 m and three replicate tests per tracer were carried out under standard 286 laboratorial conditions (20 °C). After 20 min all floating and all sunken particles were collected, dried 287 (72h, 70 °C) and weighted. The buoyancy is expressed as the ratio of the weight of floating particles to 288 the weight of all particles (Tab. 1). Thirdly, as part of the tracers' mass can get lost in the river due to 289 small detaching particle components that could not be recovered, a conversion factor is calculated by the 290 ratio of the initial DW and final DW after being in the water for 20 min. The conversion factor is used to 291 convert the final weight of tracers in the field experiments.

292 Table 1: Characteristics of the three particulate tracers: oats, wood chips and cornflakes. The mean and standard 293 deviation of the buoyancy in standing and stirred water is given. The conversion factor accounts for the detaching of 294 particle components.

		Oats	Wood chips	Cornflakes
Surface area	$\text{mm}^2 \pm \text{sd}$	$19.9\pm9.5$	$14.4\pm9.5$	$179.7\pm78.0$
Buoyancy standing	$\% \pm sd$	$0.60\pm0.2$	$57.8 \pm 1.1$	$98.4 \pm 1.4$
Buoyancy stirred	$\% \pm sd$	$0.62\pm0.7$	$34.4\pm7.3$	$45.7\pm1.5$
Conversion factor	$mean \pm sd$	$1.53\pm0.07$	$1.08\pm0.01$	$1.33\pm0.02$

295

The field experiments with oats were conducted in 2013 with a release of 3.00 kg oats at each vegetation cover. The same protocol was repeated in 2014 with 2.50 kg cornflakes and 3.00 kg wood chips. The floating part of tracers was captured downstream of the study reach with small nets in the top 0.15 m of the water column. The collected tracers were oven dried (72h, 70 °C), weighted and multiplied with the conversion factor. The retention is the mass of the tracer that remained in the study reach relative to the initial mass. The amount of particles in transport along the reach can be approached by an exponential decrease (Lamberti,1996) and is calculated using the following equation:

303 
$$P_d = P_0 e^{-kd}$$
 (Eq. 8)

with  $P_d$  (g) the amount of particles in transport downstream of the reach,  $P_0$  (g) the initial amount of particles, k (m<sup>-1</sup>) the instantaneous retention rate, d (m) the length of the study reach. The mean travel distance is the inverse of the mean instantaneous retention rate. Finally, the depositional velocity is calculated to enable the comparison of our results to reaches with different velocities and sizes. The depositional velocity expresses the rate at which tracers leave the water column (Warren et al. 2009):

$$309 V_{dep} = k u h (Eq. 9)$$

310 with  $V_{dep}$  (m s<sup>-1</sup>) the depositional velocity, k (m<sup>-1</sup>) the instantaneous retention rate, u (m s<sup>-1</sup>) the mean 311 velocity and h (m) the water depth.

**312 3. Results** 

#### 313 **1. Vegetation**

The vegetation maps are shown in Fig. 2. The species specific vegetation cover and biomass at the initial situation is given in Tab. 2. A higher initial vegetation cover is present in 2013, 89.6 %, compared to 2014, 76.6 %. The mean dry weight in the whole reach is 297.9 gDW m<sup>-2</sup> in 2013, which is also higher than 232.9 gDW m<sup>-2</sup> in 2014 (Tab. 2). In the vertical dimension (not shown in Fig. 2), the submerged vegetation fills the whole water column and the majority of the vegetation reaches the water surface.

**Table 2:** Species specific vegetation cover and biomass at the full vegetation cover. Data are collected in June 2013
 and June 2014 over a reach of 50 m.

	Ju	ne 2013	June 2014		
	Cover	Biomass	Cover	Biomass	
	(%)	$(gDW m^{-2})$	(%)	(gDW m <sup>-2</sup> )	
N. lutea	0.35	174.4	0.30	186.8	
P. crispus	0.76	NA	4.81	264.8	
P. natans	5.50	298.8	5.41	94.0	
P. pectinatus	23.71	224.8	7.74	278.4	
P. perfoliatus	36.36	383.2	32.66	455.6	
S. emersum	0.52	NA	1.28	126.8	

S. sagittifolia	20.81	376.8	15.66	146.4	
Riparian vegetation	1.41	NA	8.82	NA	
No vegetation	10.40	0	23.32	0	
Total vegetation cover	89.6	279.9	76.7	232.9	

## 322 **2.** Hydraulics

The hydraulic parameters are summarized in Tab. 3. The discharge ranges between 0.76 and 1.09 m<sup>3</sup> s<sup>-1</sup> 323 between the treatments (different days). During the experiment of one treatment in one year the discharge 324 325 is assumed to be constant. The magnitude of the reference velocity (U<sub>ref</sub>) and reference time (T<sub>ref</sub>) reflect 326 the differences in the prevailing discharge between the treatments. In both years a positive relationship is 327 observed between the Manning coefficient and vegetation cover (Tab. 3). The highest Manning 328 coefficient of 0.30 s m<sup>-1/3</sup> coincides with the highest vegetation cover of 89.6 %. The water level slope has 329 a similar trend with being 5 to 15 times higher in the vegetated reach compared to the vegetation free 330 reach (Tab. 3).

331 Table 3: Hydraulic parameters of the study reach in 2013 and 2014 at three vegetation covers (full, partial and no vegetation cover).

		June 2013		June 2014		ļ	
Vegetation cover		Full	Partial	No	Full	Partial	No
Macrophyte cover	%	89.6	56.3	0	76.7	47.9	0
Discharge upstream	m <sup>3</sup> s <sup>-1</sup>	0.82	0.90	0.78	1.09	0.76	0.82
Cross-sectional area upstream	$m^2$	7.45	8.33	7.24	9.72	8.23	8.76
U <sub>ref</sub>	$m s^{-1}$	0.110	0.108	0.108	0.112	0.092	0.093
$T_{ref}$	S	454	463	464	446	541	534
Hydraulic radius	m	0.78	0.83	0.76	0.94	0.81	0.86
Water depth	m	0.87	0.90	0.82	0.97	0.86	0.86
Mean width	m	8.1	8.1	8.1	8.9	8.9	8.9
Manning coefficient	s m <sup>-1/3</sup>	0.30	0.24	0.08	0.28	0.27	0.14
Water level slope	m m <sup>-1</sup>	0.0015	0.0009	0.0001	0.0011	0.0009	0.0002

## 3. Solute transport

An example of the breakthrough curves (BTC) for the three vegetation covers treatments is given in Fig. 3. The tracer is uniformly injected along the whole upstream width of the reach, which results in an equal distribution along the transverse direction. This is confirmed by the three sampling points downstream for the full and no vegetation cover treatment. For the partial vegetation cover a clear difference is seen between the three sampling points at the downstream end of the reach.



340

Figure 3: Breakthrough curves of NaCl concentration at the downstream edge of the reach are shown for (a) full, (b) partial and (c) no vegetation cover, results of 2014 second release. The concentrations of the center point are measured with a CTD diver (black line). Samples near the left bank (grey dots) and right bank (black dots) are taken manually.

346 The transport parameters of solutes are given in Tab. 4. The fractional mass recovery ranges between 85 347 and 106 % for all experiments. Expect for the second replica of full vegetation treatment in 2014, the 348 fractional mass recovery was 127 %. This is probably due to an overestimation of the discharge. The 349 experiments with a full vegetation cover have an average recovery of 107.2 %. Significant differences 350 between full and no vegetation cover are found for the relative start time, the relative time of the peak 351 concentration, and the relative stream velocity based on the peak concentration. This implies that 352 vegetation significantly reduces the flow velocity and increases the travel time, regardless the upstream 353 flow velocity. The differences are tested between the left and right bank in the partial vegetated reach. A 354 significant higher flow velocity ( $U_{peak}/U_{ref}$ ) 1.55 and 1.25 for the right bank and left bank, respectively 355 (p=0.03). And a significant lower time for the peak concentration ( $T_{peak}/T_{ref}$ ) are found for the right bank, 356 0.65, compared to the left bank, 0.80 (p=0.04). While the difference for U<sub>peak</sub>/U<sub>ref</sub> and T<sub>peak</sub>/T<sub>ref</sub> were not 357 significantly different between right bank and left bank for the full and empty vegetation cover.

358 Next, the second and third normalized central moments significantly vary between the full and no 359 vegetation cover (Tab. 4). For the same travel time a higher variance (second moment) and a higher 360 skewness (third moment) is found without vegetation compared to a full vegetation cover (Fig. 4). A 361 higher variance can be due to higher dispersion and a higher skewness can be related to transient storage. 362 This is confirmed by the parameters of the transient storage model. The dispersion coefficient (D) and the 363 spatially average transient storage zone volume fraction ( $\beta$ ) increase without vegetation. The goodness of 364 fit was estimated with the Nash-Sutcliffe model efficiency coefficient (E) (Nash 1959). This coefficient 365 ranges between 0.71-0.96 for full vegetation cover, 0.51-0.97 for the partial vegetation cover and 0.82-366 0.97 for the no vegetation cover. A separate parameter fit was done for the left and right bank of the partial vegetation cover, due to the significant difference of the peak flow velocity. Similar effects of the 367 presence of vegetation are found in this scenario. The dispersion coefficient is lower,  $0.134 \text{ m}^2 \text{ s}^{-1}$ , for the 368 369 left bank, which is downstream of a vegetated part (Fig. 2). While a value of 0.203 m<sup>2</sup> s<sup>-1</sup> was found for

the right bank downstream of the artificial channel (Fig. 2). The mass exchange rate ( $\alpha$ ) and the volume of the transient storage zone ( $\beta$ ) are in both cases 10<sup>-7</sup> s<sup>-1</sup> and 10<sup>-7</sup> %, respectively. When parameter settings were applied separately for both river banks, this improved the model fit between 0.68-0.88 and 0.88-0.95 for respectively the left and right bank.

Table 4: Transport of NaCl as an analogue for the behaviour of solute transport in streams. Full: n=11, partial: n=9,
 no: n=12. Letters indicate significant differences (p<0.05) based on a two-way ANOVA followed by a post-hoc</li>
 Tukey test.

		Vegetation cover				
		Full	Partial	No		
General parame	ters					
Recovery	%	$107.2\pm6.4$ $^{a}$	$89.1\pm3.5$ $^{\rm b}$	$89.7\pm2.5$ $^{\rm b}$		
$T_{start} / T_{ref}$	-	$0.47\pm0.02^{\rm a}$	$0.41\pm0.02$ $^{\rm b}$	$0.39\pm0.02^{b}$		
$T_{peak}$ / $T_{ref}$	-	$0.83\pm0.03^{\text{ a}}$	$0.76\pm0.03^{ab}$	$0.67\pm0.03^{\text{ b}}$		
$T_{med}$ / $T_{ref}$	-	$0.93\pm0.03$	$0.86\pm0.05$	$0.82\pm0.03$		
$T_{end} / T_{ref}$	-	$1.83\pm0.07$	$1.73\pm0.11$	$1.91\pm0.09$		
$U_{\text{peak}}/U_{\text{ref}}$	-	$1.22\pm0.05^{\text{ a}}$	$1.33\pm0.06^{ab}$	$1.52\pm0.07$ $^{\text{b}}$		
$U_{med}\!/U_{ref}$	-	$1.09\pm0.03$	$1.18\pm0.06$	$1.25\pm0.05$		
Duration/T <sub>ref</sub>	-	$1.36\pm0.07$	$1.31\pm0.10$	$1.52\pm0.09$		
$C_{peak}$	g m <sup>-3</sup>	$49.5\pm3.2$	$44.7\pm1.8$	$45.0\pm1.6$		
Normalized cent	ral mome	ents				
$m_1$	S	$445\pm12$	$469 \pm 19$	$446\pm10$		
$m_2 \ x 10^3$	$s^2$	$15.7 \pm 1.9^{\mathrm{a}}$	$18.5\pm2.3^{ab}$	$26.0\pm2.8^{\text{b}}$		
$m_3 \ge 10^5$	s <sup>3</sup>	$13.7\pm3.3^{\rm a}$	$17.0\pm5.4^{ab}$	$40.0\pm8.9^{b}$		
Transient storag	e model					
U (input)	m s <sup>-1</sup>	0.113	0.108	0.113		
D	$m^2 s^{-1}$	0.187	0.193	0.262		
α x 10 <sup>-7</sup>	s <sup>-1</sup>	1	1	1		
β x 10 <sup>-4</sup>	%	0.001	0.001	1.67		
Е	-	0.71-0.95	0.51-0.97	0.82-0.97		



**Figure 4: (a)** The variance (second moment) and (**b**) skewness (third moment) in function of mean travel time (first moment) for three vegetation covers: full (diamond), partial (square) and no (triangle) vegetation cover. For the partial vegetation cover different colors are used according to the location of the sampling point (Fig. 2c), because each location has different properties according to the presence of vegetation see Fig. 2c: the sampling point at left bank is behind the vegetation (drak grey, 'partial L'), the central sampling point is in the middle of the channel (grey, 'partial C'), the sampling point at the right bank is in the open channel (light grey, 'partial R').

#### 4. Particle transport and retention

387 The retention percentage and retention rate of the particular tracers is positively correlated with the vegetation cover for all substances (Fig. 5). For oats, all particles remain trapped in the study reach for the 388 389 experiments with full and partial vegetation cover. Therefore it was not possible to calculate the retention 390 rate, mean travel distance and depositional velocity (Tab. 5). The retention percentage of woodchips 391 varied between 86.01 to 97.17 % depending on the vegetation cover. Cornflakes have the lowest retention 392 percentage on the reach scale for all vegetation covers. The transport distance of the tracers is negatively 393 correlated to the vegetation cover: the particles are further transported when low vegetation covers are 394 present. A negative correlation between the depositional velocity and vegetation cover is observed for all 395 tracers.

Cornflakes Wood chips Oats



**Table 5:** Retention and transport of CPOM types, oats, wood chips and cornflakes.

			Vegetation cove	er
		Full	Partial	No
Oats				
Retention percentage	%	100.0	100.0	99.58
Retention rate	m <sup>-1</sup>	/	/	0.11
Mean travel distance	m	/	/	9.1
Depositional velocity	m s <sup>-1</sup>	/	/	0.0127
Wood chips				
Retention percentage	%	97.17	96.10	86.01
Retention rate	$m^{-1}$	0.071	0.065	0.039
Mean travel distance	m	14.0	15.4	25.4
Depositional velocity	m s <sup>-1</sup>	8.62	6.15	0.0037
Cornflakes				
Retention percentage	%	92.24	81.54	55.77

Retention rate	m <sup>-1</sup>	0.051	0.034	0.016
Mean travel distance	m	19.6	29.6	61.3
Depositional velocity	m s <sup>-1</sup>	6.18	3.20	0.0016

# 408 **4. Discussion**

409 Macrophytes are generally known to have multiple effects on the functioning of river ecosystems (Gurnell 410 2014; O'Hare 2015). The hydraulic functioning is affected by reducing the conveyance capacity, while the 411 transport capacity of dissolved and particulate matter is affected as well. The aim of this study is to 412 investigate these opposing effects by tracer experiments at three vegetation covers after experimentally 413 removing the vegetation in the same river reach. We found that (i) partially vegetated reaches had on the 414 one hand reduced hydraulic resistance compared to the fully vegetated treatment due to flow 415 concentration in to non-vegetated zones and on the other hand enlarged retention of particulate matter 416 compared to the vegetation free treatment; (ii) vegetated reaches had the highest hydraulic resistance and 417 the highest retention percentage for particulate matter; (iii) vegetation free reaches had the highest flow 418 velocity, the highest longitudinal dispersion coefficient and the largest transient storage zone.

419 The highest reduction in hydraulic roughness is observed in partially vegetated reaches (Tab.3), while still 420 maintaining the transport and retention capacity of the river (Tab. 4). The hydraulic resistance of fully 421 vegetated reaches was two to three times higher than after complete vegetation removal, while the 422 hydraulic resistance of the partially vegetated treatment ranged between them (Tab. 3). Similar results are 423 found by an extensive study that compared 35 vegetated lowland rivers with different vegetation covers (Green 2005a). Manning coefficients ranged between 0.12 and 0.39 s m<sup>-1/3</sup> for reaches with vegetation 424 425 covers of above 60 %, between 0.07 and 0.15 for covers between 35 and 60 %, whereas these values reduced to 0.09 s m<sup>-1/3</sup> for covers below 35 % for the same submerged macrophyte species and similar 426 427 flow velocities. The relatively low hydraulic resistance in the partially vegetated treatment is caused by 428 the presence of preferential flow paths (Bal et al. 2011) through deviation and concentration of flow to

429 non-vegetated zones. The occurrence of this process in our study is confirmed by the significant 430 difference of the flow velocity between the right bank and left bank in the partial vegetation cover. 431 Similar flow deviation and concentration mechanisms to non-vegetated zones of river reaches with partial 432 patchy vegetation has been show e.g. by Sand-Jensen and Mebus (1996) and Schoelynck et al. 433 (2013), Verschoren et al. (2016).

434 Increased hydraulic resistance can lead to increased water levels (Madsen et al. 2001). In our study 435 vegetation cover was experimentally varied over a limited reach of 50 m so that effects on water level are 436 small, and therefore we consider the water level slope as the most relevant proxy for the effect of the 437 varying vegetation covers on the drainage capacity. A five- to fifteen fold reduction in the water level 438 slope was observed after the removal of all vegetation (Tab. 3). Other studies where instream vegetation 439 was removed over a larger distance (5 km) showed a decrease of the water level between 17 and 28 % 440 (Old et al. 2014) and 25 to 67% (Bal and Meire 2009). Simultaneously, the Manning coefficient dropped 441 between 43 and 54 % (Old et al. 2014) and 25 to 67 % (Bal and Meire 2009) in the aforementioned 442 studies and between 50 and 73 in our study. Along with increased water levels, macrophytes reduces the 443 flow velocity relative to the incoming flow velocity (Tab. 4), this is a well-known phenomenon in 444 vegetated rivers (De Doncker et al. 2009a; Franklin et al. 2008; Sand-Jensen and Pedersen 2008). Lower 445 flow velocities reduce the advection of solutes and increase the residence time of solutes and water. 446 Residence time is one of the major factors controlling the nutrient removal in rivers (Seitzinger et al. 447 2006). Consequently, the beneficial effects of solutes for the self-purification capacity of the river are reduced when macrophytes are removed (Runkel 2007). 448

The flow concentration to non-vegetated zones, associated with strong flow reduction within the vegetated zones, can be beneficial for the trapping and deposition of fine and coarse material in the vegetation patches, while higher stream velocities in the channels are the preferential flow path of particles and there erosion of mineral sediment can be induced (Madsen et al. 2001; Old et al. 2014). Also 453 organic matter is accumulated in macrophytes patches, forming biogeochemical hotspots with efficient 454 remineralization (Schoelynck 2011). Increasing macrophyte covers enhanced the retention of all three 455 tested particle types (Fig. 5 and Tab. 5). The retention rate in vegetated reaches compared to vegetation 456 free reaches was two- and threefold higher for wood chips and cornflakes, respectively. Similar 457 observations with particles of other sizes were found for the influence of instream vegetation cover. The 458 retention rate was for (i) macrophyte stem fragments (15-20 cm) 0.02-0.12 m<sup>-1</sup> and 0.0005-0.0135 m<sup>-1</sup> 459 (Riis and Sand-Jensen 2006); (ii) circular paper chips (diameter 6 mm) 0.28-1.2 m<sup>-1</sup> and 0.02-0.26 m<sup>-1</sup> (Horvath 2004); (iii) corn pollen (diameter 85-90 µm) 0.017 m<sup>-1</sup> and 0.011 m<sup>-1</sup> (Warren et al. 2009) in 460 vegetated and vegetation free streams respectively. In addition the retention rate was inversely related to 461 462 the buoyancy of the particles, with the lowest retention rate (Tab. 5) for cornflakes which has the highest 463 buoyancy (Tab. 1). Similar effects of buoyancy were observed with lower retention rates for particles 464 with a higher buoyancy (Defina and Peruzzo 2010) for the same vegetation cover. Oats had the lowest 465 buoyancy (Tab. 1) and no retention rate could be calculated for the fully and partial vegetated treatment 466 because it cannot be known after which distance all particles were retained (Tab. 5). The effect of vegetation was overruled by the limited buoyancy of this tracer. Beside instream aquatic vegetation, other 467 468 roughness structures can have similar effects on particle trapping, like riparian vegetation (Riis and Sand-469 Jensen 2006), logs, branches and tree trunks (Cordova et al. 2008; Schneider and Sharitz 1988) and 470 substrate heterogeneity (Muotka and Laasonen 2002). This may explain the relative high retention 471 percentage of wood chips in the vegetation free reach (Tab. 5). However, in large rivers the relationship 472 between the retention percentage of particles and riparian vegetation cover was negative (Nilsson et al. 473 1991) or not significant (Andersson et al. 2000). This might be attributed to the relative smaller impact of 474 riparian vegetation in larger rivers. In these river, the majority of particles (in these studies seeds) is 475 transported in the main channel and is not obstructed by bank irregularities or riparian vegetation.

476 In addition, complex spatial flow when vegetation is present leads to distinct effects on transport and 477 mixing processes of solutes. Flow channels between the vegetation lead to a clear peak in the 478 concentration-time curves. Increased vegetation cover increased the magnitude of the peak concentration 479 of the dissolved tracer (Tab. 4) and reduced the magnitude of transient storage zones (Tab. 4). The lowest 480 dispersion coefficients are found for vegetated reaches (Tab. 4). This can be attributed to the ability of 481 dense, uniform vegetation to reduce vertical gradients in the flow velocity (Baptist et al. 2009; Nepf et al. 482 1997). However a higher mechanical dispersion is expected due to the dense network of vegetation. The 483 increase of mechanical dispersion due to the presence of vegetation can be counteracted by the decrease 484 in stream velocity which lowers the shear stress separation and total dispersion (Folkard 2011; Lightbody 485 and Nepf 2006). Similarly, the presence of vegetation mimics resulted in a decreased longitudinal 486 dispersion coefficient in a flume experiment (Nepf et al. 1997). This was explained by higher turbulence 487 and diminished vertical shear stress in the presence of vegetation. In addition, plant morphology itself can 488 also affect the longitudinal dispersion (Lightbody and Nepf 2006) and sediment retention (Rovira et al. 489 2016). An equal distribution of frontal area of the canopy along the vertical results in a fairly constant 490 velocity profile and generates relatively low vertical shear dispersion. While complex plant morphologies 491 generate pronounced variation of the vertical velocity resulting in more shear flow dispersion (Lightbody 492 and Nepf 2006). The vegetation consists in our study area of multiple species (Fig. 2), but we did not 493 distinguish the effect of individual species on the longitudinal dispersion. From the above argumentation, 494 it is clear that multiple interactions are present in these natural rivers. This leads to large variability of the 495 effect of macrophytes on the longitudinal dispersion coefficient as well as the transient storage zone. A 496 study conducted in vegetated spring-fed karst rivers found also a lower longitudinal dispersion coefficient 497 but higher transient storage zone at increased vegetation cover (Hensley and Cohen 2012). This can be 498 explained by the difference in geomorphology of the river channel dominated by karst. In contrast to our 499 results, a field study conducted with three different levels of vegetation cover at three different moments 500 in the year found a positive relationship between biomass and the longitudinal dispersion coefficient

501 (Sukhodolova et al. 2006) and the magnitude of the transient storage zone increased with vegetation cover 502 in lowland rivers. However it was difficult to isolate effects of macrophytes on the dispersion coefficient 503 and transient storage zone in this study, since changes in these parameters are mainly attributed to large 504 variation in discharge between the sampling days and the presence of recirculation zones resulting from 505 bank irregularities (Sukhodolova et al. 2006).

# 506 **5. Conclusion**

507 Our results show complex interactions between the presence of macrophytes and spatial flow patterns, 508 and hence distinct effects on transport and mixing processes are found of dissolved and particulate tracers. 509 After vegetation removal the hydraulic resistance expressed as a Manning coefficient is two to three times 510 lower and water level slope drops by two to five times. The flow velocity increases when vegetation is 511 removed, and the most heterogeneous flow field was found in the partially vegetated treatments, due to 512 flow concentration to non-vegetated zones. The effect of vegetation on the transport of solutes and 513 particles is twofold, vegetation decreases the longitudinal dispersion coefficient and decreases transient 514 storage of solutes, while the retention rate of particulate tracers is two- and threefold higher. Clear 515 differences are found between the three particulate tracers depending on their buoyancy, with the lowest 516 retention rate for cornflakes which has the highest buoyancy. The partially removing of the vegetation leads to an optimal trade-off between maximizing the residence time and retention of organic matter 517 518 while minimizing the hydraulic resistance compared to the fully vegetated and vegetation free treatment.

519

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### 527 **References**

- Andersson, E., C. Nilsson & M. E. Johansson, 2000. Plant dispersal in boreal rivers and its relation to the
   diversity of riparian flora. J Biogeogr 27(5):1095-1106 doi:DOI 10.1046/j.1365 2699.2000.00481.x.
- Bal, K. D. & P. Meire, 2009. The influence of macrophyte cutting on the hydraulic resistance of lowland
   rivers. J Aquat Plant Manage 47:65-68.
- Bal, K. D., E. Struyf, H. Vereecken, P. Viaene, L. De Doncker, E. de Deckere, F. Mostaert & P. Meire,
  2011. How do macrophyte distribution patterns affect hydraulic resistances? Ecol Eng 37(3):529533 doi:DOI 10.1016/j.ecoleng.2010.12.018.
- Baptist, M. J., V. Babovic, J. R. Uthurburu, M. Keijzer, R. E. Uittenbogaard, A. Mynett & A. Verwey,
   2009. On inducing equations for vegetation resistance Discussion. J Hydraul Res 47(2):281-281.
- Bencala, K. E. & R. A. Walters, 1983. Simulation of solute transport in a mountain pool-and-riffle stream
   a transient storage model. Water Resour Res 19(3):718-724 doi:Doi
  10.1029/Wr019i003p00718.
- Boedeltje, G., J. P. Bakker, A. Ten Brinke, J. M. Van Groenendael & M. Soesbergen, 2004. Dispersal
  phenology of hydrochorous plants in relation to discharge, seed release time and buoyancy of
  seeds: the flood pulse concept supported. J Ecol 92(5):786-796 doi:DOI 10.1111/j.00220477.2004.00906.x.
- Boerema, A., J. Schoelynck, K. Bal, D. Vrebos, S. Jacobs, J. Staes & P. Meire, 2014. Economic valuation
  of ecosystem services, a case study for aquatic vegetation removal in the Nete catchment
  (Belgium). Ecosyst Serv 7:46-56 doi:10.1016/j.ecoser.2013.08.001.
- 548 Chow, V.T., 1959. Open-channel hydraulics. McGraw-Hill, New York, USA.
- Church, M., 2006. Bed material transport and the morphology of alluvial river channels. Annu Rev Earth
   Pl Sc 34:325-354 doi:10.1146/annurev.earth.33.092203.122721.
- Cordova, J. M., E. J. Rosi-Marshall, J. L. Tank & G. A. Lamberti, 2008. Coarse particulate organic matter
   transport in low-gradient streams of the Upper Peninsula of Michigan. J N Am Benthol Soc
   27(3):760-771 doi:10.1899/06-119.1.
- Cotton, J. A., G. Wharton, J. A. B. Bass, C. M. Heppell & R. S. Wotton, 2006. The effects of seasonal changes to in-stream vegetation cover on patterns of flow and accumulation of sediment. Geomorphology 77(3-4):320-334 doi:10.1016/j.geomorph.2006.01.010.
- 557 Czernuszenko, W. & P. M. Rowinski, 1997. Properties of the dead-zone model of longitudinal dispersion
   558 in rivers. J Hydraul Res 35(4):491-504.
- Danvind, M. & C. Nilsson, 1997. Seed floating ability and distribution of alpine plants along a northern
   Swedish river. J Veg Sci 8(2):271-276 doi:Doi 10.2307/3237356.
- Das, B. S., R. S. Govindaraju, G. J. Kluitenberg, A. J. Valocchi & J. M. Wraith, 2002. Theory and
   applications of time moment analysis to study the fate of reactive solutes in soil. American
   Society of Civil Engineers.
- De Doncker, L., P. Troch, R. Verhoeven, K. Bal, N. Desmet & P. Meire, 2009a. Relation between
   resistance characteristics due to aquatic weed growth and the hydraulic capacity of the river Aa.
   River Res Appl 25(10):1287-1303 doi:Doi 10.1002/Rra.1240.
- De Doncker, L., P. Troch, R. Verhoeven, K. Bal, P. Meire & J. Quintelier, 2009b. Determination of the
   Manning roughness coefficient influenced by vegetation in the river Aa and Biebrza river.
   Environ Fluid Mech 9(5):549-567 doi:DOI 10.1007/s10652-009-9149-0.
- 570 Defina, A. & P. Peruzzo, 2010. Floating particle trapping and diffusion in vegetated open channel flow.
   571 Water Resour Res 46 doi:Artn W1152510.1029/2010wr009353.

 <sup>572</sup> Dietrich, W. E., 1982. Settling velocity of natural particles. Water Resour Res 18(6):1615-1626 doi:Doi
 573 10.1029/Wr018i006p01615.

- 574 Ehrman, T. P. & G. A. Lamberti, 1992. Hydraulic and Particulate Matter Retention in a 3rd-Order Indiana
   575 Stream. J N Am Benthol Soc 11(4):341-349 doi:Doi 10.2307/1467556.
- 576 Folkard, A. M., 2011. Vegetated flows in their environmental context: A review. Proc. ICE -577 Eng.Comput.Mech.164(1):3-24 doi:10.1680/eacm.8.00006.
- 578 Franklin, P., M. Dunbar & P. Whitehead, 2008. Flow controls on lowland river macrophytes: A review.
  579 Sci Total Environ 400(1-3):369-378 doi:DOI 10.1016/j.scitotenv.2008.06.018.
- Gonzalez-Pinzon, R., R. Haggerty & M. Dentz, 2013. Scaling and predicting solute transport processes in
   streams. Water Resour Res 49(7):4071-4088 doi:10.1002/wrcr.20280.
- 582 Govindaraju, R. S. & B. S. Das, 2002. Moment analysis for subsurface hydrological applications.
- Green, J. C., 2005a. Comparison of blockage factors in modelling the resistance of channels containing
   submerged macrophytes. River Res Appl 21(6):671-686 doi:Doi 10.1002/Rra.854.
- Green, J. C., 2005b. Velocity and turbulence distribution around lotic macrophytes. Aquat Ecol 39(1):1 10 doi:DOI 10.1007/s10452-004-1913-0.
- 587 Gurnell, A. M., 2014. Plants as river system engineers. Earth Surf Process Landf 39(1):4-25.
- Hensley, R. T. & M. J. Cohen, 2012. Controls on solute transport in large spring-fed karst rivers. Limnol
   Oceanogr 57(4):912-924 doi:10.4319/lo.2012.57.4.0912.
- Horvath, T. G., 2004. Retention of particulate matter by macrophytes in a first-order stream. Aquat Bot
   78(1):27-36 doi:10.1016/j.aquabot.2003.09.003.
- Jackman, A. P., R. A. Walters & V. C. Kennedy, 1984. Transport and concentration controls for chloride,
   strontium, potassium and lead in Uvas Creek, a small cobble-bed stream in Santa-Clara County,
   California, USA .2. Mathematical-Modeling. J Hydrol 75(1-4):111-141 doi:Doi 10.1016/0022 1694(84)90047-7.
- Lamberti, G. A., S. V. Gregory, L. R. Ashkenas, R. C. Wildman, A. D. Steinman, 1996. Influence of
   channel geomorphology on retention of dissolved and particulate matter in a cascade mountain
   stream. USDA Forest Service Gen. Tech. Rep. PSW-110.
- Lees, M. J., L. A. Camacho & S. Chapra, 2000. On the relationship of transient storage and aggregated
   dead zone models of longitudinal solute transport in streams. Water Resour Res 36(1):213-224
   doi:Doi 10.1029/1999wr900265.
- Lightbody, A. F. & H. M. Nepf, 2006. Prediction of velocity profiles and longitudinal dispersion in
   emergent salt marsh vegetation. Limnol Oceanogr 51(1):218-228.
- Lopez, F. & M. H. Garcia, 2001. Mean flow and turbulence structure of open-channel flow through non emergent vegetation. J Hydraul Eng-Asce 127(5):392-402.
- Madsen, J. D., P. A. Chambers, W. F. James, E. W. Koch & D. F. Westlake, 2001. The interaction
   between water movement, sediment dynamics and submersed macrophytes. Hydrobiologia
   444(1-3):71-84 doi:Doi 10.1023/A:1017520800568.
- Muotka, T. & P. Laasonen, 2002. Ecosystem recovery in restored headwater streams: the role of enhanced
   leaf retention. J Appl Ecol 39(1):145-156 doi:DOI 10.1046/j.1365-2664.2002.00698.x.
- Nash, J. E., 1959. Systematic determination of unit hydrograph parameters. J Geophys Res 64(1):111-115
   doi:Doi 10.1029/Jz064i001p00111.
- Nepf, H. M., C. G. Mugnier & R. A. Zavistoski, 1997. The effects of vegetation on longitudinal dispersion. Estuar Coast Shelf S 44(6):675-684.
- Newbold, J. D., R. V. Oneill, J. W. Elwood & W. Vanwinkle, 1982. Nutrient Spiralling in Streams Implications for Nutrient Limitation and Invertebrate Activity. Am Nat 120(5):628-652 doi:Doi
   10.1086/284017.
- Nilsson, C., A. Ekblad, M. Gardfjell & B. Carlberg, 1991. Long-term effects of river regulation on river
   margin vegetation. J Appl Ecol 28(3):963-987 doi:Doi 10.2307/2404220.
- O'Hare, M. T., 2015. Aquatic vegetation a primer for hydrodynamic specialists. J Hydraul Res 53(6):687-698 doi:10.1080/00221686.2015.1090493.

- Old, G. H., P. S. Naden, P. Rameshwaran, M. C. Acreman, S. Baker, F. K. Edwards, J. P. R. Sorensen, O.
  Mountford, D. C. Gooddy, C. J. Stratford, P. M. Scarlett, J. R. Newman & M. Neal, 2014.
  Instream and riparian implications of weed cutting in a chalk river. Ecol Eng 71:290-300
  doi:10.1016/j.ecoleng.2014.07.006.
- Pedersen, F. B., 1977. Prediction of longitudinal dispersion in natural streams. Ser Pap 14(Inst. of
   Hydrodyn. and Hydraul. Eng., Univ. of Denmark, Lyngby).
- Pluntke, T. & H. P. Kozerski, 2003. Particle trapping on leaves and on the bottom in simulated submerged
   plant stands. Hydrobiologia 506(1-3):575-581 doi:Doi 10.1023/B:Hydr.0000008569.29286.Ec.
- Riis, T. & K. Sand-Jensen, 2006. Dispersal of plant fragments in small streams. Freshwater Biol 51(2):274-286 doi:10.1111/j.1365-2427.2005.01496.x.
- Rovira, A., C. Alcaraz & R. Trobajo, 2016. Effects of plant architecture and water velocity on sediment
   retention by submerged macrophytes. Freshwater Biol 61(5):758-768 doi:10.1111/fwb.12746.
- Runkel, R. L., 2007. Toward a transport-based analysis of nutrient spiraling and uptake in streams.
   Limnol Oceanogr-Meth 5:50-62.
- Sand-Jensen, K., K. Andersen & T. Andersen, 1999. Dynamic properties of recruitment, expansion and
   mortality of macrophyte patches in streams. Int Rev Hydrobiol 84(5):497-508.
- Sand-Jensen, K. & J. R. Mebus, 1996. Fine-scale patterns of water velocity within macrophyte patches in streams. Oikos 76(1):169-180 doi:Doi 10.2307/3545759.
- Sand-Jensen, K. & M. L. Pedersen, 2008. Streamlining of plant patches in streams. Freshwater Biol
   53(4):714-726 doi:DOI 10.1111/j.1365-2427.2007.01928.x.
- Schneider, R. L. & R. R. Sharitz, 1988. Hydrochory and Regeneration in a Bald Cypress Water Tupelo
   Swamp Forest. Ecology 69(4):1055-1063 doi:Doi 10.2307/1941261.
- Schoelynck, J., 2011. Macrophytes patches as biogeochemical hotspots: what is the impact on river water
   quality? University Press Antwerp.
- Schoelynck, J., D. Meire, K. Bal, K. Buis, P. Troch, T. Bouma, P. Meire & S. Temmerman, 2013.
  Submerged macrophytes avoiding a negative feedback in reaction to hydrodynamic stress. Limnologica 43(5):371-380 doi:DOI 10.1016/j.limno.2013.05.003.
- Seitzinger, S., J. A. Harrison, J. K. Bohlke, A. F. Bouwman, R. Lowrance, B. Peterson, C. Tobias & G.
  Van Drecht, 2006. Denitrification across landscapes and waterscapes: A synthesis. Ecol Appl 16(6):2064-2090 doi:Doi 10.1890/1051-0761(2006)016[2064:Dalawa]2.0.Co;2.
- Sukhodolova, T., A. Sukhodolov, H. P. Kozerski & J. Kohler, 2006. Longitudinal dispersion in a lowland
   river with submersed vegetation. River Flow 2006, Vols 1 and 2:631-638.
- Taylor, 1954. The dispersion of matter in solvent flowing slowly through a tube. Proc Royal Soc 222(1155):446-468.
- Thankston, E. I. & K. B. Schnelle, 1970. Predicting effects of dead zones on stream mixing. Journal of
   Environmental Engineering 96:319-331.
- van den Broek, T., R. van Diggelen & R. Bobbink, 2005. Variation in seed buoyancy of species in
  wetland ecosystems with different flooding dynamics. J Veg Sci 16(5):579-586 doi:Doi
  10.1658/1100-9233(2005)16[579:Visbos]2.0.Co;2.
- Vereecken, H., J. Baetens, P. Viaene, F. Mostaert & P. Meire, 2006. Ecological management of aquatic
   plants: effects in lowland streams. Hydrobiologia 570:205-210 doi:10.1007/s10750-006-0181-5.
- Verschoren, V., D. Meire, J. Schoelynck, K. Buis, K. D. Bal, P. Troch, P. Meire & S. Temmerman, 2016.
   Resistance and reconfiguration of natural flexible submerged vegetation in hydrodynamic river
   modelling Environ Fluid Mech doi:10.1007/s10652-015-9432-1.
- Warren, L. L., R. S. Wotton, G. Wharton, J. A. B. Bass & J. A. Cotton, 2009. The transport of fine
  particulate organic matter in vegetated chalk streams. Ecohydrology 2(4):480-491 doi:Doi
  10.1002/Eco.86.

- Wilcock, R. J., P. D. Champion, J. W. Nagels & G. F. Croker, 1999. The influence of aquatic macrophytes on the hydraulic and physico-chemical properties of a New Zealand lowland stream.
  Hydrobiologia 416:203-214 doi:Doi 10.1023/A:1003837231848.
- Wood, P. J. & P. D. Armitage, 1997. Biological effects of fine sediment in the lotic environment. Environ
   Manage 21(2):203-217 doi:DOI 10.1007/s002679900019.