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Analytical performance of a platform for point-of-care CRP testing in adults consulting for lower respiratory tract infection in primary care

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1 Analytical performance of a platform for point-of-care CRP testing in adults consulting for  
2 lower respiratory tract infection in primary care

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22 **Abstract**

23 Background: C-reactive protein (CRP) is a biomarker widely used for disease severity  
24 assessment and treatment of inflammatory conditions. Point-of-care testing (POCT) devices  
25 should ideally be rapid and provide similar results to standard tests done in laboratories.

26 Methods: 2922 serum samples were obtained from adult patients presenting to primary care  
27 with symptoms of lower respiratory infection in a European diagnostic study. The analytic  
28 performance of the CRP QuikRead POCT device (Orion Diagnostica) was evaluated by  
29 comparing results with a central laboratory method (Dimension Vista, Siemens), with both  
30 tests performed in a laboratory setting.

31 Results: For a CRP cut-off concentration of  $\geq 30$  mg/L, the QuikRead test had a sensitivity of  
32 92.2%, and specificity of 99.4%. The mean difference between QuikRead and the central lab  
33 test was 0.4 mg/L. The slope of the Passing-Bablok regression was 0.94 (95% CI 0.93-0.95)  
34 indicating an underestimation of CRP levels of 6% by QuikRead.

35 Conclusions: CRP estimates obtained from the QuikRead test correlate well with a central  
36 laboratory assay and the measurement displays low inter-assay variation. Therefore, the  
37 QuikRead test is a good candidate for CRP testing in primary care.

38

39 Keywords: C-reactive protein; point-of-care test; rapid test; bedside test; respiratory infection

## 40 **Introduction**

41 C-reactive protein (CRP) is the first described acute-phase protein and is now a universal  
42 biomarker and early indicator of infectious or inflammatory conditions, related to diversified  
43 diseases, disorders and pathological conditions [1]. Measurement of CRP levels in blood are  
44 routinely performed in central hospital laboratories and many automated methods are available.  
45 However, near-patient or point-of-care testing (POCT) to measure CRP levels may be helpful  
46 in diagnosis and early management, for example to avoid unnecessary hospital referrals for  
47 acutely ill children [2] and to guide antibiotic treatment for lower respiratory tract infections  
48 [3,4]. Several POCT CRP test platforms have been developed in recent years with the  
49 advantage of providing a result within minutes. In order for these devices to be used in daily  
50 practice they should provide results comparable to laboratory based platforms for a wide  
51 variety of patients with varying CRP levels, and indications.

52 In a European diagnostic study on lower respiratory tract infections in primary care, we have  
53 previously demonstrated the added value of CRP levels in predicting pneumonia [5] which  
54 can be useful to start appropriate treatment and advice and avoid unnecessary use of antibiotics.  
55 In the current study we evaluate the analytical performance of a POCT device (QuikRead,  
56 Orion Diagnostica) for CRP measurement in serum samples of the above mentioned trial by  
57 comparing results with those obtained of our central laboratory method (Dimension Vista,  
58 Siemens), with assays on both platforms done in a laboratory.

59

## 60 **Materials and methods**

61 Serum samples from adults ( $\geq 18$  years of age) with symptoms of lower respiratory tract  
62 infection (acute cough,  $\leq 28$  days) who presented to primary care were obtained in a  
63 prospective observational study as part of GRACE-09 (Genomics to Combat Resistance

64 against Antibiotics in Community-Acquired Lower Respiratory Tract Infection in Europe;  
65 [www.grace-lrti.org](http://www.grace-lrti.org)) which was conducted from October 2007 to April 2010. A total of 3104  
66 patients were included at 16 primary care networks in 12 European countries. The mean age of  
67 patients was 49.8 years (range 18 – 92 years), 40% were male and 4.5% were diagnosed with  
68 pneumonia [5,6]. Six percent of blood test data were lacking which resulted in 2922 remaining  
69 individual patient results. Venous blood was drawn at the primary care facility, serum was  
70 prepared at local laboratories and stored at -70°C. After transport to the central laboratory of  
71 the University Hospital of Antwerp, serum samples were analyzed in batch by both the central  
72 laboratory test and the POCT method on the same sample.

73 Dimension Vista® (Siemens) was the central laboratory platform used. This is a high-  
74 throughput analyzer using conventional particle enhanced nephelometry to determine CRP  
75 levels. The lower limit of detection of this method is 2.9 mg/L. The upper limit of detection is  
76 290.0 mg/L. If this limit is exceeded, samples are diluted 2-fold and reanalyzed. The results  
77 are evaluated by comparison to a low-level (CRP range: 4.4 – 7.0 mg/L) and high-level (CRP  
78 range: 40.7 – 54.8 mg/L) standard (Liquichek Immunology Control, Bio-Rad).

79 The QuikRead 101 (Orion Corporation, Orion Diagnostica, Espoo, Finland) is a POCT or  
80 bedside immunoturbidimetric assay based on micro particles coated with anti-human CRP. It  
81 requires 20 µL of whole blood, plasma or serum and has an analytical measurement range of  
82 8-160 mg/L. Values exceeding the upper detection limit are diluted and reanalyzed. A one-  
83 level CRP control (CRP range: 42-62 mg/L) was analyzed on every analysis day.

84 Analyses were all performed by trained lab technicians who were blinded to the results of the  
85 reference test and vice versa. Sensitivity, specificity, positive (PPV) and negative predictive  
86 values (NPV) were calculated by comparing the measurements obtained by the POCT CRP  
87 with the results from the central laboratory CRP. The following CRP cut-off values were used:

88 10 mg/L as the universally applied cut-off value to indicate production of CRP, 30 mg/L  
89 previously shown to add most diagnostic value to a “symptoms and signs” model to predict  
90 pneumonia [5] or the presence of bacterial pathogens in the airways [7] and 100 mg/L which  
91 is a critical level indicating severe infection and justifying antibiotic prescription [8].

92 Considering only samples with numeric values (excluding <2.9 mg/l and <8 mg/L for the  
93 central lab and POCT test, respectively), the agreement of measurements between both  
94 methods was evaluated using Passing-Bablok regression analysis, Bland-Altman plot and  
95 Spearman for the correlation coefficient. Criteria for acceptable correlation were defined as  
96 follows: the 95% confidence interval (CI) of the slope includes 1.0, the 95% CI of the intercept  
97 includes 0.0 and the correlation coefficient > 0.95. Statistical analyses were performed with  
98 MedCalc Statistical Software version 17.5.5 (MedCalc Software bvba, Ostend, Belgium).

99

## 100 **Results**

101 The inter-assay variation of both the POCT and central lab CRP test was determined. With the  
102 QuikRead test, a variation of 3.4% was obtained, while variations of 5.9% (low level) and 5.2%  
103 (high level) were observed for the central lab test.

104 Considering positive CRP concentrations to be  $\geq 10$  mg/L, the CRP bedside test has a  
105 sensitivity of 75.5% and a specificity of 98.8% when compared with the central lab CRP test  
106 (Table 1). However, considering CRP concentrations  $\geq 30$  mg/L, the sensitivity of the CRP  
107 bedside test increases to 92.2% and the specificity to 99.4% when compared with the central  
108 laboratory CRP test, with a PPV of 96.5 % and an NPV of 98.6% (Table 2).The performance  
109 of the QuikRead test increases even further at a CRP cut-off level of  $\geq 100$  mg/l (sensitivity =  
110 92.6% and specificity = 99.8%) (Table 3).

111 The median and mean CRP values as well as the CRP range obtained using the QuikRead  
112 POCT CRP assay on the 834 serum samples with CRP concentration  $\geq 10$  mg/L are similar to  
113 CRP levels measured by the central laboratory assay: 32 mg/l, 50 mg/L (SD 51.2 mg/L) and  
114 12 – 393 mg/L respectively for the QuikRead assay and 32.5 mg/L, 49.6 mg/L (SD 48.2 mg/L)  
115 and 10.1 – 392.1 mg/L for the central laboratory method.

116 Taking into account only the samples with a numeric value (excluding  $<2.9$  mg/L samples for  
117 the central lab test and  $<8$  mg/L samples for the QuikRead test) for the correlation analysis,  
118 854 values remained. Passing-Bablok regression analysis shows a small constant bias  
119 (indicated by the intercept of 1.5) and a proportional bias (indicated by the slope of the  
120 regression line of 0.94) indicating that the QuikRead underestimates the CRP values with 6%  
121 compared to those of the central lab analysis (Fig. 1). The Spearman correlation coefficient is  
122 0.976 but a significant deviation from linearity was shown. The Bland-Altman plot shows a  
123 mean difference between both methods of 0.4 mg/L (95% limits of agreement:  $-18.8$  to  $19.5$   
124 mg/L) which is not statistically significant (Fig. 2). The plot also shows that the reliability of  
125 the POCT CRP test decreases with increased CRP values.

126

## 127 **Discussion**

128 In this study, the analytical performance of the commercially available POCT CRP device  
129 QuikRead was evaluated by comparing results obtained from this platform in a laboratory (and  
130 not at the point of care) to results from a central laboratory CRP analyzer. A CRP cut-off value  
131 of 30 mg/L has previously been shown to add most diagnostic value in predicting pneumonia  
132 [5] or the presence of a bacterial infection [7] in this patient population. Taking into account  
133 this CRP cut-off, QuikRead shows good sensitivity and excellent specificity. Analysis of the  
134 correlation between both methods, demonstrates both a small constant bias and a proportional

135 bias. The deviation at the intercept has a relatively small impact and is less relevant than the  
136 slope of the linear regression line, which deviates with 6%. An underestimation of CRP levels  
137 by the QuikRead test compared to central lab tests is in accordance with previous studies (17%  
138 compared to the Tina-quant CRP Hitachi 912, Roche, n = 59 [9]; 15% compared to Synchron®,  
139 Beckman Coulter, n = 100 [10]). However, others report good concordance (Cobas Integra,  
140 Roche, n = 231 [11]; Architect c8000, Abbott, n = 250 [12]) or even an overestimation of CRP  
141 levels (7% compared to the BN II System, Dade Behring, n = 72 [13]) by the QuikRead test. It  
142 is clear that the large sample size in our study is an asset compared to the previously published  
143 reports.

144 Compared to other CRP POCT devices, the QuikRead has a more complicated pre-analytical  
145 handling of samples which takes 2.5 min [10] and is not the most user friendly apparatus [14]  
146 making it more susceptible to flaws in the procedure. This could not be tested in our study  
147 performing the analyses in a laboratory setting, which we preferred due to practical issues when  
148 testing such a large number of samples, with qualified lab technicians. The robustness of the  
149 performance of the QuikRead POCT device should therefore be evaluated using results  
150 obtained from use by different clinicians and practice staff in routine primary care. Moreover,  
151 the performance of the POCT device in that setting might also be influenced by the use of  
152 capillary whole blood, a sampling method prone to variation, instead of the serum from venous  
153 blood used in our study. However, a previous study showed no difference in CRP determination  
154 in the venous and capillary blood samples using the QuikRead device [15].

155 All tested CRP POCT devices in the multi-device study performed well in general with  
156 correlation coefficients  $> 0.95$  [10]. QuikRead was shown to have low within- and between-  
157 day variation (5.7% and 6.3% respectively at a CRP concentration of 100 mg/L) compared to  
158 the other analyzers [14]. In our hands, the between-day variation was even lower, most  
159 probably due to the execution by trained lab technicians which does not reflect the real POCT



160 environment. Furthermore, a rather short total analysis time (without pre-analytical time) of 2  
161 min [10] to 3 min 20 sec [14] was described. Another advantage of the POCT device, besides  
162 the rapid availability of the result, is the requirement of only small volumes of blood which can  
163 be very advantageous for use in small children, a patient group not studied here.

164 POCT CRP use in primary care may increase even further in the future as obtaining chest  
165 radiographs in all patients is not feasible. However, there is ongoing debate about the clinical  
166 utility of CRP POCTs. Severe pneumonias are likely to be evident clinically, so the value of  
167 POCT CRP tests should be where there is more doubt, in helping clinicians diagnose milder  
168 pneumonias – which are more difficult to detect clinically. There should also be caution about  
169 the potential for the inappropriate and inefficient use of CRP as it is introduced in practice [16],  
170 given wider concerns about the danger of ‘medicalising’ largely self-limiting illness. CRP test  
171 results should also always be interpreted together with clinical findings, as it was shown  
172 previously that CRP levels can be low in people with pneumonia [5].

173 To the best of our knowledge, this is the first study evaluating the analytical performance of  
174 the QuikRead POCT CRP test in this high number of patients with symptoms of respiratory  
175 tract infection. The data presented here are part of a large multicenter observational study and  
176 indicate the feasibility of the QuikRead POCT CRP test in primary care. However, large  
177 prospective trials in primary care or at the point of care are needed to determine the effect of  
178 CRP POCT on clinical decision-making.

179

## 180 **Compliance with ethical standards**

181 **Funding** The study was part of the European Union FP6 funded Network of Excellence  
182 GRACE. Orion Diagnostics provided the QuikRead instruments and kits for this study. The

183 study sponsors played no role in the study design; in the collection, analysis, and interpretation  
184 of data; in the writing of the report; or in the decision to submit the paper for publication.

185 **Ethical approval** All procedures performed in studies involving human participants were in  
186 accordance with the ethical standards of the institutional and/or national research committee  
187 and with the 1964 Helsinki declaration and its later amendments or comparable ethical  
188 standards.

189 **Informed consent** Informed consent was obtained from all individual participants included in  
190 the study.

191 **Conflict of interest** The authors declare that they have no conflict of interest.

192

## 193 **References**

194 1. Vashist SK, Venkatesh AG, Marion Schneider E, Beaudoin C, Luppia PB, Luong JH (2016)  
195 Bioanalytical advances in assays for C-reactive protein. *Biotechnol Adv* 34 (3):272-290.  
196 doi:10.1016/j.biotechadv.2015.12.010

197 2. Verbakel JY, Lemiengre MB, De Burghgraeve T, De Sutter A, Aertgeerts B, Shinkins B,  
198 Perera R, Mant D, Van den Bruel A, Buntinx F (2016) Should all acutely ill children in primary  
199 care be tested with point-of-care CRP: a cluster randomised trial. *BMC Med* 14 (1):016-0679.  
200 doi:10.1186/s12916-2

201 3. Cals JW, Butler CC, Hopstaken RM, Hood K, Dinant GJ (2009) Effect of point of care  
202 testing for C reactive protein and training in communication skills on antibiotic use in lower  
203 respiratory tract infections: cluster randomised trial. *Bmj* 5 (338). doi:10.1136/bmj.b1374

204 4. Little P, Stuart B, Francis N, Douglas E, Tonkin-Crine S, Anthierens S, Cals JW, Melbye H,  
205 Santer M, Moore M, Coenen S, Butler C, Hood K, Kelly M, Godycki-Cwirko M, Mierzecki A,  
206 Torres A, Llor C, Davies M, Mullee M, O'Reilly G, van der Velden A, Geraghty AW, Goossens

207 H, Verheij T, Yardley L (2013) Effects of internet-based training on antibiotic prescribing rates  
208 for acute respiratory-tract infections: a multinational, cluster, randomised, factorial, controlled  
209 trial. *Lancet* 382 (9899):1175-1182. doi:10.1016/S0140-6736(13)60994-0

210 5. van Vugt SF, Broekhuizen BD, Lammens C, Zuithoff NP, de Jong PA, Coenen S, Ieven M,  
211 Butler CC, Goossens H, Little P, Verheij TJ (2013) Use of serum C reactive protein and  
212 procalcitonin concentrations in addition to symptoms and signs to predict pneumonia in  
213 patients presenting to primary care with acute cough: diagnostic study. *Bmj* 30 (346).  
214 doi:10.1136/bmj.f2450

215 6. Ieven M, Coenen S, Loens K, Lammens C, Coenjaerts F, Vanderstraeten A, Henriques-  
216 Normark B, Crook D, Huygen K, Butler CC, Verheij TJ, Little P, Zlateva K, van Loon A, Claas  
217 EC, Goossens H (2018) Aetiology of Lower Respiratory Tract Infection in Adults in Primary  
218 Care: A prospective Study in 11 European Countries. *Clin Microbiol Infect* 12 (18):30152-  
219 30156. doi:10.1016/j.cmi.2018.02.004

220 7. Teepe J, Broekhuizen BD, Loens K, Lammens C, Ieven M, Goossens H, Little P, Butler CC,  
221 Coenen S, Godycki-Cwirko M, Verheij TJ (2016) Predicting the presence of bacterial  
222 pathogens in the airways of primary care patients with acute cough. *CMAJ* 24:151364.  
223 doi:10.1503/cmaj.151364

224 8. Cals JW, Schot MJ, de Jong SA, Dinant GJ, Hopstaken RM (2010) Point-of-care C-reactive  
225 protein testing and antibiotic prescribing for respiratory tract infections: a randomized  
226 controlled trial. *Ann Fam Med* 8 (2):124-133. doi:10.1370/afm.1090

227 9. Monteny M, ten Brinke MH, van Brakel J, de Rijke YB, Berger MY (2006) Point-of-care  
228 C-reactive protein testing in febrile children in general practice. *Clin Chem Lab Med* 44  
229 (12):1428-1432. doi:10.1515/CCLM.2006.270

230 10. Brouwer N, van Pelt J (2015) Validation and evaluation of eight commercially available  
231 point of care CRP methods. *Clin Chim Acta* 439:195-201. doi:10.1016/j.cca.2014.10.028

- 232 11. Esposito S, Tremolati E, Begliatti E, Bosis S, Gualtieri L, Principi N (2005) Evaluation of  
233 a rapid bedside test for the quantitative determination of C-reactive protein. *Clin Chem Lab*  
234 *Med* 43 (4):438-440. doi:10.1515/CCLM.2005.077
- 235 12. Hernandez-Bou S, Trenchs V, Vanegas MI, Valls AF, Luaces C (2017) Evaluation of the  
236 bedside Quikread go(R) CRP test in the management of febrile infants at the emergency  
237 department. *Eur J Clin Microbiol Infect Dis* 36 (7):1205-1211. doi:10.1007/s10096-017-2910-  
238 2
- 239 13. Zecca E, Barone G, Corsello M, Romagnoli C, Tiberi E, Tirone C, Vento G (2009)  
240 Reliability of two different bedside assays for C-reactive protein in newborn infants. *Clin Chem*  
241 *Lab Med* 47 (9):1081-1084. doi:10.1515/CCLM.2009.246
- 242 14. Minnaard MC, van de Pol AC, Broekhuizen BD, Verheij TJ, Hopstaken RM, van Delft S,  
243 Kooijman-Buiting AM, de Groot JA, De Wit NJ (2013) Analytical performance, agreement  
244 and user-friendliness of five C-reactive protein point-of-care tests. *Scand J Clin Lab Invest* 73  
245 (8):627-634. doi:10.3109/00365513.2013.841985
- 246 15. Papaevangelou V, Papassotiriou I, Sakou I, Ferentinos G, Liapi G, Kyrka A,  
247 Konstantopoulos A (2006) Evaluation of a quick test for C-reactive protein in a pediatric  
248 emergency department. *Scand J Clin Lab Invest* 66 (8):717-721.  
249 doi:10.1080/00365510600977869
- 250 16. Minnaard MC, van de Pol AC, Hopstaken RM, van Delft S, Broekhuizen BD, Verheij TJ,  
251 de Wit NJ (2016) C-reactive protein point-of-care testing and associated antibiotic prescribing.  
252 *Fam Pract* 33 (4):408-413. doi:10.1093/fampra/cmw039

253

254 **Table legends**

255 **Table 1:** Number of serum samples found positive by the QuikRead CRP assay or the central  
256 lab assay using a cut-off of  $\geq 10$  mg/L (n = 2922)

257 **Table 2:** Number of serum samples found positive by the QuikRead CRP assay or the central  
258 lab assay using a cut-off of  $\geq 30$  mg/L [5,7] (n = 2922)

259 **Table 3:** Number of serum samples found positive by the QuikRead CRP assay or the central  
260 lab assay using a cut-off of  $\geq 100$  mg/L (n = 2922)

261 **Figure legends**

262 **Fig. 1:** Comparison of QuikRead CRP test with central laboratory CRP values using Passing-  
263 Bablok regression analysis (n = 854).

264 **Fig. 2:** Comparison of QuikRead CRP test with central laboratory CRP values using the  
265 Bland-Altman plot (n = 854). Mean differences  $\pm 1.96$  SD for the difference are given.

266 **Table 1:** Number of serum samples found positive by the QuikRead CRP assay or the central  
 267 lab assay using a cut-off of  $\geq 10$  mg/L (n = 2922)

		CRP central lab		TOTAL
		< 10 mg/L	$\geq 10$ mg/L	
CRP QuikRead	< 10 mg/L	1797	270	2067
	$\geq 10$ mg/L	21	834	855
TOTAL		1818	1104	2922

268

269 **Table 2:** Number of serum samples found positive by the QuikRead CRP assay or the central  
 270 lab assay using a cut-off of  $\geq 30$  mg/L [5,7] (n = 2922)

		CRP central lab		TOTAL
		< 30 mg/L	$\geq 30$ mg/L	
CRP QuikRead	< 30 mg/L	2457	35	2492
	$\geq 30$ mg/L	15	415	430
TOTAL		2472	450	2922

271

272 **Table 3:** : Number of serum samples found positive by the QuikRead CRP assay or the  
 273 central lab assay using a cut-off of  $\geq 100$  mg/L (n = 2922)

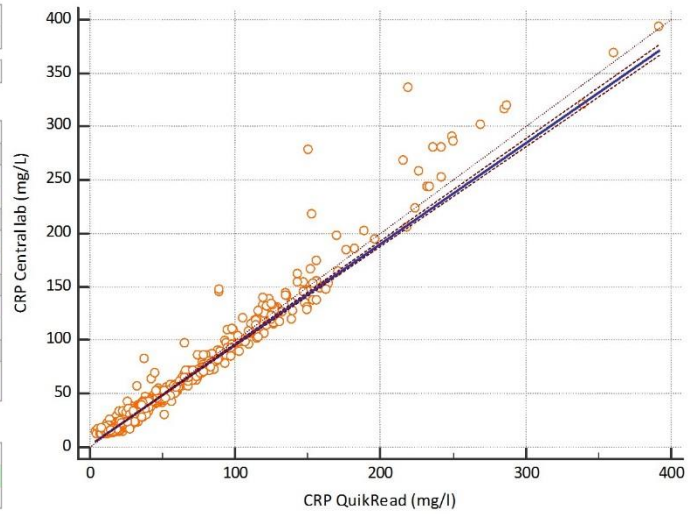
		CRP central lab		TOTAL
		< 100 mg/L	$\geq 100$ mg/L	
CRP QuikRead	< 100 mg/L	2822	7	2829
	$\geq 100$ mg/L	5	88	93
TOTAL		2827	95	2922

274



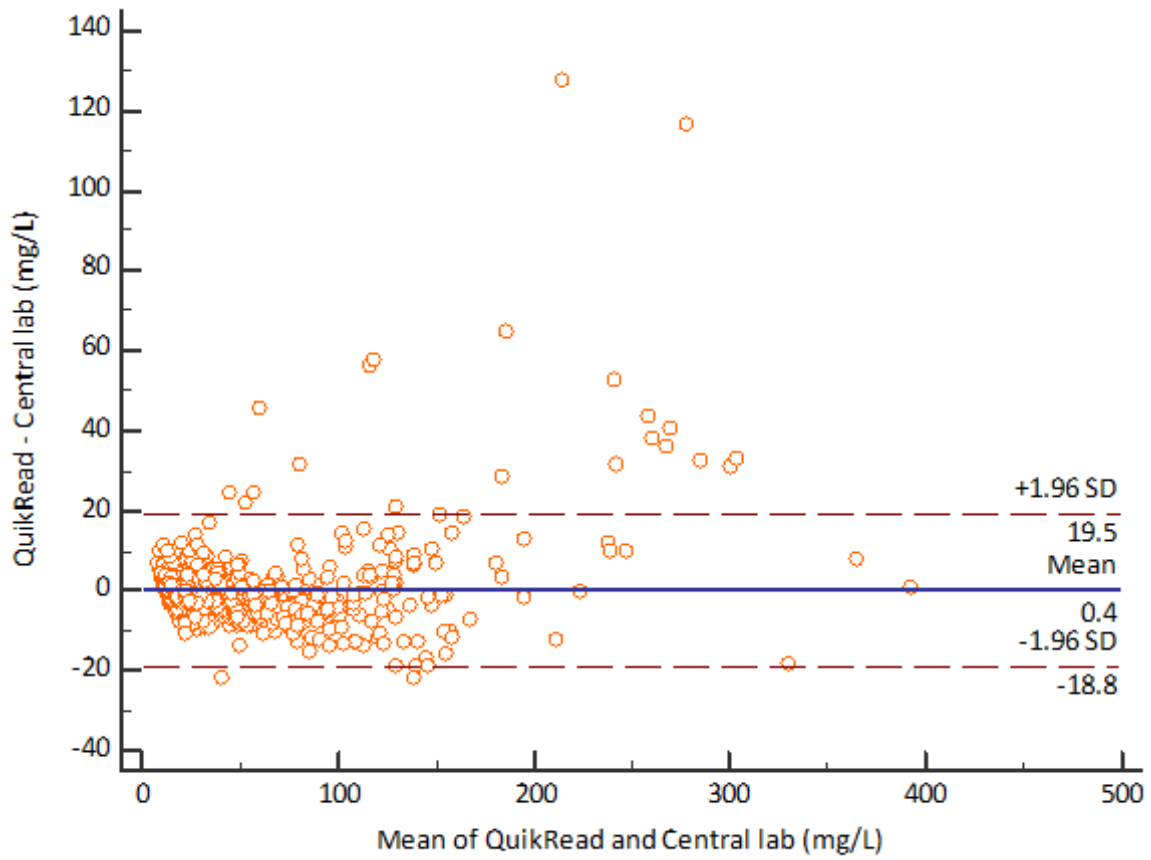
275 **Figure 1:** Comparison of QuikRead CRP test with central laboratory CRP values using  
 276 Passing-Bablok regression analysis (n = 854)

Variable X	CRP_Central_lab
Variable Y	CRP_QuikRead
Sample size	854
<b>Regression Equation</b>	
$y = 1.513640 + 0.943102 x$	
<b>Systematic differences</b>	
Intercept A	1.5136
95% CI	1.1141 to 1.8700
<b>Proportional differences</b>	
Slope B	0.9431
95% CI	0.9319 to 0.9544
<b>Random differences</b>	
Residual Standard Deviation (RSD)	7.8369
± 1.96 RSD Interval	-15.3603 to 15.3603
<b>Linear model validity</b>	
Cusum test for linearity	Significant deviation from linearity (P<0.01)
<b>Spearman rank correlation coefficient</b>	
Correlation coefficient	0.976
Significance level	P<0.0001
95% CI	0.973 to 0.979



277

278 **Figure 2:** Comparison of QuikRead CRP test with central laboratory CRP values using the  
279 Bland-Altman plot (n = 854). Mean differences  $\pm$  1.96 SD for the difference are given



280