

**This item is the archived peer-reviewed author-version of:**

Performance of aerobic nitrite granules treating an anaerobic pre-treated wastewater originating from the potato industry

**Reference:**

Dobbeleers Thomas, Daens Dominique, Miele Solange, D' aes Jolien, Caluw é Michel, Geuens Luc, Dries Jan.- Performance of aerobic nitrite granules treating an anaerobic pre-treated wastewater originating from the potato industry  
Bioresource technology - ISSN 0960-8524 - 226(2017), p. 211-219  
Full text (Publisher's DOI): <https://doi.org/10.1016/J.BIORTECH.2016.11.117>  
To cite this reference: <https://hdl.handle.net/10067/1387820151162165141>

1 **Title:** Performance of aerobic nitrite granules treating an anaerobic pre-treated  
2 wastewater originating from the potato industry.

3

4 **Authors:** Thomas Dobbeleers<sup>a\*</sup>, Dominique Daens<sup>a</sup>, Solange Miele<sup>b</sup>, Jolien D'aes<sup>a</sup>,  
5 Michel Caluwé<sup>a</sup>, Luc Geuens<sup>a</sup> and Jan Dries<sup>a</sup>

6 **Affiliations:**

7 <sup>a</sup>Research group BioGEM, Bio-Chemical Green Engineering & Materials, Faculty of  
8 Applied Engineering, University of Antwerp, Salesianenlaan 90, 2660 Antwerp,  
9 Belgium

10 <sup>b</sup>National University of Quilmes, Basic and Applied Microbiology Institute, Genetic  
11 Engineering and Cellular and Molecular Biology, Buenos Aires, Argentina

12 **ABSTRACT**

13 In this study nitrogen removal via nitrite > 80% was achieved after approximately 80  
14 days in an Sequencing Batch Reactor (SBR) treating pre-treated industrial wastewater  
15 originating from the potato industry. Thereafter SBR performance was investigated  
16 during the formation of aerobic nitrite granules (ANG). The first granules appeared after  
17 26 days leading to full granulation after 64 days. ANG showed excellent settling  
18 properties, as the Sludge Volume Index (SVI) went down to 16 mL/g and a SVI<sub>10</sub>/SVI<sub>30</sub>  
19 =1 was obtained. qPCR analysis showed that slow growing organisms, especially  
20 Polyphosphate Accumulating Organisms (PAO) were stimulated by an anaerobic  
21 feeding strategy. The average nitrogen removal was 95.3% over the entire operational  
22 period, and it mainly followed the “nitrite-route”. Moreover, with ANG also phosphorus  
23 removal efficiencies up to 65.7% could be achieved. However, it has to be mentioned

24 that nitrous oxide was an important denitrification product, which implies some  
25 environmental concerns.

26

27 **KEYWORDS:** aerobic granular sludge; nitrite-pathway; nitrous oxide; industrial  
28 wastewater; control strategies, PAO

29

## 30 **1. INTRODUCTION**

31 Industrial wastewaters are characterized by a great variability in terms of (bio)chemical  
32 composition pH, temperature, intermittent and shock loadings which are all inseparably  
33 dependent of the variety in industrial activities (Rosenwinkel et al. 2014). During the  
34 last century, many activated sludge (AS) technologies such as conventional activated  
35 sludge (CAS) systems, multiple stages AS system, membrane bioreactor (MBR)  
36 systems, hybrid processes and discontinue-flow sequencing batch reactor (SBR)  
37 processes were implemented for industrial wastewaters (Rosenwinkel et al. 2014).

38

39 Typically, potato processing industries deal with high-strength wastewater as it contains  
40 high amounts of starch, proteins, amino acids and sugars. On one hand, this high  
41 organic load is favourable for primary anaerobic pre-treatment as this initial treatment  
42 step goes along with the production of biogas. However, because proteins and amino  
43 acids are a primary source of nitrogen (N) and potatoes contain 0.6 mg/g phosphorus (P)  
44 (Bosak et al., 2016), this wastewater contains a substantial amount of nutrients as well.  
45 Biological nutrient removal (BNR) processes require a sufficient fraction of easily  
46 biodegradable COD. Therefore, a significant bypass of raw potato processing  
47 wastewater is unavoidable to achieve an enhanced BNR performance.

48

49 A combination of simultaneous nitrification-denitrification and aerobic granular sludge  
50 (Bassin et al., 2012; Yilmaz et al., 2008) might be an interesting **strategy** to reduce the  
51 amount of bypass **needed** to purge anaerobic pre-treated wastewaters, or in general for  
52 wastewaters with a low COD/N ratio. The main advantages of a nitrogen removal via  
53 nitrite are: (1) 25% less oxygen demand during nitrification; (2) 40% less COD demand  
54 during denitrification, (3) 20% reduction in CO<sub>2</sub> emission and (4) 30-50% less sludge  
55 production (Peng and Zhu, 2006). **In addition**, aerobic granular sludge has the potential  
56 for simultaneous removal of nutrients in a single tank (N and P) (De Kreuk et al., 2005;  
57 Lochmatter et al., 2013). Simultaneous nitrification-denitrification can be achieved  
58 through the formation of anoxic zones, which in granules typically occur near the core  
59 as a result of the decreasing oxygen-gradient (Beun et al., 2001; Zeng et al., 2003).  
60 Furthermore, **by periodically alternating anaerobic and aerobic conditions**, phosphorus  
61 removal can be achieved as well (De Kreuk et al., 2005; Lochmatter et al., 2013). Under  
62 phosphate rich conditions, polyphosphate accumulating organisms (PAO) can store  
63 carbon sources such as volatile fatty acids (VFA) as polyhydroxyalkanoates (PHA)  
64 (Mino et al., 1998). These internal PHA storage pools can be used for denitrification in  
65 combination with P-removal through denitrifying polyphosphate accumulating  
66 organisms (DPAO) (Carvalho et al., 2007; de Kreuk and van Loosdrecht, 2004).  
67 **However**, (D)PAO **compete** for VFA with (denitrifying) glycogen accumulating  
68 organisms (D)GAO (Bassin et al., 2012; Oehmen et al., 2007). In contrast to (D)PAO,  
69 (D)GAO are unable to contribute to P-removal, which makes them less favorable for  
70 BNR processes.

71

72 In recent years, research regarding granulation and short-cut nitrogen removal processes  
73 **mainly** focused on partial-nitritation/anammox (Liang et al., 2015; Lotti et al., 2014;  
74 Wang et al., 2012). Only **a** few studies reported granulation and simultaneous nutrient  
75 removal on industrial wastewater (Arrojo et al., 2004; Cassidy and Belia, 2005; Liu et  
76 al., 2015; Wang et al., 2007). To **the** best of our knowledge, simultaneous BNR by the  
77 use of aerobic nitrite granules (ANG) has not been studied on industrial wastewater  
78 before.

79 **Environmental concerns, related to BNR, such as the reduction of greenhouse gas**  
80 **emissions have become more and more critical.** Nitrous oxide is an important  
81 greenhouse gas and contributes to the destruction of the stratospheric ozon layer (IPCC,  
82 2007). Previous research (Kampschreur et al., 2009, Kishida et al., 2004, Wunderlin et  
83 al., 2012) showed that the main factors stimulating the formation of nitrous oxide are:  
84 (1) nitrite accumulation, (2) low dissolved oxygen (DO) concentrations and low (3)  
85 COD/N values. According to these findings, N<sub>2</sub>O analysis should be taken into account  
86 for ANG.

87 This study, focuses on treating industrial wastewater originating from the potato  
88 industry, aims to: (1) **obtain a degree of** nitritation-denitritation for >80% in floccular  
89 sludge, (2) transform the floccular sludge into ANG, (3) analyse BNR performance of  
90 ANG, (4) **monitor N<sub>2</sub>O production and calculate emissions for ANG.**

91

## 92 **2. MATERIALS AND METHODS**

### 93 **2.1. Reactor Set-up And Operating Conditions**

94 A lab-scale sequencing batch reactor (SBR) with H/D ratio of 1.74 was operated at  
95 room temperature (18-22°C). The reactor was inoculated with sludge originating from a

96 lab-scale reactor treating similar industrial wastewater and performing conventional  
97 BNR. The SBR process was controlled by a Siemens PLC, process settings and  
98 visualization were controlled by a **specifically** programmed LabVIEW (National  
99 Instruments, Austin, Texas, USA) supervision program. Table 1 summarizes the  
100 different experimental periods.

101 **In experimental period, obtaining N-shortcut (ONS), during each cycle a total volume**  
102 **of 1L was fed to the reactor spread over 4 separate feeding steps. To obtain a profound**  
103 **nitrogen shortcut via nitrite, the typical SBR sequence during ONS started with of a pre-**  
104 **aerobic phase (15 min), to make sure the sludge was endogenous, and four consecutive**  
105 **anoxic feeding phases (30 min), each followed by an aeration step with variable**  
106 **duration (15-35 min). To end the SBR cycle, there was one additional anoxic (30 min)**  
107 **and aerobic (15-35 min) phase without feeding followed by a prolonged anoxic phase**  
108 **(90min), a settling phase (120 min) and effluent withdrawal (5 min).**

109 **During the ANG periods, starting from day 104, the SBR sequence consisted of a pre-**  
110 **aerobic phase (30 min), an extended anaerobic feeding phase (90 min), a maximum of 5**  
111 **reaction (aerobic/anoxic) steps with variable duration (see 2.2), a settling phase (60 –**  
112 **2.7 min) and effluent withdrawal (5 min). During reactor operation the volume**  
113 **exchange ratio (VER) was increased from 9% (working volume 11L) during**  
114 **experimental period ONS to 26% (working volume 13.5L) in experimental period**  
115 **“ANG – P II” (table 1).**

116

## 117 **2.2. Aeration control strategy**

118 To obtain the nitrite pathway for BNR, an aeration control strategy, based on the  
119 oxygen uptake rate (OUR) was applied. It was already demonstrated by Blackburne et

120 al., 2008 and Lemaire et al., 2008 that a sharp drop in the OUR appears as soon as  $\text{NH}_4^+$   
121 oxidation is finished. This aeration control strategy was the main criterion to end the  
122 aeration period.

123 In order to promote aerobic granulation and retain nitrogen removal via nitrite, an  
124 additional control strategy based on pH-decrease was applied. This pH-slope method  
125 assured alternation between the first aerobic and anoxic period in order to control nitrite  
126 accumulation, to maintain nitrogen removal via nitrite, and to achieve a degree  
127 simultaneous nitrification-denitrification (SND) as high as possible. **The subsequent aerobic**  
128 **phases (limited to 5) had a maximum time of 35 min and were also supervised by the**  
129 **OUR aeration control strategy.**

130 The dissolved oxygen (DO) level in the aerobic step was **controlled using an on-off**  
131 **regulation between 1.0 and 2.0 mg  $\text{O}_2$ /L. The aeration rate was kept between 1.5 and**  
132 **2.0 L air/min.** A constant SRT of 20 days was maintained by periodically removing the  
133 excess sludge from the reactor.

134

### 135 **2.3. Microbial activity measurements**

136 **A custom-build respirometer was controlled by a WAGO PLC (GmbH & Co. KG,**  
137 **Germany) and consisted of a DO probe (Hamilton, Switzerland) and air pump. It was**  
138 **supervised by a custom-build LabVIEW (National Instruments, Austin, Texas, USA)**  
139 **control program, which automatically calculated oxygen uptakes rates (OUR), whereafter**  
140 **the maximum specific oxygen uptake rates (SOUR) of AOB ( $\text{SOUR}_{\text{AOB}}$ ) and NOB**  
141 **( $\text{SOUR}_{\text{NOB}}$ ) could be determined. Biomass was collected at the end of the SBR cycle**  
142 **and aerated until endogenous respiration  $\text{SOUR}_{\text{endo}}$  was reached. Then, a pulse of nitrite**  
143 **( $\text{NaNO}_2$ ) was dosed to achieve an initial concentration of 10 mg  $\text{NO}_2^-$ -N/L. After three**

144 OUR measurements, a pulse of ammonium (NH<sub>4</sub>Cl) was dosed to achieve an initial  
145 concentration of 10 mg NH<sub>4</sub><sup>+</sup>-N/L. The DO was controlled between 4 mg O<sub>2</sub>/L and 6  
146 mg O<sub>2</sub>/L. Based on Jubany et al., 2009, following calculations were used to evaluate the  
147 specific nitrogen removal rates (SR) and NOB/AOB activity ratio:

148

$$149 \text{ SOUR}_{\text{NOB}} (\text{mg O}_2/\text{h. gVSS}) = \text{SOUR}_{\text{NOB+endo}} - \text{SOUR}_{\text{endo}}$$

$$150 \text{ SR}_{\text{NOB}} = \text{SOUR}_{\text{NOB}}/(1.14 \text{ mg O}_2/\text{mg N})$$

151

$$152 \text{ SOUR}_{\text{AOB}} (\text{mg O}_2/\text{h. gVSS}) = \text{SOUR}_{\text{AOB+NOB+endo}} - \text{SOUR}_{\text{NOB+endo}}$$

$$153 \text{ SR}_{\text{AOB}} = \text{SOUR}_{\text{AOB}}/(3.43 \text{ mg O}_2/\text{mg N})$$

154

$$155 \text{ NOB/AOB activity ratio (\%)} = (\text{SR}_{\text{NOB}}/\text{SR}_{\text{AOB}}) \times 100$$

156 This NOB/AOB activity ratio can be used in order to evaluate the nitrification-  
157 denitrification process, a low nitrification-rate (SR<sub>NOB</sub>) will result in a low activity ratio,  
158 which is a good indicator for the performance of “nitrite-route” nitrification.

159

#### 160 **2.4.qPCR Procedure For Molecular Quantification**

161 DNA extraction from sludge samples was conducted using the NaTCA method  
162 (McIlroy et al., 2009). Resulting DNA concentrations were analysed by a fluorometric  
163 quantification using an Qubit 3.0 Fluorometer (ThermoFisher Scientific, Waltham,  
164 Massachusetts). In order to determine the abundance of AOB, NOB, PAO and GAO  
165 specific target genes (table 2) were quantified through SYBR Green quantitative  
166 polymerase chain reaction (qPCR) assays. Real-time PCR was performed on a CFX96  
167 Touch (BioRad, Hercules, California, USA). Standard curves were constructed by two

168 series of 10 fold dilutions of plasmids carrying the target genes, which varied between  
169  $1.0 \times 10^3$  and  $1.0 \times 10^8$ . Only standard curves with an R square  $> 0.99$  were withhold.  
170 Furthermore, for each DNA sample, three replicates were run and all qPCR assays  
171 contained no-template control reactions. For every sample, the average target copy  
172 numbers per genome (table 2), MLVSS and specific DNA concentration, were used to  
173 calculate the amount of target cells per g biomass. This finally led to the NOB/AOB  
174 absolute quantification ratio.

175

## 176 **2.5. The Industrial Wastewater**

177 The wastewater was sampled bimonthly directly at the UASB reactor outlet of a potato  
178 processing enterprise and stored immediately at 4°C. Most of the easily degradable  
179 COD was consumed during the pre-treated anaerobic step. Therefore, in the three first  
180 operational periods (ONS, ANG I and ANG II), acetate was dosed to maintain an  
181 average COD/N ratio of 6.5. In ANG P III, part of the acetate was gradually substituted  
182 (10 – 50%) by raw wastewater (sampled at the USAB inlet, containing on average 6030  
183 mg COD/L and 5080 mg SCOD/L), also to maintain an average COD/N ratio of 6.5.  
184 The average influent parameters throughout the different operational periods (ONS and  
185 ANG) were:  $1546 \pm 389$  mg COD /L,  $1334 \pm 335$  mg SCOD/L,  $236.8 \pm 26.9$  mg NH<sub>4</sub>-  
186 N/L,  $53.8 \pm 27.0$  mg PO<sub>4</sub>-P/L.

187

## 188 **2.6. Analytical methods**

189 All samples were filtered over a glass microfiber filter (particle retention 1.2 µm).  
190 Subsequently concentrations of phosphate (Hach Lange, Germany), ammonium (Hanna  
191 Instruments, Belgium), nitrite (Hach Lange, Germany), nitrate (Hanna Instruments,

192 Belgium) and COD (Hanna Instruments, Belgium) were analysed with standard cuvette  
193 tests (Hanna Instruments, Temse, Belgium). The evolution of the sludge morphology  
194 was examined using an MOTIC (Xiamen, China) microscope. Biomass concentration  
195 and sludge volume index (SVI) measurements were conducted according to the standard  
196 methods (APHA, 1998).

197

## 198 **2.7. In-situ cycle measurements**

199 During period ONS, in-situ cycle measurements during aerobic steps were carried out to  
200 determine the nitrite accumulation rate (NAR).

201

$$202 \text{ NAR (\%)} = \frac{[\text{NO}_2^- - \text{N}]_{t_0}^{\text{in}}}{([\text{NO}_2^- - \text{N}]_{t_0}^{\text{in}} + [\text{NO}_3^- - \text{N}]_{t_0}^{\text{in}})} \times 100$$

203

204 With  $t_0$  the start of the aeration phase and  $t_n$  the end of the aeration phase.

205

206 Subsequently, during the ANG periods, in-situ cycle measurements were performed  
207 during standard reactor operation. Grab samples were taken every 10 – 30 min in order  
208 to obtain profiles of nitrite, nitrate, ammonium and phosphate.  $\text{N}_2\text{O}$  profiles were  
209 monitored in the liquid phase using a Unisense (Aarhus, Denmark) micro sensor with  
210 data logging. A two-point calibration of the micro sensor was done before each  
211 measurement. A calculation example regarding  $\text{N}_2\text{O}$  emissions can be found in the  
212 supplementary data (A).

213

214

215

216 **2.8. SND efficiency**

217 Calculation of the simultaneous nitrification-denitrification (SND) was based on the total  
218 amount of  $\text{NO}_x\text{-N}$  formed, divided by the amount of  $\text{NH}_4\text{-N}$  oxidized. A distinction was  
219 made between  $\text{SND}_1$ , during the first aeration step, and the cumulative  $\text{SND}_c$  throughout  
220 the entire reaction cycle.

221

222  $\text{SND}_1 (\%) = ([\text{NO}_x\text{-N}]_{t_0}^{\text{in}} / [\text{NH}_4\text{-N}]_{t_n}^{\text{in}}) \times 100$

223 with  $t_0$  start of aeration 1 and  $t_n$  the end of aeration 1

224

225  $\text{SND}_c (\%) = (([\text{NO}_x\text{-N}]_{t_{10}}^{t_1} + [\text{NO}_x\text{-N}]_{t_{20}}^{t_2} + [\text{NO}_x\text{-N}]_{t_{30}}^{t_3}) / ([\text{NH}_4\text{-N}]_{t_{1n}}^{t_{10}} + [\text{NH}_4\text{-N}]_{t_{2n}}^{t_{20}} + [\text{NH}_4\text{-N}]_{t_{3n}}^{t_{30}})) \times 100;$

226

227  
228 With  $t_{10}$  start of aeration 1 and  $t_{1n}$  the end of aeration ;  $t_{20}$  start of aeration 2 and  $t_{2n}$  the  
229 end of aeration 2; ....

### 230 3. RESULTS AND DISCUSSION

#### 231 3.1. Repression Of NOB And Achievement Of The Nitrite-pathway

232 The first experimental period, ONS lasted 103 days. During this period, the NAR was  
233 determined regularly. In the subsequent operational periods (ANG I – III) maintaining  
234 the nitrite-pathway was an important objective, but due to SND processes, it was  
235 unfeasible to determine the NAR during the ANG periods. Therefore, specific activity  
236 measurements, to determine  $SR_{AOB}$ ,  $SR_{NOB}$  and to calculate NOB/AOB activity ratio,  
237 were carried out throughout the whole study. On day 0 (the inoculum), 99, 174, 205,  
238 408, 427 and 455 measurements for molecular quantification by qPCR were conducted  
239 to verify the absolute populations of AOB.

240

241 Figure 1 shows the results of the NAR (during stage ONS), microbial activity  
242 measurements, molecular quantification, NOB/AOB activity/quantification ratio and  
243 nitrogen removal efficiency. It can be observed that, at the onset of ONS, nitrogen  
244 removal followed the conventional two-step nitrification. Through the use of the  
245 aeration control strategy, a continued increase in NAR and decline in the NOB/AOB  
246 activity ratio could be observed. As from day 79 the NAR reached 87.5%, NOB/AOB  
247 activity ratio went down below 30% and a quantification ratio of 0.093 was achieved.  
248 These results indicate that nitrification was shifted to a one step nitrification followed by a  
249 denitrification step to close the nitrogen removal loop. Aeration control strategies already  
250 proved their value for achieving nitrification-denitrification (Blackburne et al., 2008;  
251 Lemaire et al., 2008).

252

253 During the following operational periods (ANG PI – PIII), the nitrite-route for nitrogen  
254 removal could easily be maintained, the NOB/AOB activity ratio was below 25%  
255 **almost constantly**. Only at the start of ANG PIII, a slight increase of the NOB/AOB  
256 activity ratio to about 40% could be noticed. In addition, the NOB/AOB quantification  
257 ratio was below 0.05 **for every sample taken** during the ANG periods. **Overall, a close**  
258 **relationship between NOB/AOB activity ratio and quantification ratio was observed.**

259 .

### 260 **3.2.Factors Affecting The Transformation From Floccular Sludge Into**

#### 261 **ANG**

262 **An important aim of the present study** was to transform the floccular sludge into ANG.  
263 Therefore, sludge characteristics as biomass concentration, SVI and morphology were  
264 examined during the different operational periods. In order to survey the microbial  
265 population, molecular quantification by qPCR was conducted.

266

267 The biomass concentration of the seed sludge was 6.31 gMLSS/L. During ONS, the  
268 VER and settling time were kept constant. As a result of these settings, biomass  
269 concentrations (ML(V)SS) sharply decreased (figure 2). The initial SVI values varied  
270 between 200 and 300 mL/g.

271 The formation and maintenance of the ANG granules can be divided in 3 consecutive  
272 periods (ANG - P I; ANG - P II; ANG - P III). During ANG - P I settling time was  
273 decreased from 25 min on day 104 to 5 min on day 208, **while** the VER was increased  
274 from 11% to 20%. The combination of these two factors resulted in a strong hydraulic  
275 selection pressure, which **has been shown** to lead to the selection of quickly settling  
276 particles (Liu and Tay, 2004). As figure 2 shows, the ML(V)SS fluctuated, while **in**

277 general a slightly rising trend is visible. The sludge morphology of the seed sludge  
278 showed irregular, loose and even filamentous structures (supplementary Fig. B.1(A)).  
279 At the start of ANG – P I (day 104), some preliminary aggregation was already visible.  
280 Granule development was especially clear from day 127 up to day 205 leading to well-  
281 shaped particles with dimensions up to 0,5 mm. Another parameter demonstrating well-  
282 formed granules is the  $SVI_{10}/SVI_{30}$  ratio (de Kreuk et al., 2007). Figure 2 shows the  
283 decreasing SVI values (at day 167,  $SVI_{10} = SVI_{30} = 16$  mL/g). Due to some aeration  
284 issues a minor increase in SVI values was observed from day 174 until day 208  
285 (average  $SVI_{10} = 77$  mL/g;  $SVI_{30} = 55$ ).

286

287 At the start of ANG - P II, the industrial wastewater was contaminated with anaerobic  
288 sludge originating from the potato enterprise UASB reactor. This contamination is  
289 clearly visible in figure 2, which shows that the MLVSS/MLSS ratio decreased and  
290 indicating an increase of inorganic matter in the reactor. With a slight delay, also the  
291 SVI values increased. From the microscopic point of view (supplementary Fig. B.1(A)),  
292 granule degradation and accumulation of loose and filamentous structures were  
293 observed. At day 226, the batch of wastewater was replaced. Nevertheless, it should be  
294 emphasized that a significant portion of the previously formed granules remained stable  
295 during this disturbance (supplementary Fig. B.1(A)).

296

297 ANG - P III was initiated with a reduction of the settling time from 6 min on day 307 to  
298 a final minimum of 3 min on day 362. As figure 2 shows, this modification affected the  
299 ML(V)SS values, which dropped significantly as results of biomass washout (effluent  
300 TSS not measured). Furthermore, sludge settling greatly improved, resulting in

301 decreasing SVI values (at day 384,  $SVI_{30} = 30$ ), indicating highly granulated biomass.  
302 This was confirmed by microscopic analysis of sludge morphology (supplementary  
303 Fig.B.1(B)), demonstrating the presence of mature granules, between day 336 and 395,  
304 with sharp edges and dimensions varying between 0.5 and 1.0 mm. Because a slight  
305 drop in nitrogen removal (76% at day 405) was observed, the VER was decreased to  
306 17% from day 405. This modification also affected sludge settleability (at day 419,  
307  $SVI_{30} = 119$ ), due to the growth of suspended biomass. However, again settleability (at  
308 day 440,  $SVI_{30} = 47$ ) as well as nitrogen removal (98% at day 440) recovered fairly  
309 quickly.

310

311 Figure 3, shows the amount of target cells per g biomass for PAO and GAO. It can be  
312 observed that the inoculum already contained a fair amount of PAO , while the GAO  
313 population was much smaller. The specific reactor operation during ONS, completely  
314 turned over this balance (day 99). Subsequent, introduction of an anaerobic feeding  
315 phase in ANG PI – PIII definitely stimulated progressive growth of PAO, while the  
316 GAO population first showed a decline whereafter, starting from ANG PIII, the  
317 population recovered again. These observations are in line with the results of previous  
318 research from de Kreuk and van Loosdrecht, 2004 and Pronk et al., 2015, who stated  
319 that selection of slow growing organisms, such as PAO and GAO are one of the most  
320 important strategies to stimulate aerobic granulation. Both the rising PAO levels, and  
321 recovery of GAO, during ANG PIII, might be a consequence of the partial replacement  
322 of the acetate by raw wastewater, containing different VFA substrates (Oehmen et al.,  
323 2007).

324

325

326 During experimental period ANG PI – PIII, also phosphorus removal was examined.  
327 These results, together with the anaerobic PO<sub>4</sub>-P release are summarized in table 3. A  
328 significant increase for both parameters can be observed between ANG PI and PII.  
329 Similar phosphorus removal efficiencies **concerning** the treatment of industrial  
330 wastewater with aerobic granular sludge were already reported by(Arrojo et al., 2004;  
331 Liu et al., 2015; Yilmaz et al., 2008).

332

### 333 **3.3.The Effect Of ANG On The Sludge Loading Rate And SND**

334 High specific conversion rates are one of the major benefits of aerobic granulation (Liu  
335 and Tay, 2004). Table 4 shows the average COD (gCOD/gVSS.day) and nitrogen  
336 sludge loading rates (mgNH<sub>4</sub>-N/gVSS.day) for the different experimental periods. A  
337 progressive increase in both loading rates can be observed over the whole operational  
338 period, **starting at** 0.162 gCOD/gVSS.day during “ONS” **up to** 0.274 gCOD/gVSS.day  
339 in period ANG-P III. The enhanced nitrogen loading rate is even more remarkable,  
340 which **was increased** from 22.8 mgNH<sub>4</sub>-N/gVSS.day to 54.2 mgNH<sub>4</sub>-N/gVSS.day.

341

342 **The possibility to employ** aerobic granules for simultaneous nitrification-denitrification  
343 was already proved by Bassin et al., 2012; Kishida et al., 2006; Pronk et al., 2015.

344 Moreover, SND was also observed in the present study, the results are summarized in  
345 table 3, which **displays** the average %SND<sub>1</sub> and %SND<sub>c</sub> for the different experimental  
346 periods. These results confirm the favourable effect of granulation, and COD storage,  
347 during the anaerobic phase on the SND (Gao et al., 2011; Guo et al., 2013; Zeng et al.,  
348 2003). Moreover, it is clearly visible that after the first loop (one alternation between

349 aerobic and anoxic steps), the stored organic carbon becomes more and more depleted,  
350 leading to a decrease in %SND<sub>c</sub> in the following loops (SND<sub>c</sub>).

351

### 352 **3.4. Effect Of ANG On Nitrous Oxide Emissions**

353 Nitrous oxide can be formed during autotrophic nitrification and heterotrophic  
354 denitrification. In both processes, N<sub>2</sub>O acts as an inevitable intermediate. For ANG the  
355 two most important pathways leading to N<sub>2</sub>O production (Kampschreur et al., 2009)  
356 are: (1) nitrifier denitrification and (2) heterotrophic denitrification. Nitrifier  
357 denitrification covers the reduction from NO<sub>2</sub><sup>-</sup> by AOB under oxygen limiting  
358 conditions, low organic carbon content and increased nitrite concentrations (Wrage et  
359 al., 2001; Wunderlin et al., 2012). Incomplete heterotrophic denitrification leads to the  
360 formation of N<sub>2</sub>O as main product instead of N<sub>2</sub>. The key factors affecting this  
361 incomplete denitrification are high nitrite levels and the absence of sufficient carbon  
362 sources (Itokawa et al., 2001, Kishida et al., 2004). According to Quan et al., 2012,  
363 granules could be a trigger for enhanced N<sub>2</sub>O emissions as well.

364

365 Due to the high solubility of N<sub>2</sub>O in water, it was assumed that during non-aerated  
366 periods, the emissions were negligible. Therefore, emissions were only considered  
367 during aeration events. For experimental period ONS, N<sub>2</sub>O profiles were assessed for  
368 one specific aeration step on day 59 and 97 (figure 4 A/B). The on/off aeration  
369 regulation affected N<sub>2</sub>O formation, resulting in a specific saw - tooth profile. However,  
370 in general, a progressive increase in soluble N<sub>2</sub>O could be observed for both  
371 experiments. In order to estimate N<sub>2</sub>O emissions, calculations (supplementary data A)  
372 were performed, resulting in the cumulative N<sub>2</sub>O - N emissions, which can be compared

373 to the total amount of  $\text{NH}_4^+$  - N consumed. For both experiments, comparable results of  
374 approximately 4.0% emission as  $\text{N}_2\text{O}$  - N were obtained. As already stated before, the  
375 main trigger for these  $\text{N}_2\text{O}$  emissions is expected to be nitrifier denitrification caused by  
376 low DO and elevated  $\text{NO}_2^-$  concentrations.

377 During ANG – PIII, online  $\text{N}_2\text{O}$  measurements were conducted on day 308 and 449,  
378 next to a number of grab samples analysed offline, in order to obtain a general overview  
379 of the nitrogen mass balance (figure 5). Both types of measurements show a significant  
380 formation of  $\text{N}_2\text{O}$  during aerobic steps as well as a significant production of  $\text{N}_2\text{O}$  during  
381 anoxic periods. Due to subsequent aeration events, anoxic  $\text{N}_2\text{O}$  production caused  
382 strongly increased  $\text{N}_2\text{O}$  emissions (Kampschreur et al., 2008).

383 At day 308, the  $\text{N}_2\text{O}$  - N emission was 15.05 mg/L. According to figure 5A, nitrite was  
384 completely denitrified after the anoxic periods. However, high concentrations of  
385 dissolved nitrous oxide at the end of an anoxic period caused elevated  $\text{N}_2\text{O}$  emissions  
386 after the anoxic phases. In total 5.05 mg  $\text{N}_2\text{O}$  - N /L was emitted during the first  
387 aeration pulse after an anoxic phase, which represents 33.5% of the  $\text{N}_2\text{O}$  emitted.

388 Similar results were obtained at day 449 as 5.20 mg  $\text{N}_2\text{O}$  - N /L (31.1%) of the total of  
389 16.68 mg  $\text{N}_2\text{O}$  - N/L was emitted directly after the anoxic phase. However, the overall  
390  $\text{N}_2\text{O}$  emission at day 449 was almost twofold compared with the emission on day 308.

391 The availability of organic carbon compounds during aerobic periods might explain this  
392 difference, as the acetate was partially substituted by raw wastewater during ANG PIII.

393 As most of the easily degradable carbon is stored intracellularly during anaerobic  
394 conditions, significant  $\text{N}_2\text{O}$  production is caused by simultaneous nitrification-  
395 denitrification. Similarly, Gao et al., 2016 found that the combination of (1) internal  
396 storage denitrification and (2) higher ammonium removal rates led to nitrite

397 accumulation. They suggested that under these conditions, incomplete denitrification  
398 occurs, which might at least partially explain why the emission at day 449 is almost  
399 twofold higher than the emission on day 308. These findings correspond with the results  
400 of previous studies (Lemaire et al., 2006; Zeng et al., 2003). Consequently, strategies to  
401 mitigate N<sub>2</sub>O emissions for ANG should be investigated in more detail. Desloover et al.,  
402 2012 proposed several strategies to mitigate the N<sub>2</sub>O emissions in biological nitrogen  
403 removal systems. Most strategies rely on minimizing N<sub>2</sub>O precursors, as low DO,  
404 elevated NO<sub>2</sub><sup>-</sup> concentrations, low COD/N ratio's. Yang et al., 2013 found that applying  
405 a step-feed on a partial nitrifying granular sludge reactor could reduce the N<sub>2</sub>O  
406 emissions with 53.8%. The use of a step-feed might be a promising strategy for ANG  
407 processes as well. During period ONS, step-feed is used in order to obtain nitrogen  
408 shortcut, which coincides with the fact that N<sub>2</sub>O emissions are significant lower than  
409 during the ANG periods. In addition, it can be interesting to investigate elevated DO  
410 set points and there might be a great potential in developing a control strategy, based on  
411 the dissolved N<sub>2</sub>O accumulation in order to supervise the anoxic periods Recently,  
412 Dries, 2016 also emphasized that dynamic SBR cycles might be interesting, in order to  
413 develop novel N<sub>2</sub>O mitigation strategies.

414

#### 415 4. CONCLUSIONS

416 In conclusion, formation of ANG with pre-treated industrial wastewater, originating  
417 from the potato industry, is possible. Short-cut nitrification was achieved after  
418 approximately 80 days. After changing the operational strategy, granules became the  
419 dominant fraction from day 168. Additionally, the nitrification-denitrification pathway  
420 could be maintained at all times. BNR efficiencies for N and P reached 95.3% and

421 65.7%. Nevertheless, an important denitrification product of ANG seems to be nitrous  
422 oxide, which implies some environmental issues. **Future investigation should examine**  
423 **how to mitigate N<sub>2</sub>O formation and emission in more detail.**

424

#### 425 **ACKNOWLEDGMENTS**

426 This work was funded by the by the Flanders Innovation and Entrepreneurship Agency  
427 (grant number: 131325)

428

## REFERENCES

429

1. APHA, 1998. Standard methods for the examination of water and wastewater, 20<sup>th</sup> ed. American Public Health Association. Washington DC.

430

431

2. Arrojo, B., Mosquera-Corral, A., Garrido, J.M., Méndez, R., 2004. Aerobic granulation with industrial wastewater in sequencing batch reactors. *Water Res.* 38, 3389–3399.

432

433

434

3. Bassin, J.P., Kleerebezem, R., Dezotti, M., van Loosdrecht, M.C.M., 2012.

435

Simultaneous nitrogen and phosphate removal in aerobic granular sludge

436

reactors operated at different temperatures. *Water Res.* 46, 3805–3816.

437

4. Beun, J., Heijnen, J.J., van Loosdrecht, M.C.M., 2001. N-removal in a granular sludge sequencing batch airlift reactor. *Biotechnol. Bioeng.* 75, 82–92.

438

439

5. Blackburne, R., Yuan, Z., Keller, J., 2008. Demonstration of nitrogen removal via nitrite in a sequencing batch reactor treating domestic wastewater. *Water Res.* 42, 2166–76.

440

441

442

6. Bosak, V., VanderZaag, A., Crolla, A., Kinsley, C., Gordon, R., 2016.

443

Performance of a constructed wetland and pretreatment system receiving potato farm wash water. *Water (Switzerland)* 8, 1–14.

444

445

7. Carvalho, G., Lemos, P.C., Oehmen, A., Reis, M. a M., 2007. Denitrifying phosphorus removal: Linking the process performance with the microbial community structure. *Water Res.* 41, 4383–4396.

446

447

448

8. Cassidy, D.P., Belia, E., 2005. Nitrogen and phosphorus removal from an abattoir wastewater in a SBR with aerobic granular sludge. *Water Res.* 39, 4817–23.

449

450

451

9. Crocetti R., G., Banfield F., J.F., Keller, J., Bond, P.L., Blackall, L.L., 2002.

- 452 Glycogen-accumulating organisms in laboratory-scale and full-scale wastewater  
453 treatment processes. *Microbiology* 148, 3353–3364.
- 454 10. De Kreuk, M.K., Heijnen, J.J., Van Loosdrecht, M.C.M., 2005. Simultaneous  
455 COD, nitrogen, and phosphate removal by aerobic granular sludge. *Biotechnol.*  
456 *Bioeng.* 90, 761–769.
- 457 11. de Kreuk, M.K., Kishida, N., van Loosdrecht, M.C.M., 2007. Aerobic granular  
458 sludge – state of the art. *Water Sci. Technol.* 55, 75-81.
- 459 12. de Kreuk, M.K., van Loosdrecht, M.C.M., 2004. Selection of slow growing  
460 organisms as a means for improving aerobic granular sludge stability. *Water Sci.*  
461 *Technol.* 49, 9–17.
- 462 13. Desloover, J., Vlaeminck, S.E., Clauwaert, P., Verstraete, W., Boon, N., 2012.  
463 Strategies to mitigate N<sub>2</sub>O emissions from biological nitrogen removal systems.  
464 *Curr. Opin. Biotechnol.* 23, 474–482.
- 465 14. Dries, J., 2016. Dynamic control of nutrient-removal from industrial wastewater  
466 in a sequencing batch reactor, using common and low-cost online sensors. *Water*  
467 *Sci. Technol.* 73, 740–745.
- 468 15. Fukushima, T., Uda, N., Onuki, M., Satoh, H., Mino, T., 2007. Development of  
469 the Quantitative PCR Method for *Candidatus “Accumulibacter phosphatis”* and  
470 Its Application to Activated Sludge. *J. Water Environ. Technol.* 5, 37–43.
- 471 16. Gao, D., Yuan, X., Liang, H., Wu, W.M., 2011. Comparison of biological  
472 removal via nitrite with real-time control using aerobic granular sludge and  
473 flocculent activated sludge. *Appl. Microbiol. Biotechnol.* 89, 1645–1652.
- 474 17. Gao, M., Yang, S., Wang, M., Wang, X.H., 2016. Nitrous oxide emissions from  
475 an aerobic granular sludge system treating low-strength ammonium wastewater.

- 476 J. Biosci. Bioeng. 122, 601–605.
- 477 18. Guo, J., Zhang, L., Chen, W., Ma, F., Liu, H., Tian, Y., 2013. The regulation  
478 and control strategies of a sequencing batch reactor for simultaneous nitrification  
479 and denitrification at different temperatures. *Bioresour. Technol.* 133, 59–67.
- 480 19. Hiroki Itokawa, Hanaki, K., Matsuo, T., 2001. Nitrous Oxide Production in  
481 High-Loading Biological Nitrogen Removal Process Under Low Cod / N Ratio  
482 Condition. *Water Res.* 35, 657–664.
- 483 20. Jubany, I., Lafuente, J., Baeza, J. a, Carrera, J., 2009. Total and stable washout  
484 of nitrite oxidizing bacteria from a nitrifying continuous activated sludge system  
485 using automatic control based on Oxygen Uptake Rate measurements. *Water*  
486 *Res.* 43, 2761–72.
- 487 21. Kampschreur, M.J., Temmink, H., Kleerebezem, R., Jetten, M.S.M., van  
488 Loosdrecht, M.C.M., 2009. Nitrous oxide emission during wastewater treatment.  
489 *Water Res.* 43, 4093–103.
- 490 22. Kampschreur, M.J., van der Star, W.R.L., Wielders, H.A., Mulder, J.W., Jetten,  
491 M.S.M., van Loosdrecht, M.C.M., 2008. Dynamics of nitric oxide and nitrous  
492 oxide emission during full-scale reject water treatment. *Water Res.* 42, 812–826.
- 493 23. Kishida, N., Kim, J., Tsuneda, S., Sudo, R., 2006. Anaerobic/oxic/anoxic  
494 granular sludge process as an effective nutrient removal process utilizing  
495 denitrifying polyphosphate-accumulating organisms. *Water Res.* 40, 2303–10.
- 496 24. Kishida, N., Kim, J.H., Kimochi, Y., Nishimura, O., Sasaki, H., Sudo, R., 2004.  
497 Effect of C/N ratio on nitrous oxide emission from swine wastewater treatment  
498 process. *Water Sci Technol* 49, 359–365.
- 499 25. Kuba, T., Van Loosdrecht, M.C.M., Heijnen, J.J., 1996. Phosphorus and

- 500 nitrogen removal with minimal COD requirement by integration of denitrifying  
501 dephosphatation and nitrification in a two-sludge system. *Water Res.* 30, 1702–  
502 1710.
- 503 26. Lemaire, R., Marcelino, M., Yuan, Z., 2008. Achieving the nitrite pathway using  
504 aeration phase length control and step-feed in an SBR removing nutrients from  
505 abattoir wastewater. *Biotechnol. Bioeng.* 100, 1228–36.
- 506 27. Lemaire, R., Meyer, R., Taske, A., Crocetti, G.R., Keller, J., Yuan, Z., 2006.  
507 Identifying causes for N<sub>2</sub>O accumulation in a lab-scale sequencing batch reactor  
508 performing simultaneous nitrification, denitrification and phosphorus removal. *J.*  
509 *Biotechnol.* 122, 62–72.
- 510 28. Liang, Y., Li, D., Zeng, H., Zhang, C., Zhang, J., 2015. Rapid start-up and  
511 microbial characteristics of partial nitrification granular sludge treating domestic  
512 sewage at room temperature. *Bioresour. Technol.* 741–745.
- 513 29. Liu, Y., Kang, X., Li, X., Yuan, Y., 2015. Performance of aerobic granular  
514 sludge in a sequencing batch bioreactor for slaughterhouse wastewater  
515 treatment. *Bioresour. Technol.* 190, 487–491.
- 516 30. Liu, Y., Tay, J.-H., 2004. State of the art of biogranulation technology for  
517 wastewater treatment. *Biotechnol. Adv.* 22, 533–563.
- 518 31. Lochmatter, S., Gonzalez-Gil, G., Holliger, C., 2013. Optimized aeration  
519 strategies for nitrogen and phosphorus removal with aerobic granular sludge.  
520 *Water Res.* 47, 6187–97.
- 521 32. Lotti, T., Kleerebezem, R., Hu, Z., Kartal, B., Jetten, M.S.M., van Loosdrecht,  
522 M.C.M., 2014. Simultaneous partial nitritation and anammox at low temperature  
523 with granular sludge. *Water Res.* 66, 111–21.

- 524 33. McIlroy, S.J., Porter, K., Seviour, R.J., Tillett, D., 2009. Extracting nucleic acids  
525 from activated sludge which reflect community population diversity. *Antonie*  
526 *van Leeuwenhoek*, Int. J. Gen. Mol. Microbiol. 96, 593–605.
- 527 34. Mino, T., Van Loosdrecht, M.C.M., Heijnen, J.J., 1998. Microbiology and  
528 biochemistry of the enhanced biological phosphate removal process. *Water Res.*  
529 32, 3193–3207.
- 530 35. Oehmen, A., Lemos, P.C., Carvalho, G., Yuan, Z., Keller, J., Blackall, L.L.,  
531 Reis, M. a M., 2007. Advances in enhanced biological phosphorus removal:  
532 from micro to macro scale. *Water Res.* 41, 2271–300.
- 533 36. Peng, Y., Zhu, G., 2006. Biological nitrogen removal with nitrification and  
534 denitrification via nitrite pathway. *Appl. Microbiol. Biotechnol.* 73, 15–26.
- 535 37. Pester, M., Maixner, F., Berry, D., Rattei, T., Koch, H., L??cker, S., Nowka, B.,  
536 Richter, A., Spieck, E., Lebedeva, E., Loy, A., Wagner, M., Daims, H., 2014.  
537 NxrB encoding the beta subunit of nitrite oxidoreductase as functional and  
538 phylogenetic marker for nitrite-oxidizing Nitrospira. *Environ. Microbiol.* 16,  
539 3055–3071.
- 540 38. Pronk, M., de Kreuk, M.K., de Bruin, B., Kamminga, P., Kleerebezem, R., van  
541 Loosdrecht, M.C.M., 2015. Full scale performance of the aerobic granular  
542 sludge process for sewage treatment. *Water Res.* 84, 207–217.
- 543 39. Quan, X., Zhang, M., Lawlor, P.G., Yang, Z., Zhan, X., 2012. Nitrous oxide  
544 emission and nutrient removal in aerobic granular sludge sequencing batch  
545 reactors. *Water Res.* 46, 4981–4990.
- 546 40. Rosenwinkel, K.H., Verstraete, W., Vlaeminck, S.E., Wagner, M., Kipp, S.,  
547 Manig, N., 2014. Industrial wastewater treatment, in: Jenkins, D., Wanner, J.,

- 548 Activated Sludge 100 years and counting., Londen, pp. 343-367.
- 549 41. Rotthauwe, J., Witzel, K., 1997. The Ammonia Monooxygenase Structural Gene  
550 amoA as a Functional Marker : Molecular Fine-Scale Analysis of Natural  
551 Ammonia-Oxidizing Populations. *Applied and Environmental Microbiology* 63,  
552 4704–4712.
- 553 42. Third, K. a, Burnett, N., Cord-Ruwisch, R., 2003. Simultaneous nitrification and  
554 denitrification using stored substrate (PHB) as the electron donor in an SBR.  
555 *Biotechnol. Bioeng.* 83, 706–20.
- 556 43. Wang, S.-G., Liu, X.-W., Gong, W.-X., Gao, B.-Y., Zhang, D.-H., Yu, H.-Q.,  
557 2007. Aerobic granulation with brewery wastewater in a sequencing batch  
558 reactor. *Bioresour. Technol.* 98, 2142–2147.
- 559 44. Wang, X.H., Jiang, L.X., Shi, Y.J., Gao, M.M., Yang, S., Wang, S.G., 2012.  
560 Effects of step-feed on granulation processes and nitrogen removal  
561 performances of partial nitrifying granules. *Bioresour. Technol.* 123, 375–381.
- 562 45. Wrage, N., Velthof, G.L., Van Beusichem, M.L., Oenema, O., 2001. Role of  
563 nitrifier denitrification in the production of nitrous oxide. *Soil Biol. Biochem.*  
564 33, 1723–1732.
- 565 46. Wunderlin, P., Mohn, J., Joss, A., Emmenegger, L., Siegrist, H., 2012.  
566 Mechanisms of N<sub>2</sub>O production in biological wastewater treatment under  
567 nitrifying and denitrifying conditions. *Water Res.* 46, 1027–37.
- 568 47. Yang, S., Gao, M.M., Liang, S., Wang, S.G., Wang, X.H., 2013. Effects of step-  
569 feed on long-term performances and N<sub>2</sub>O emissions of partial nitrifying  
570 granules. *Bioresour. Technol.* 143, 682–685.
- 571 48. Yilmaz, G., Lemaire, R., Keller, J., Yuan, Z., 2008. Simultaneous nitrification,

572           denitrification, and phosphorus removal from nutrient-rich industrial wastewater  
573           using granular sludge. *Biotechnol. Bioeng.* 100, 529–541.

574   49. Zanetti, L., Frison, N., Nota, E., Tomizioli, M., Bolzonella, D., Fatone, F., 2012.  
575           Progress in real-time control applied to biological nitrogen removal from  
576           wastewater. A short-review. *Desalination* 286, 1–7.

577   50. Zeng, R.J., Lemaire, R., Yuan, Z., Keller, J., 2003. Simultaneous nitrification,  
578           denitrification, and phosphorus removal in a lab-scale sequencing batch reactor.  
579           *Biotechnol. Bioeng.* 84, 170–178.

580

<b>Day</b>	<b>Period</b>	<b>VER (%)</b>	<b>Aim</b>
1 - 103	obtaining N-shortcut (ONS)	9%	Repressing NOB; promoting AOB
104 - 208	ANG period I (ANG P I)	9% - 17%	Promoting granulation
209 - 306	ANG period II (ANG P II)	17% - 26%	Examination of granule stability
307 - 456	ANG period II (ANG P III)	26% - 17%	Recovering and maintaining ANG

581 **Table 1: Overview different experimental periods.**

582

<b>Target</b>	<b>Cell copy number</b>	<b>Primers</b>	<b>Conditions</b>	<b>Reference</b>
<b>amoA</b> <i>Nitrosomonas</i>	2	amoA-1F / amoA-1R	98° - 3' [98° - 15'' / 56° - 15'' / 72° - 30''] x 40	Rotthauwe and Witzel, 1997
<b>nxB</b> <i>Nitrospira</i>	2	nxBF169 / nxBR638	98° - 3' [98° - 15'' / 56° - 15'' / 72° - 30''] x 40	Pester et al., 2014
<b>16S rRNA</b> <i>Candidatus</i> <i>Accumulibacter</i> <i>phosphatis</i>	2	PAO651f / PAO846r	98° - 3' [98° - 15'' / 57° - 15'' / 72° - 15''] x 40	Fukushima et al., 2007
<b>16S rRNA</b> <i>Candidatus</i> <i>Competibacter</i> <i>phosphatis</i>	5,66	GAOQ989f / GAM1278r	98° - 3' [98° - 15'' / 59° - 15'' / 72° - 30''] x 40	Crocetti R. et al., 2002

583

584 Table 2: Target, cell copy number, primers, qPCR conditions and references

585

	<b>PO<sub>4</sub>-P removal (%)</b>	<b>Anaerobic PO<sub>4</sub>-P release (mg PO<sub>4</sub>-P/g.VSS)</b>
	Average ± SD	Average ± SD
<b>ANG P I (day 104 – 208)</b>	41.0 ± 15.1% (n=15)	9.2 ± 9.5 (n=4)
<b>ANG P II (day 209 – 306)</b>	65.7 ± 18.1% (n=14)	21.1 ± 2.3 (n=3)
<b>ANG P III (day 307 – 456)</b>	61.5 ± 27.1% (n=21)	20.6 ± 6.4 (n=12)

586 **Table 3: Average PO<sub>4</sub>-P removal (%) and anaerobic PO<sub>4</sub>-P release (mg PO<sub>4</sub>-P/g.VSS) for the**  
587 **different experimental periods.**

588

	COD loading rate (g COD/g.VSS.day)				N loading rate (mg NH <sub>4</sub> -N/g.VSS.day)			
	<i>Min.</i>	<i>Max.</i>	<i>Average + SD</i>	<i>%CV</i>	<i>Min.</i>	<i>Max.</i>	<i>Average + SD</i>	<i>%CV</i>
<b>ONS (day 1 - 103)</b>	0.0539	0.269	0.162 ± 0.0455	28	6.76	44.6	22.8 ± 8.46	37
<b>ANG P I (day 104 - 208)</b>	0.0918	0.291	0.195 ± 0.0456	23	16.3	45.5	29.6 ± 7.78	26
<b>ANG P II (day 209 - 306)</b>	0.109	0.303	0.203 ± 0.0463	23	20.0	56.4	40.9 ± 11.2	27
<b>ANG P III (day 307 - )</b>	0.130	0.444	0.274 ± 0.0677	25	34.2	81.1	54.2 ± 9.40	17

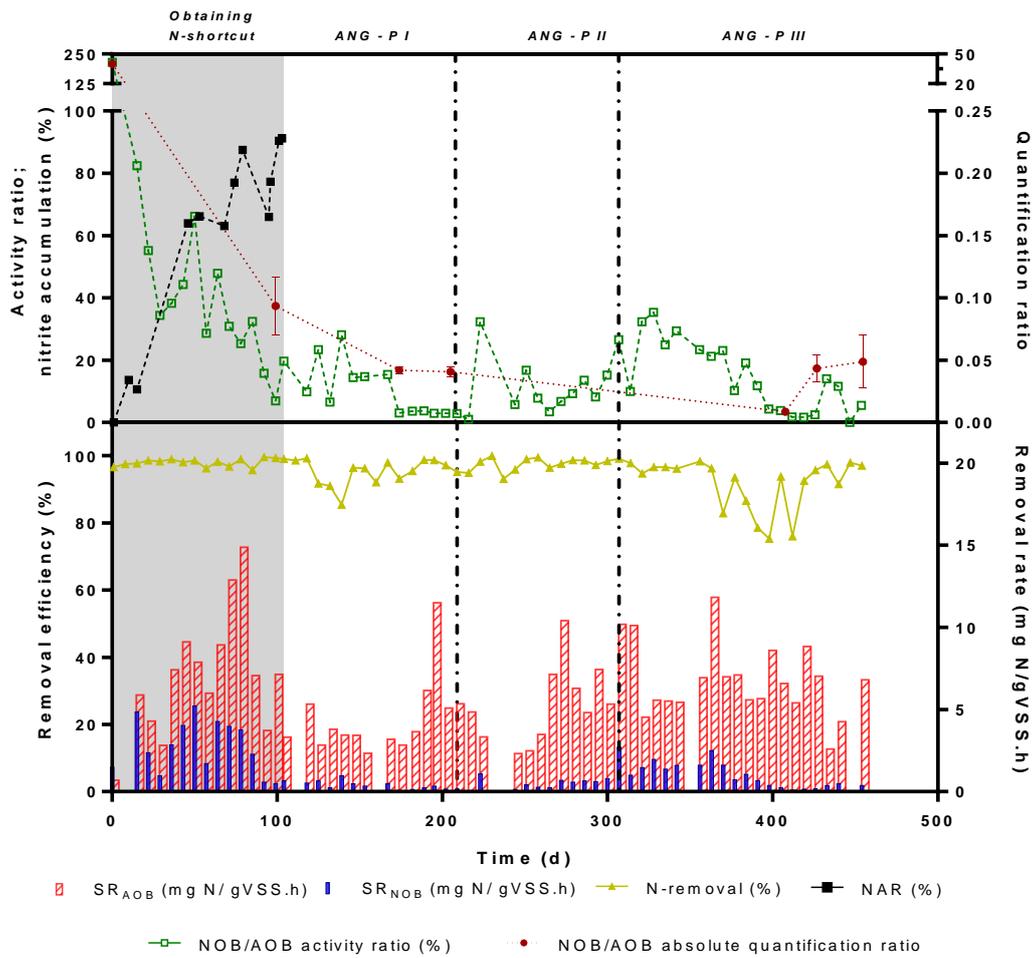
589 Table 4: Average COD and nitrogen loading rate during the different experimental periods.

590

	<b>SND<sub>1</sub> (%)</b>				<b>SND<sub>c</sub> (%)</b>			
	<i>Min.</i>	<i>Max.</i>	<i>Average + SD</i>	<i>%CV</i>	<i>Min.</i>	<i>Max.</i>	<i>Average + SD</i>	<i>%CV</i>
<b>ONS (day 1 - 103)</b>	39.2	85.2	64.9 ± 14.5	22				
<b>ANG P I (day 104-208)</b>	45.0	87.0	72.4 ± 15.2	21	37.9	71.2	49.0 ± 14.3	29
<b>ANG P II (day 209-306)</b>	65.3	85.5	75.3 ± 10.1	13	44.0	66.1	55.1 ± 15.6	28
<b>ANG P II (day 307 - )</b>	48.9	88.6	74.0 ± 12.4	17	35.9	66.2	50.0 ± 9.3	19

591 **Table 5: Average SND<sub>1</sub> and SND<sub>c</sub> during the different experimental periods.**

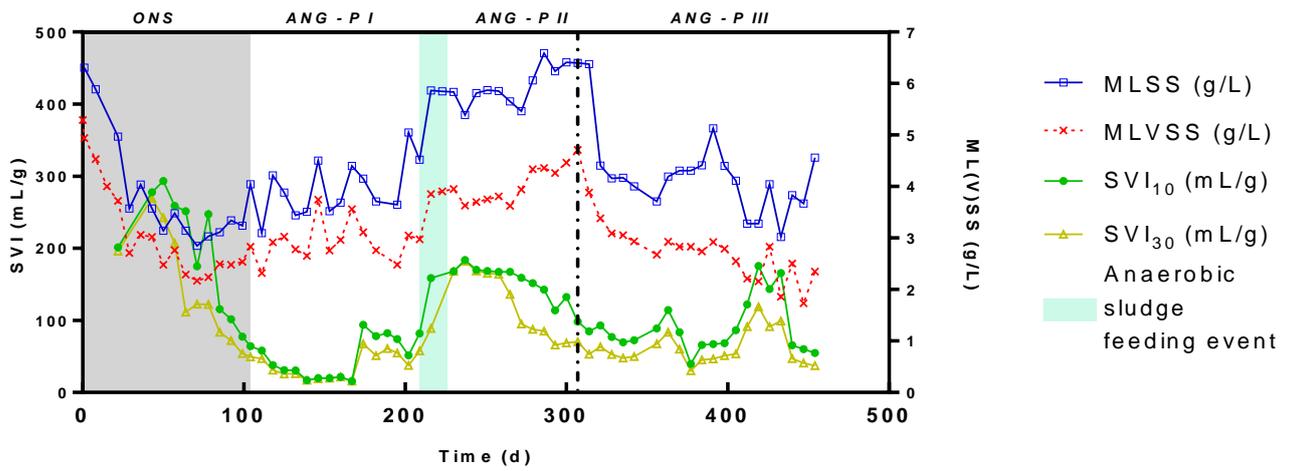
592



593

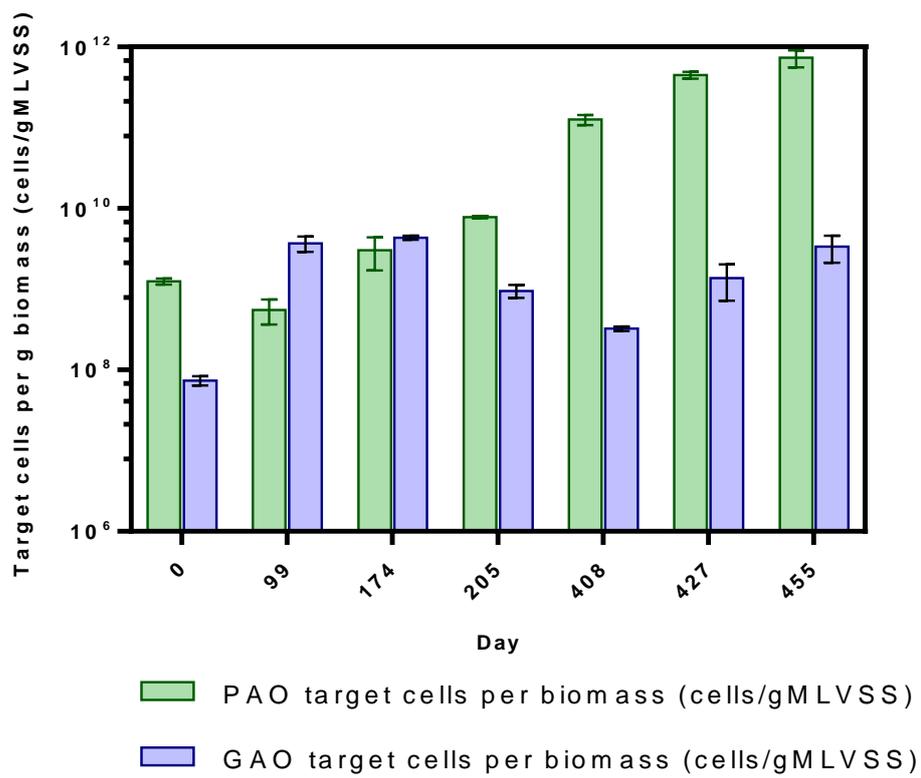
594 **Figure 1. Nitrite accumulation rate (%), Specific nitrogen removal rates (SR) for AOB and**  
 595 **NOB (mg N/gMLVSS.h), NOB/AOB activity ratio (%), NOB/AOB absolute quantification**  
 596 **ratio and nitrogen removal (%) throughout the different experimental periods.**

597



598  
 599 **Figure 2. Evolution of sludge parameters ML(V)SS, SVI<sub>10</sub> and SVI<sub>30</sub> throughout the**  
 600 **different operational periods (ONS – ANG P III)**

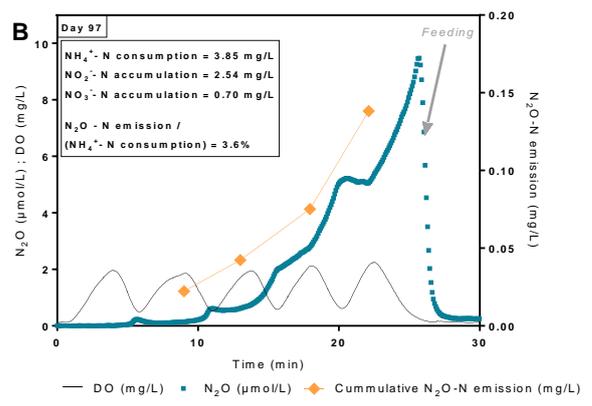
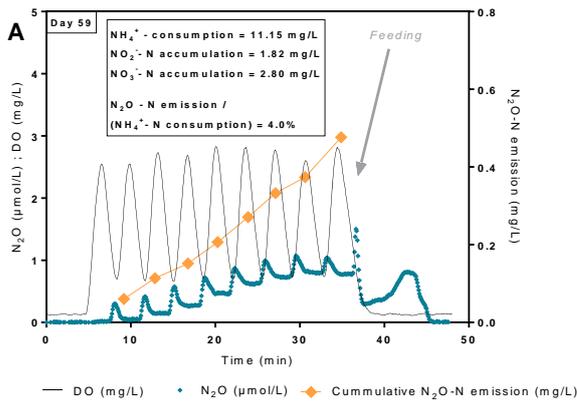
601



602

603 **Figure 3. Evolution of specific PAO and GAO abundance (target cells per biomass) during**  
 604 **the operational period.**

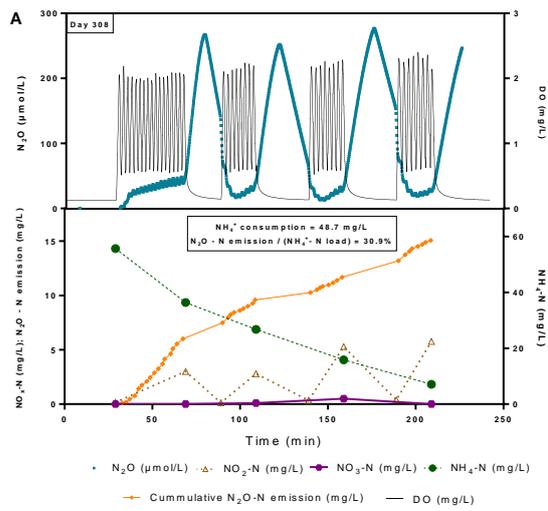
605



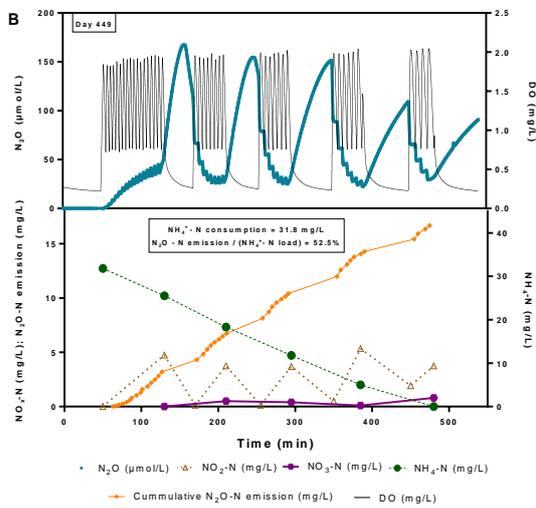
606  
607

**Figure 4. Nitrous oxide formation and emission on day 59 (A) and 97 (B).**

608



609



610

611 **Figure 5. Online cycle measurements showing nitrous oxide formation and emission in**  
 612 **combination with  $\text{NO}_2^-$ ,  $\text{NO}_3^-$  and  $\text{NH}_4^+$  offline grab samples on day 308 (A) and 445 (B).**

613

614