

**This item is the archived peer-reviewed author-version of:**

Prevalence of Ureaplasma spp. and Mycoplasma hominis in healthy women and patients with flora alterations

**Reference:**

Rumyantseva Tatiana, Khayrullina Guzel, Guschin Alexander, Donders Gilbert.- Prevalence of Ureaplasma spp. and Mycoplasma hominis in healthy women and patients with flora alterations

Diagnostic microbiology and infectious disease - ISSN 0732-8893 - 93:3(2019), p. 227-231

Full text (Publisher's DOI): <https://doi.org/10.1016/J.DIAGMICROBIO.2018.10.001>

To cite this reference: <https://hdl.handle.net/10067/1585870151162165141>

## Accepted Manuscript

Prevalence of *Ureaplasma* spp. and *Mycoplasma hominis* in healthy women and patients with flora alterations

Tatiana Rumyantseva, Guzel Khayrullina, Alexander Guschin, Gilbert Donders



PII: S0732-8893(18)30441-3

DOI: doi:[10.1016/j.diagmicrobio.2018.10.001](https://doi.org/10.1016/j.diagmicrobio.2018.10.001)

Reference: DMB 14686

To appear in: *Diagnostic Microbiology & Infectious Disease*

Received date: 26 July 2018

Revised date: 25 September 2018

Accepted date: 1 October 2018

Please cite this article as: Tatiana Rumyantseva, Guzel Khayrullina, Alexander Guschin, Gilbert Donders , Prevalence of *Ureaplasma* spp. and *Mycoplasma hominis* in healthy women and patients with flora alterations. *Dmb* (2018), doi:[10.1016/j.diagmicrobio.2018.10.001](https://doi.org/10.1016/j.diagmicrobio.2018.10.001)

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

**Prevalence of *Ureaplasma spp.* and *Mycoplasma hominis* in healthy women and patients  
with flora alterations.**

**Running title.** Mycoplasmas in normal and altered flora.

Tatiana Rumyantseva, PhD, <sup>1</sup> Guzel Khayrullina, PhD, <sup>2</sup> Alexander Guschin, PhD, <sup>1</sup> Gilbert Donders, PhD.<sup>3,4</sup>

1. Laboratory of molecular diagnostics of reproductive tract infections, Federal Budget Institute of Science “Central Research Institute for Epidemiology” of Rospotrebnadzor, 111123, Novogireevskaya st. 3a, Moscow, Russia
2. Laboratory for studying pathogenesis and clinics of socially important infectious and parasitic diseases, Peoples’ Friendship University of Russia, 117198, Moscow Miklukho-Maklaya str.6, Moscow, Russia
3. Femicare Clinical Research for Women, 3300, Gasthuismolenstraat 31, Tienen, Belgium
4. Department of Obstetrics and Gynecology, Antwerp University, 2000, Prinsstraat 13, Antwerp, Belgium

**Corresponding author:** T.A. Rumyantseva (ivanovatatiana86@yandex.ru), 111123

Novogireevskaya st., 3a, Moscow, Russia.

Tel (personal): +79161547067,

Tel (business): +74959749646

Fax: +74953042209

**Abstract**

**Objective.** To estimate the prevalence of *Mycoplasma hominis*, *Ureaplasma parvum*, *Ureaplasma urealyticum* in healthy women and patients with altered vaginal microflora.

**Methods.** Vaginal samples from 2594 unselected female patients were divided into Normal, BV and AV groups and tested for *U. parvum*, *U. urealyticum* and *M. hominis*.

**Results.** Normal flora was detected in 1773 patients (68.4%), BV - in 754 patients (29.1%), and AV - in 67 patients (2.6%). In the control group 771 (43.5%) patients were *U. parvum* positive, 104 (5.9%) - *U. urealyticum* positive, 158 (8.9%) - *M. hominis* positive. In the BV group those bacteria were detected in 452 (59.9%), 102 (13.5%) and 202 (26.8%) patients, respectively ( $p < 0.001$ ); in AV group – in 16 (23.9%), 3 (4.5%) and 4 (6.0%) patients, respectively ( $p < 0.001$ ; 0.63 and 0.40, respectively).

**Conclusions.** This study demonstrated that mycoplasmas may be a marker or a symbiont of the BV flora, but not AV flora.

**Keywords:** bacterial vaginosis, aerobic vaginitis, *Mycoplasma hominis*, *Ureaplasma parvum*, *Ureaplasma urealyticum*, polymerase chain reaction.

## 1. Introduction

Genital mycoplasmas are a diverse group of bacteria variable in virulence and pathogenicity. *Mycoplasma genitalium* is a sexually transmitted bacteria and is a cause of urethritis in males<sup>1</sup> and cervicitis, PID, preterm birth and spontaneous abortion<sup>2</sup>. *Mycoplasma hominis* and *Ureaplasma spp.* (*Ureaplasma parvum* and *Ureaplasma urealyticum*) are opportunistic bacteria found in both healthy women and severe diseases and complications during or outside pregnancy<sup>3, 4</sup>. Prevalence of mycoplasmas varies in many studies (4.5-40%, 28.8-63.8%, 1.3-51% for *U.urealyticum*, *U.parvum* and *M.hominis*, respectively<sup>3, 5-8</sup> reaching very high numbers even for healthy individuals in some studies: 85.2% (summarized prevalence for *U.urealyticum*, *U.parvum* and *M.hominis*<sup>9</sup>) and 95.5% for *U.parvum*<sup>10</sup>.

*M. hominis* is associated with a very common flora alteration - bacterial vaginosis (BV)<sup>3, 11</sup>, whereas the role or association of *Ureaplasma spp.* and BV is still a topic for discussion<sup>10</sup>. Apart from flora alterations, *M. hominis* and *Ureaplasma spp.* have been linked to a number of morbidities: pyelonephritis, PID, chorioamnionitis, endometritis, postpartum fever, infertility, low birth weight, spontaneous abortion, stillbirth, preterm delivery, perinatal mortality<sup>4, 8, 12-15</sup>.

As *M. hominis* and *Ureaplasma spp.* are found both in healthy women as well as in patients with BV and other morbidities, the need for the detection of those species in female patients is being argued. At the moment it is suggested that detection of *Mycoplasma hominis* and *Ureaplasma spp.* alone without flora evaluation seems useless in women<sup>3</sup>.

Flora alterations include not only BV but also symptomatic *Trichomonas vaginalis* infection, *Candida* vaginitis and aerobic vaginitis (AV)<sup>16</sup>. *M. hominis* and *Ureaplasma spp.* are frequently found in BV-positive patients, whereas their prevalence in AV-positive patients has not been thoroughly investigated yet. Investigation of the association between mycoplasmas and two types of flora alteration (BV and AV) may give a deeper insight if mycoplasmas are associated with elevated pH in BV or AV - positive women, are a specific constituent of BV

only, or whether they are pathogenic without that lactobacillus deficient climate found in BV/AV – positive patients. Despite the focus on the prevalence of mycoplasmas in women with healthy or BV or AV type vaginal flora, some loads of *M. hominis* and *Ureaplasma* spp. may play a role in the balance of health and disease as well. Thus the aim of the present study was to evaluate the prevalence and loads of *U.parvum*, *U.urealyticum* and *M.hominis* in healthy, BV-positive and AV-positive women.

## 2. Methods

### 2.1.Samples.

A total of 2705 vaginal samples from reproductive-aged female patients were included in this study. Samples were collected from both asymptomatic women (planning pregnancy; after unprotected intercourse or requesting STI testing for any other reasons) and patients with vaginal complaints attending gynecologists in a variety of clinics in Moscow, Russia (clinics sending samples for laboratory testing to Federal Budget Institute of Science “Central Research Institute for Epidemiology”), from August to September 2015. All participants signed the informed consent form.

### 2.2.Testing.

Vaginal samples from all participants were tested by real-time PCR. DNA-sorb-AM kit (InterLabService, Russia) was used for DNA extraction. The AmpliSens® N.gonorrhoeae/C.trachomatis/M.genitalium/T.vaginalis-MULTIPRIME-FRT kit was used to detect the DNA of four STIs (*Chlamydia trachomatis*, *Neisseria gonorrhoeae*, *Mycoplasma genitalium*, *Trichomonas vaginalis*)<sup>17</sup>. Samples positive for one or more of these STIs were excluded from the further analysis. Flora was further assessed using AmpliSens® Florocenosis/Aerobes-FRT (detection and quantification of *Enterobacteriaceae* spp., *Staphylococcus* spp., *Streptococcus* spp.)<sup>18</sup> and AmpliSens® Florocenosis/Bacterial vaginosis-FRT (detection and quantification of total bacterial DNA, *Lactobacillus* spp, *G.vaginalis*, *A.vaginae*)<sup>19</sup> kits (InterLabService, Russia). Those

kits allow discrimination between normal flora, BV and AV-like flora by making the distinction between 1) normal flora with dominance of *Lactobacilli*, 2) BV with dominance of *G.vaginalis*/ *A.vaginae* in the presence of low *Lactobacilli* counts and 3) AV with dominance *Enterobacteriaceae spp.*/ *Staphylococcus spp.*/ *Streptococcus spp.* with low *Lactobacilli* counts.

Calculations and interpretation were performed based on the DNA loads of the detected bacteria, according to the manufacturer's instructions. In brief, the ratio coefficients (RC) for AmpliSens® Florocenosis/Bacterial vaginosis-FRT were calculated as follows:  $RC1 = \lg [Lactobacillus \text{ spp.}] - \lg [G. vaginalis + A. vaginae]$ ,  $RC2 = \lg [Bacteria] - \lg [Lactobacillus \text{ spp.}]$ , and  $RC3 = \lg [Bacteria] - \lg [G. vaginalis + A. vaginae]$ .

In summary, the result of the test was interpreted as normal vaginal microbiota (Control group), if *G. vaginalis* and/or *A. vaginae* as well as *Enterobacteriaceae spp.* + *Staphylococcus spp.* + *Streptococcus spp.* were absent or their cumulative load was less than the *Lactobacillus spp.* load. A sample was categorized as BV (BV group), if the *G. vaginalis* and/or *A. vaginae* loads were equal to or exceeded the *Lactobacillus spp.* load ( $RC1 < 0.5$ ). Finally, a sample was categorized as AV-like flora (AV group), if the *Lactobacillus spp.* load was decreased and the *G. vaginalis* and/or *A. vaginae* load was substantially lower than the load of total bacteria ( $RC2 > 1$  and  $RC3 > 2$ ) with summarized load of *Enterobacteriaceae spp.* + *Staphylococcus spp.* + *Streptococcus spp.* dominating over both *Lactobacillus spp.* and *G.vaginalis*/ *A.vaginae*. This analysis allowed us to divide the sample into BV, AV and Control groups.

*U. parvum*, *U. urealyticum* and *M. hominis* DNA were detected and quantified in all samples using AmpliSens® Florocenosis/Mycoplasma-FRT kit. Testing was performed according to manufacturer's instructions.

### 2.3. Data analysis.

Descriptive statistics used for summarizing quantitative PCR results included median values. For testing differences between the groups of patients, chi-square statistics were used for categorical variables (presence of bacteria), and the Mann-Whitney U test was used for loads of bacteria. All tests for significance were two-sided, and statistically significant differences were assumed when  $p < 0.05$ .

## 2.4. Funding.

This work was supported by the Ministry of Education and Science of Russian Federation, project no. 03.G25.31.0226, March 3, 2017.

## 3. Results.

### 3.1. Prevalence

After exclusion of STI-positive samples, 2594 samples (mean age  $31 \pm 5$  y.o.) were included in the analysis. Normal lactobacillary-dominated flora was detected in 1773 patients (68.4%), BV - in 754 patients (29.1%), and AV - in 67 (2.6%) patients (**Figure 1**).

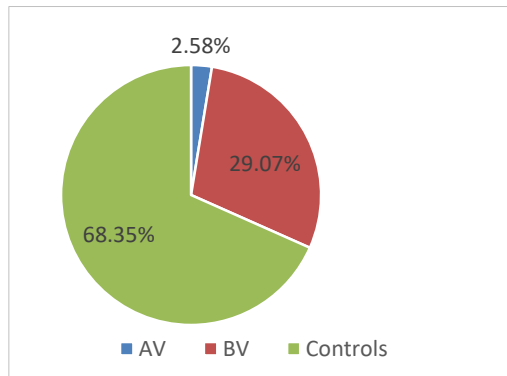


Figure 1. Results of flora assessment in 2594 participants.

AV – aerobic vaginitis; BV – bacterial vaginosis.

DNA of one or more Mycoplasmas was discovered in 1475 (56.9%) women. *U. parvum* DNA was detected in 1239 (47.8%) patients, *U. urealyticum* – in 209 (8.1%) patients, *M. hominis* – in 364 (14.0%) patients. In the control group 771 (43.5%) patients were *U. parvum* positive, 104 (5.9%) patients were *U. urealyticum* positive and 158 (8.9%) patients were *M. hominis* positive. In the BV group those bacteria were detected in 452 (59.9%), 102



(13.5%) and 202 (26.8%) patients, respectively ( $p < 0.001$  versus Controls); in the AV group – in 16 (23.9%), 3 (4.5%) and 4 (6.0%) patients, respectively ( $p$ -values versus Controls  $< 0.001$ ; 0.63 and 0.40, respectively;  $p$ -values versus BV group  $< 0.001$ ; 0.034;  $< 0.001$ , respectively). *U. parvum*, *U. urealyticum* and *M. hominis* were significantly more prevalent in the BV group ( $p < 0.001$ ) compared to Control and AV groups. *U. parvum* was significantly less prevalent ( $p < 0.001$ ) in the AV group compared to the Control group (Figure 2).

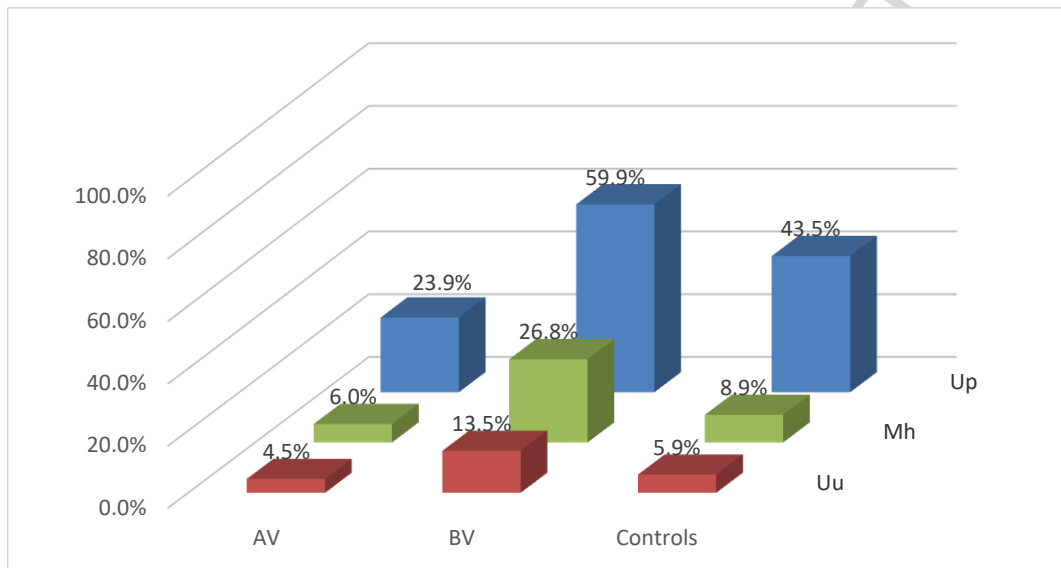


Figure 2. Prevalence of *U. parvum*, *U. urealyticum* and *M. hominis* in the BV, AV and Control groups.

AV – aerobic vaginitis; BV – bacterial vaginosis.

Uu - *U. urealyticum*, Mh - *M. hominis*, Up - *U. parvum*

### 3.2. Bacterial loads

Median loads of *U. parvum* and *M. hominis* were significantly higher in BV group compared to AV and Control groups ( $p < 0.001$ ). Median load of *U. urealyticum* did not vary significantly in three groups ( $p = 0.21$ ) (Table 1, Figure 3)

Table 1.

Median loads of *U. parvum*, *U. urealyticum* and *M. hominis* in BV, AV and Control groups (Geq/ml).

	AV	BV	Control	p-value
<i>U.parvum</i>	$1*10^5$	$1*10^6$	$2*10^5$	AV vs Control p=0.30 BV vs Control p<0.001
<i>U.urealyticum</i>	$1*10^5$	$8*10^4$	$1*10^4$	p=0.21
<i>M.hominis</i>	$5*10^2$	$1.5*10^6$	$2*10^2$	AV vs Control p=0.50 BV vs Control p<0.001

AV – aerobic vaginitis; BV – bacterial vaginosis.

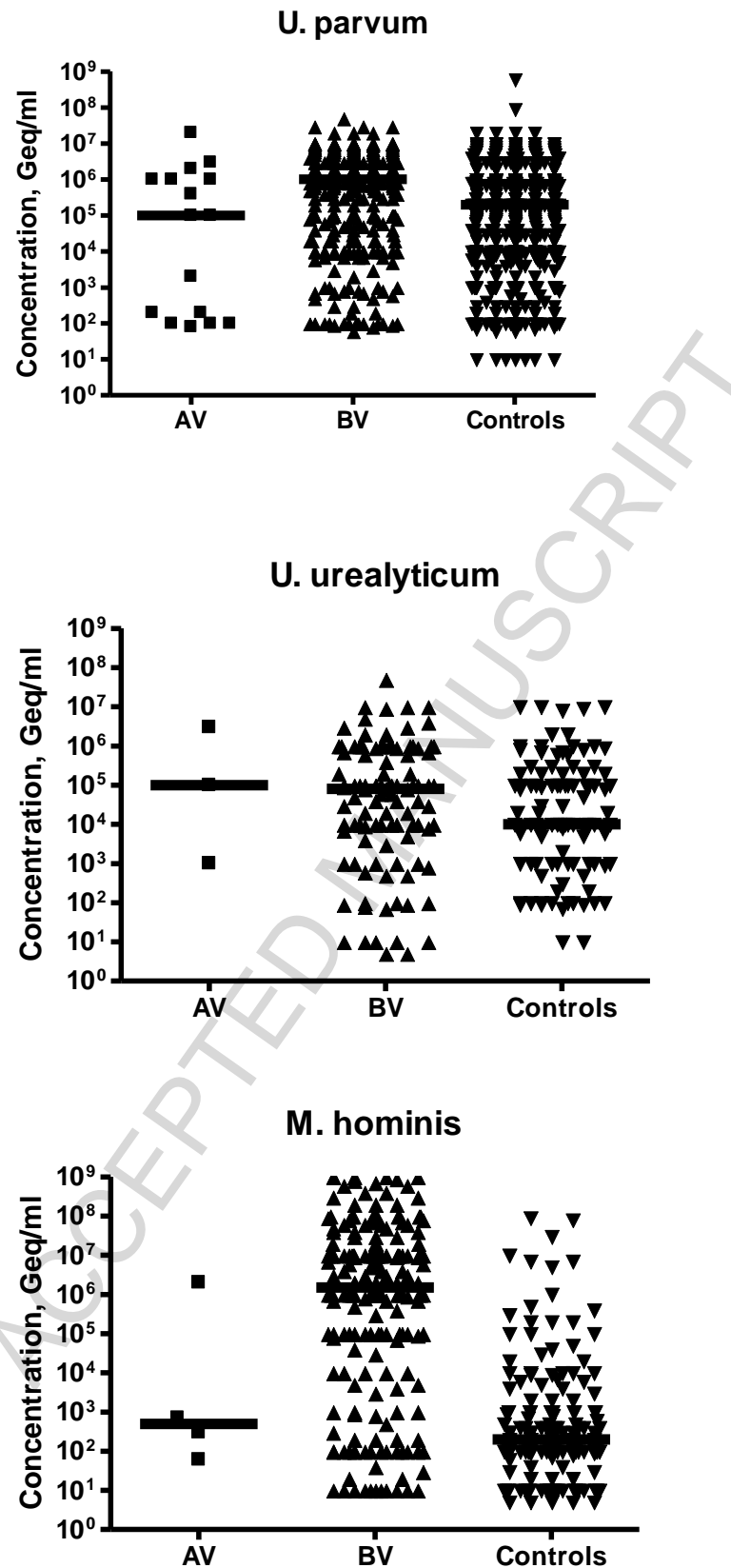


Figure 3. *U. parvum*, *U. urealyticum* and *M. hominis* load distribution in BV, AV and Control groups (Geq/ml). Horizontal bar = median load of mycoplasmas in each group. AV – aerobic vaginitis; BV – bacterial vaginosis.

Overall *U. parvum* was the most prevalent mycoplasma in all groups, whereas *M. hominis* was most strongly associated with BV (demonstrated a 3-fold increase in prevalence and a 7500 - fold increase in load compared to the Control group,  $p < 0.001$ ).

#### **4. Discussion.**

The role of mycoplasmas (*U. parvum*, *U. urealyticum* and *M. hominis*) in the reproductive health and disease is still being evaluated. Conflicting results are being reported, and one of the reasons for that may be the insufficient evaluation of vaginal microflora in some studies devoted to mycoplasmas only. Thus in this study we investigated the prevalence of mycoplasmas in healthy, BV-positive and AV-positive women.

##### **4.1. Main Findings.**

All mycoplasmas were less prevalent in the AV group as compared to the BV group; *U. parvum* was less prevalent in the AV group compared to Controls; loads of mycoplasmas did not vary significantly in AV and Control groups, whereas *U. parvum* and *M. hominis* were detected in BV-positive patients in significantly higher loads.

##### **4.2. Interpretation.**

In our setting of 2594 female patients BV was detected in 29.1%, AV in 2.6% and normal flora in 68.4% of women. These data correlate well with known prevalence, BV being the most prevalent lower genital tract bacterial infection in women of reproductive age worldwide<sup>20</sup>. In most settings the real prevalence of BV may be lower than described before, due to the underestimation of AV, e.g. Donders et al. found BV in 25% and AV in 11% of women in Uganda, whereas most authors claim that 35 to 50% prevalence of BV in central African populations<sup>21</sup>. Prevalence of AV in our study is lower than the prevalence demonstrated in other studies, as AV was encountered in 8.3%-10.8% of pregnant women<sup>22, 23</sup> and in 5%-23.7% of women reporting vaginal complaints<sup>21, 24-26</sup>. Besides accepting real differences between specific

populations, this may also be due to the variations of diagnostic tools used or to the specific population studied and the proportion of symptomatic patients.

The prevalence of mycoplasmas in this study was lower than in previous studies. Cox et al. detected *U. parvum* in 78.6% of BV-positive patients and in 61,3% of healthy controls<sup>27</sup>. In our study *U. parvum* was detected in 59.9% of BV-positive patients and 43.5% of healthy controls. *U. urealyticum* was previously detected in 65% of BV-positive patients and in 48% of healthy controls by Keane et al.<sup>28</sup> and in 17.6% of BV-positive patients and in 22.5% of healthy controls by Cox et al<sup>27</sup>. We report the prevalence of *U. urealyticum* being 13.5% and 5.9% in BV and healthy women, respectively. For *M. hominis*, a prevalence of 60.7% and 11.3%; 81% and 31% in BV-positive and healthy women was reported previously<sup>27, 29</sup>, whereas in our study *M. hominis* was detected in only 26.8% and 8.9% of participants, respectively. In one study, *M. hominis* was not detected in healthy controls<sup>28</sup>, demonstrating even lower prevalence than in our study, but in the BV group they still found a prevalence of 53%, which is much higher compared to our results. In one study a 17% prevalence of *M. hominis* was demonstrated in the “AV-like group”<sup>30</sup>, compared to 6% in our study. Further comparison of the prevalence of mycoplasmas in AV-positive patients is impossible due to the lack of data.

According to our data, prevalence of all mycoplasmas was significantly higher in the BV-group than in both the AV group and healthy controls. This leads us to the conclusion that mycoplasmas need more to thrive than a favorable pH in the environment, but may survive better in the symbiotic relationships with anaerobic BV-associated bacteria, e.g. *Gardnerella vaginalis*, *Atopobium vaginae*, *Mobiluncus spp.* and other anaerobes. The most prominent difference was observed by the 3-fold increase in prevalence for *M. hominis* between healthy and BV-positive women, which is in line with previous findings and supports the suggestion made by Cox et al. that *M. hominis* and *G. vaginalis* have a symbiotic relationship<sup>27</sup>. Some previous studies did not find any increase in the prevalence of *Ureaplasma spp.* in BV-positive women versus healthy controls<sup>27, 28</sup>, whereas in our study both *Ureaplasmas* were more prevalent in the BV group.

Ethnical characteristics of the groups, different diagnostic tools used, various principles of groups' formation (e.g. selection of AV-positive patients in our study) could have accounted for that difference.

In our study, quantitative parameters (DNA load) showed no significant variation of *U. urealyticum* in the normal, BV or AV groups, similar to their bacterial loads found in BV-positive and healthy women in Cox et al's study<sup>27</sup>. Compared to both healthy and AV-positive participants the load of *U. parvum* in BV-positive women was significantly higher in our study, whereas no significant difference was observed by Cox et al.<sup>27</sup> This disagreement may be explained by varying quantification techniques and sample size and variations in populations studied. Quantitative data for *M. hominis* load, on the other hand, correlate well with earlier studies and demonstrated a 7500-fold increase in BV-positive participants versus controls, which is in agreement with findings by Rosenstein et al.<sup>30</sup>

#### **4.3.Strengths and Limitations.**

Discussion of the features of the present study requires special attention to the laboratory methods used. We used molecular-based techniques for the detection and quantification of mycoplasmas as well as for the flora assessment. Although this technique does not include clinical data or microscopy findings, it demonstrated good ability for BV, AV and normal flora discrimination in previous studies<sup>17-19</sup>. In the majority of previous studies of other authors, on the other hand, culture-based tools were used for the detection and quantification of mycoplasmas, and/or non-molecular methods, like microscopy were used to diagnose microflora abnormalities. The PCR-based assay used in the current study, however, has two major advantages: standardized quantification of the microorganisms, validated testing of microflora alterations, and reliable discrimination between *Ureaplasma* species (*U.parvum* and *U.urealyticum*).

The present study has three major limitations: lack of clinical data, relatively small amount of AV-positive patients and no opportunity for follow-up (to determine prevalence and load after BV or AV treatment).

## 5. Conclusion.

This study demonstrates that mycoplasmas, in special *M. hominis*, are associated with BV, but not with AV. Vaginal mycoplasmas even seem to be slightly less prevalent in women with AV, but larger numbers are needed to prove this is statistically significant. Further studies are needed to demonstrate if flora normalization or treatment leads to the elimination or decrease of the DNA load of *U.parvum*, *U.urealyticum* and/or *M.hominis* in the vaginal fluid.

## 6. Disclosure of Interests

No conflict of interests declared.

## 7. Contribution to Authorship

TR performed data analysis, produced the draft of the paper, worked on the paper text after corrections and comments of other authors, approved the final version of the manuscript. GK performed data collection, data analysis, produced the draft of the paper, approved the final version of the manuscript. AG planned the project, read and revised the manuscript, and approved the final version. GD read and revised the manuscript, and approved the final version. All authors accept responsibility for the paper as published.

## 8. Details of ethics approval

This study was approved by the Ethical Committee of the Federal Budget Institute of Science “Central Research Institute for Epidemiology” of Rospotrebnadzor on June, 26, 2015 (Protocol #34).

**9. Funding:**

This work was supported by the Ministry of Education and Science of Russian Federation, project no. 03.G25.31.0226, March 3, 2017

ACCEPTED MANUSCRIPT



## 10. References

1. Taylor-Robinson D, Jensen JS. *Mycoplasma genitalium*: from Chrysalis to Multicolored Butterfly. Clin Microbiol Rev. 2011;24:498–514.
2. Lis R, Rowhani-Rahbar A, Manhart LE. *Mycoplasma genitalium* infection and female reproductive tract disease: a meta-analysis. Clin Infect Dis. 2015;61:418–26.
3. Donders GGG, Ruban K, Bellen G, Petricevic L. *Mycoplasma/Ureaplasma* infection in pregnancy: to screen or not to screen. J Perinat Med. 2017;45(5):505-15. doi: 10.1515/jpm-2016-0111.
4. Taylor-Robinson D, Lamont RF. Mycoplasmas in pregnancy. BJOG. 2011;118(2):164-74
5. Camporiondo MP, Farchi F, Ciccozzi M, Denaro A, Gallone D, Maracchioni F, et al. Detection of HPV and co-infecting pathogens in healthy Italian women by multiplex real-time PCR. Infez Med. 2016;24(1):12-7.
6. Tomusiak A, Heczko PB, Janeczko J, Adamski P, Pilarczyk-Zurek M, Strus M. Bacterial infections of the lower genital tract in fertile and infertile women from the southeastern Poland. Ginekol Pol. 2013;84(5):352-8.
7. Menard JP, Fenollar F, Henry M, Bretelle F, Raoult D. Molecular quantification of *Gardnerella vaginalis* and *Atopobium vaginae* loads to predict bacterial vaginosis. Clin Infect Dis 2008;47:33–43.
8. Cox C, Saxena N, Watt AP, Gannon C, McKenna JP, Fairley DJ, et al. The common vaginal commensal bacterium *Ureaplasma parvum* is associated with chorioamnionitis in extreme preterm labor. J Matern Fetal Neonatal Med. 2016;26:1-6.
9. Chaban B, Links MG, Jayaprakash TP, Wagner EC, Bourque DK, Lohn Z, et al. Characterization of the vaginal microbiota of healthy Canadian women through the menstrual cycle. Microbiome. 2014;2:23.
10. Marovt M, Keše D, Kotar T, Kmet N, Miljković J, Šoba B, et al. *Ureaplasma parvum* and *Ureaplasma urealyticum* detected with the same frequency among women with and

- without symptoms of urogenital tract infection. *Eur J Clin Microbiol Infect Dis.* 2015;34(6):1237-45.
11. Taylor-Robinson D, Rosenstein IJ. Is *Mycoplasma hominis* a vaginal pathogen? *Sex Transm Infect.* 2001;77(4):302.
  12. Kataoka S, Yamada T, Chou K et al. Association between preterm birth and vaginal colonization by mycoplasmas in early pregnancy. *J Clin Microbiol.* 2006;44(1):51-5.
  13. Kwak DW, Hwang HS, Kwon JY, Park YW, Kim YH. Co-infection with vaginal *Ureaplasma urealyticum* and *Mycoplasma hominis* increases adverse pregnancy outcomes in patients with preterm labor or preterm premature rupture of membranes. *J Matern Fetal Neonatal Med.* 2014;27(4):333-7. doi: 10.3109/14767058.2013.818124.
  14. Capoccia R, Greub G, Baud D. *Ureaplasma urealyticum*, *Mycoplasma hominis* and adverse pregnancy outcomes. *Curr Opin Infect Dis.* 2013;26(3):231-40. doi: 10.1097/QCO.0b013e328360db58.
  15. Haggerty CL, Totten PA, Tang G Astete SG, Ferris MJ, Norori J, et al. Identification of novel microbes associated with pelvic inflammatory disease and infertility. *Sex Transm Infect.* 2016;92(6):441-6. doi: 10.1136/sextrans-2015-052285.
  16. Donders GG, Vereecken A, Bosmans E, Dekeersmaecker A, Salembier G, Spitz B. Definition of a type of abnormal vaginal flora that is distinct from bacterial vaginosis: aerobic vaginitis. *BJOG.* 2002;109(1):34-43.
  17. Rumyantseva T, Golparian D, Nilsson CS, Johansson E, Falk M, Fredlund H, et al. Evaluation of the new AmpliSens multiplex real-time PCR assay for simultaneous detection of *Neisseria gonorrhoeae*, *Chlamydia trachomatis*, *Mycoplasma genitalium*, and *Trichomonas vaginalis*. *APMIS.* 2015;123(10):879-86. doi: 10.1111/apm.12430.
  18. Rumyantseva TA, Bellen G, Savochkina YA, Guschin AE, Donders GG. Diagnosis of aerobic vaginitis by quantitative real-time PCR. *Arch Gynecol Obstet.* 2016;294(1):109-14. doi: 10.1007/s00404-015-4007-4

19. Rumyantseva TA, Bellen G, Romanuk TN, Shipulina OI, Guschin AE, Shipulin GA, et al. Utility of Microscopic Techniques and Quantitative Real-time Polymerase Chain Reaction for the Diagnosis of Vaginal Microflora Alterations. *J Low Genit Tract Dis.* 2015;19(2):124-28. doi: 10.1097/LGT.0000000000000060.
20. Schwebke JR. New concepts in the etiology of bacterial vaginosis. *Curr Infect Dis Rep.* 2009;11(2):143-7.
21. Donders GG, Gonzaga A, Marconi C, Donders F, Michiels T, Eggermont N, et al. Increased vaginal pH in Ugandan women: what does it indicate? *Eur J Clin Microbiol Infect Dis.* 2016;35(8):1297-303. doi: 10.1007/s10096-016-2664-2.
22. Donders GGG, Van Calsteren K, Bellen G, Reybrouck R, Van den Bosch T, Riphagen I, et al. Predictive value for preterm birth of abnormal vaginal flora, bacterial vaginosis and aerobic vaginitis during the first trimester of pregnancy. *BJOG.* 2009;116(10):1315–24. doi:10.1111/j.1471-0528.2009.02237.x
23. Zodzika J, Rezeberga D, Jermakova I, Vasina O, Vedmedovska N, Donders G. Factors related to elevated vaginal pH in the first trimester of pregnancy. *Acta Obstet Gynecol Scand.* 2011;90(1):41–6. doi:10.1111/j.1600-0412.2010.01011.x
24. Fan A, Yue Y, Geng N, Zhang H, Wang Y, Xue F. Aerobic vaginitis and mixed infections: comparison of clinical and laboratory findings. *Arch Gynecol Obstet.* 2013;287(2):329-35. doi: 10.1007/s00404-012-2571-4.
25. Bologno R, Díaz YM, Giraudo MC, Fernández R, Menéndez V, Brizuela JC, et al. Importance of studying the balance of vaginal content (BAVACO) in the preventive control of sex workers. *Rev Argent Microbiol.* 2011;43(4):246-50. doi: 10.1590/S0325-75412011000400002.
26. Marconi C, Donders GG, Bellen G, Brown DR, Parada CM, Silva MG. Sialidase activity in aerobic vaginitis is equal to levels during bacterial vaginosis. *Eur J Obstet Gynecol Reprod Biol.* 2013;167(2):205-9. doi: 10.1016/j.ejogrb.2012.12.003.

27. Cox C, Watt AP, McKenna JP, Coyle PV. *Mycoplasma hominis* and *Gardnerella vaginalis* display a significant synergistic relationship in bacterial vaginosis. *Eur J Clin Microbiol Infect Dis*. 2016;35(3):481-7. doi: 10.1007/s10096-015-2564-x.
28. Keane FE, Thomas BJ, Gilroy CB, Renton A, Taylor-Robinson D. The association of *Mycoplasma hominis*, *Ureaplasma urealyticum* and *Mycoplasma genitalium* with bacterial vaginosis: observations on heterosexual women and their male partners. *Int J STD AIDS*. 2000;11(6):356-60.
29. Zozaya-Hinchliffe M, Lillis R, Martin DH, Ferris MJ. Quantitative PCR assessments of bacterial species in women with and without bacterial vaginosis. *J Clin Microbiol*. 2010;48(5):1812-9.
30. Rosenstein IJ, Morgan DJ, Sheehan M, Lamont RF, Taylor-Robinson D. Bacterial vaginosis in pregnancy: distribution of bacterial species in different gram-stain categories of the vaginal flora. *J Med Microbiol*. 1996;45(2):120-6.

Figure 1. Results of flora assessment in 2594 participants.

AV – aerobic vaginitis; BV – bacterial vaginosis.

Figure 2. Prevalence of *U. parvum*, *U. urealyticum* and *M. hominis* in the BV, AV and Control groups.

AV – aerobic vaginitis; BV – bacterial vaginosis.

Uu - *U. urealyticum*, Mh - *M. hominis*, Up - *U. parvum*

Figure 3. *U. parvum*, *U. urealyticum* and *M. hominis* load distribution in BV, AV and Control groups (Geq/ml). Horizontal bar = median load of mycoplasmas in each group. AV – aerobic vaginitis; BV – bacterial vaginosis.

## Highlights

- Genital mycoplasmas (*U. parvum*, *U. urealyticum* and *M. hominis*) are positively associated with bacterial vaginosis (BV), not aerobic vaginitis (AV).
- *U. parvum* and *M. hominis* are detected in higher loads among patients with BV compared to healthy individuals and patients with AV.
- Genital mycoplasmas may have symbiotic relationship with BV-associated microflora rather than be attracted by elevated pH in BV-positive patients.

ACCEPTED MANUSCRIPT