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1 Analytical performance of a platform for point-of-care CRP testing in adults consulting for
2 lower respiratory tract infection in primary care

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22 **Abstract**

23 Background: C-reactive protein (CRP) is a biomarker widely used for disease severity
24 assessment and treatment of inflammatory conditions. Point-of-care testing (POCT) devices
25 should ideally be rapid and provide similar results to standard tests done in laboratories.

26 Methods: 2922 serum samples were obtained from adult patients presenting to primary care
27 with symptoms of lower respiratory infection in a European diagnostic study. The analytic
28 performance of the CRP QuikRead POCT device (Orion Diagnostica) was evaluated by
29 comparing results with a central laboratory method (Dimension Vista, Siemens), with both
30 tests performed in a laboratory setting.

31 Results: For a CRP cut-off concentration of ≥ 30 mg/L, the QuikRead test had a sensitivity of
32 92.2%, and specificity of 99.4%. The mean difference between QuikRead and the central lab
33 test was 0.4 mg/L. The slope of the Passing-Bablok regression was 0.94 (95% CI 0.93-0.95)
34 indicating an underestimation of CRP levels of 6% by QuikRead.

35 Conclusions: CRP estimates obtained from the QuikRead test correlate well with a central
36 laboratory assay and the measurement displays low inter-assay variation. Therefore, the
37 QuikRead test is a good candidate for CRP testing in primary care.

38

39 Keywords: C-reactive protein; point-of-care test; rapid test; bedside test; respiratory infection

40 **Introduction**

41 C-reactive protein (CRP) is the first described acute-phase protein and is now a universal
42 biomarker and early indicator of infectious or inflammatory conditions, related to diversified
43 diseases, disorders and pathological conditions [1]. Measurement of CRP levels in blood are
44 routinely performed in central hospital laboratories and many automated methods are available.
45 However, near-patient or point-of-care testing (POCT) to measure CRP levels may be helpful
46 in diagnosis and early management, for example to avoid unnecessary hospital referrals for
47 acutely ill children [2] and to guide antibiotic treatment for lower respiratory tract infections
48 [3,4]. Several POCT CRP test platforms have been developed in recent years with the
49 advantage of providing a result within minutes. In order for these devices to be used in daily
50 practice they should provide results comparable to laboratory based platforms for a wide
51 variety of patients with varying CRP levels, and indications.

52 In a European diagnostic study on lower respiratory tract infections in primary care, we have
53 previously demonstrated the added value of CRP levels in predicting pneumonia [5] which
54 can be useful to start appropriate treatment and advice and avoid unnecessary use of antibiotics.

55 In the current study we evaluate the analytical performance of a POCT device (QuikRead,
56 Orion Diagnostica) for CRP measurement in serum samples of the above mentioned trial by
57 comparing results with those obtained of our central laboratory method (Dimension Vista,
58 Siemens), with assays on both platforms done in a laboratory.

59

60 **Materials and methods**

61 Serum samples from adults (≥ 18 years of age) with symptoms of lower respiratory tract
62 infection (acute cough, ≤ 28 days) who presented to primary care were obtained in a
63 prospective observational study as part of GRACE-09 (Genomics to Combat Resistance

64 against Antibiotics in Community-Acquired Lower Respiratory Tract Infection in Europe;
65 www.grace-lrti.org) which was conducted from October 2007 to April 2010. A total of 3104
66 patients were included at 16 primary care networks in 12 European countries. The mean age of
67 patients was 49.8 years (range 18 – 92 years), 40% were male and 4.5% were diagnosed with
68 pneumonia [5,6]. Six percent of blood test data were lacking which resulted in 2922 remaining
69 individual patient results. Venous blood was drawn at the primary care facility, serum was
70 prepared at local laboratories and stored at -70°C. After transport to the central laboratory of
71 the University Hospital of Antwerp, serum samples were analyzed in batch by both the central
72 laboratory test and the POCT method on the same sample.

73 Dimension Vista® (Siemens) was the central laboratory platform used. This is a high-
74 throughput analyzer using conventional particle enhanced nephelometry to determine CRP
75 levels. The lower limit of detection of this method is 2.9 mg/L. The upper limit of detection is
76 290.0 mg/L. If this limit is exceeded, samples are diluted 2-fold and reanalyzed. The results
77 are evaluated by comparison to a low-level (CRP range: 4.4 – 7.0 mg/L) and high-level (CRP
78 range: 40.7 – 54.8 mg/L) standard (Liquichek Immunology Control, Bio-Rad).

79 The QuikRead 101 (Orion Corporation, Orion Diagnostica, Espoo, Finland) is a POCT or
80 bedside immunoturbidimetric assay based on micro particles coated with anti-human CRP. It
81 requires 20 µL of whole blood, plasma or serum and has an analytical measurement range of
82 8-160 mg/L. Values exceeding the upper detection limit are diluted and reanalyzed. A one-
83 level CRP control (CRP range: 42-62 mg/L) was analyzed on every analysis day.

84 Analyses were all performed by trained lab technicians who were blinded to the results of the
85 reference test and vice versa. Sensitivity, specificity, positive (PPV) and negative predictive
86 values (NPV) were calculated by comparing the measurements obtained by the POCT CRP
87 with the results from the central laboratory CRP. The following CRP cut-off values were used:

88 10 mg/L as the universally applied cut-off value to indicate production of CRP, 30 mg/L
89 previously shown to add most diagnostic value to a “symptoms and signs” model to predict
90 pneumonia [5] or the presence of bacterial pathogens in the airways [7] and 100 mg/L which
91 is a critical level indicating severe infection and justifying antibiotic prescription [8].

92 Considering only samples with numeric values (excluding <2.9 mg/l and <8 mg/L for the
93 central lab and POCT test, respectively), the agreement of measurements between both
94 methods was evaluated using Passing-Bablok regression analysis, Bland-Altman plot and
95 Spearman for the correlation coefficient. Criteria for acceptable correlation were defined as
96 follows: the 95% confidence interval (CI) of the slope includes 1.0, the 95% CI of the intercept
97 includes 0.0 and the correlation coefficient > 0.95. Statistical analyses were performed with
98 MedCalc Statistical Software version 17.5.5 (MedCalc Software bvba, Ostend, Belgium).

99

100 **Results**

101 The inter-assay variation of both the POCT and central lab CRP test was determined. With the
102 QuikRead test, a variation of 3.4% was obtained, while variations of 5.9% (low level) and 5.2%
103 (high level) were observed for the central lab test.

104 Considering positive CRP concentrations to be ≥ 10 mg/L, the CRP bedside test has a
105 sensitivity of 75.5% and a specificity of 98.8% when compared with the central lab CRP test
106 (Table 1). However, considering CRP concentrations ≥ 30 mg/L, the sensitivity of the CRP
107 bedside test increases to 92.2% and the specificity to 99.4% when compared with the central
108 laboratory CRP test, with a PPV of 96.5 % and an NPV of 98.6% (Table 2).The performance
109 of the QuikRead test increases even further at a CRP cut-off level of ≥ 100 mg/l (sensitivity =
110 92.6% and specificity = 99.8%) (Table 3).

111 The median and mean CRP values as well as the CRP range obtained using the QuikRead
112 POCT CRP assay on the 834 serum samples with CRP concentration ≥ 10 mg/L are similar to
113 CRP levels measured by the central laboratory assay: 32 mg/l, 50 mg/L (SD 51.2 mg/L) and
114 12 – 393 mg/L respectively for the QuikRead assay and 32.5 mg/L, 49.6 mg/L (SD 48.2 mg/L)
115 and 10.1 – 392.1 mg/L for the central laboratory method.

116 Taking into account only the samples with a numeric value (excluding <2.9 mg/L samples for
117 the central lab test and <8 mg/L samples for the QuikRead test) for the correlation analysis,
118 854 values remained. Passing-Bablok regression analysis shows a small constant bias
119 (indicated by the intercept of 1.5) and a proportional bias (indicated by the slope of the
120 regression line of 0.94) indicating that the QuikRead underestimates the CRP values with 6%
121 compared to those of the central lab analysis (Fig. 1). The Spearman correlation coefficient is
122 0.976 but a significant deviation from linearity was shown. The Bland-Altman plot shows a
123 mean difference between both methods of 0.4 mg/L (95% limits of agreement: -18.8 to 19.5
124 mg/L) which is not statistically significant (Fig. 2). The plot also shows that the reliability of
125 the POCT CRP test decreases with increased CRP values.

126

127 **Discussion**

128 In this study, the analytical performance of the commercially available POCT CRP device
129 QuikRead was evaluated by comparing results obtained from this platform in a laboratory (and
130 not at the point of care) to results from a central laboratory CRP analyzer. A CRP cut-off value
131 of 30 mg/L has previously been shown to add most diagnostic value in predicting pneumonia
132 [5] or the presence of a bacterial infection [7] in this patient population. Taking into account
133 this CRP cut-off, QuikRead shows good sensitivity and excellent specificity. Analysis of the
134 correlation between both methods, demonstrates both a small constant bias and a proportional

135 bias. The deviation at the intercept has a relatively small impact and is less relevant than the
136 slope of the linear regression line, which deviates with 6%. An underestimation of CRP levels
137 by the QuikRead test compared to central lab tests is in accordance with previous studies (17%
138 compared to the Tina-quant CRP Hitachi 912, Roche, n = 59 [9]; 15% compared to Synchron®,
139 Beckman Coulter, n = 100 [10]). However, others report good concordance (Cobas Integra,
140 Roche, n = 231 [11]; Architect c8000, Abbott, n = 250 [12]) or even an overestimation of CRP
141 levels (7% compared to the BN II System, Dade Behring, n = 72 [13]) by the QuikRead test. It
142 is clear that the large sample size in our study is an asset compared to the previously published
143 reports.

144 Compared to other CRP POCT devices, the QuikRead has a more complicated pre-analytical
145 handling of samples which takes 2.5 min [10] and is not the most user friendly apparatus [14]
146 making it more susceptible to flaws in the procedure. This could not be tested in our study
147 performing the analyses in a laboratory setting, which we preferred due to practical issues when
148 testing such a large number of samples, with qualified lab technicians. The robustness of the
149 performance of the QuikRead POCT device should therefore be evaluated using results
150 obtained from use by different clinicians and practice staff in routine primary care. Moreover,
151 the performance of the POCT device in that setting might also be influenced by the use of
152 capillary whole blood, a sampling method prone to variation, instead of the serum from venous
153 blood used in our study. However, a previous study showed no difference in CRP determination
154 in the venous and capillary blood samples using the QuikRead device [15].

155 All tested CRP POCT devices in the multi-device study performed well in general with
156 correlation coefficients > 0.95 [10]. QuikRead was shown to have low within- and between-
157 day variation (5.7% and 6.3% respectively at a CRP concentration of 100 mg/L) compared to
158 the other analyzers [14]. In our hands, the between-day variation was even lower, most
159 probably due to the execution by trained lab technicians which does not reflect the real POCT

160 environment. Furthermore, a rather short total analysis time (without pre-analytical time) of 2
161 min [10] to 3 min 20 sec [14] was described. Another advantage of the POCT device, besides
162 the rapid availability of the result, is the requirement of only small volumes of blood which can
163 be very advantageous for use in small children, a patient group not studied here.

164 POCT CRP use in primary care may increase even further in the future as obtaining chest
165 radiographs in all patients is not feasible. However, there is ongoing debate about the clinical
166 utility of CRP POCTs. Severe pneumonias are likely to be evident clinically, so the value of
167 POCT CRP tests should be where there is more doubt, in helping clinicians diagnose milder
168 pneumonias – which are more difficult to detect clinically. There should also be caution about
169 the potential for the inappropriate and inefficient use of CRP as it is introduced in practice [16],
170 given wider concerns about the danger of ‘medicalising’ largely self-limiting illness. CRP test
171 results should also always be interpreted together with clinical findings, as it was shown
172 previously that CRP levels can be low in people with pneumonia [5].

173 To the best of our knowledge, this is the first study evaluating the analytical performance of
174 the QuikRead POCT CRP test in this high number of patients with symptoms of respiratory
175 tract infection. The data presented here are part of a large multicenter observational study and
176 indicate the feasibility of the QuikRead POCT CRP test in primary care. However, large
177 prospective trials in primary care or at the point of care are needed to determine the effect of
178 CRP POCT on clinical decision-making.

179

180 **Compliance with ethical standards**

181 **Funding** The study was part of the European Union FP6 funded Network of Excellence
182 GRACE. Orion Diagnostics provided the QuikRead instruments and kits for this study. The

183 study sponsors played no role in the study design; in the collection, analysis, and interpretation
184 of data; in the writing of the report; or in the decision to submit the paper for publication.

185 **Ethical approval** All procedures performed in studies involving human participants were in
186 accordance with the ethical standards of the institutional and/or national research committee
187 and with the 1964 Helsinki declaration and its later amendments or comparable ethical
188 standards.

189 **Informed consent** Informed consent was obtained from all individual participants included in
190 the study.

191 **Conflict of interest** The authors declare that they have no conflict of interest.

192

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253

254 **Table legends**

255 **Table 1:** Number of serum samples found positive by the QuikRead CRP assay or the central
256 lab assay using a cut-off of ≥ 10 mg/L (n = 2922)

257 **Table 2:** Number of serum samples found positive by the QuikRead CRP assay or the central
258 lab assay using a cut-off of ≥ 30 mg/L [5,7] (n = 2922)

259 **Table 3:** Number of serum samples found positive by the QuikRead CRP assay or the central
260 lab assay using a cut-off of ≥ 100 mg/L (n = 2922)

261 **Figure legends**

262 **Fig. 1:** Comparison of QuikRead CRP test with central laboratory CRP values using Passing-
263 Bablok regression analysis (n = 854).

264 **Fig. 2:** Comparison of QuikRead CRP test with central laboratory CRP values using the
265 Bland-Altman plot (n = 854). Mean differences ± 1.96 SD for the difference are given.

266 **Table 1:** Number of serum samples found positive by the QuikRead CRP assay or the central
 267 lab assay using a cut-off of ≥ 10 mg/L (n = 2922)

| | | CRP central lab | | TOTAL |
|-----------------|----------------|-----------------|----------------|-------|
| | | < 10 mg/L | ≥ 10 mg/L | |
| CRP QuikRead | < 10 mg/L | 1797 | 270 | 2067 |
| | ≥ 10 mg/L | 21 | 834 | 855 |
| TOTAL | | 1818 | 1104 | 2922 |

268

269 **Table 2:** Number of serum samples found positive by the QuikRead CRP assay or the central
 270 lab assay using a cut-off of ≥ 30 mg/L [5,7] (n = 2922)

| | | CRP central lab | | TOTAL |
|-----------------|----------------|-----------------|----------------|-------|
| | | < 30 mg/L | ≥ 30 mg/L | |
| CRP QuikRead | < 30 mg/L | 2457 | 35 | 2492 |
| | ≥ 30 mg/L | 15 | 415 | 430 |
| TOTAL | | 2472 | 450 | 2922 |

271

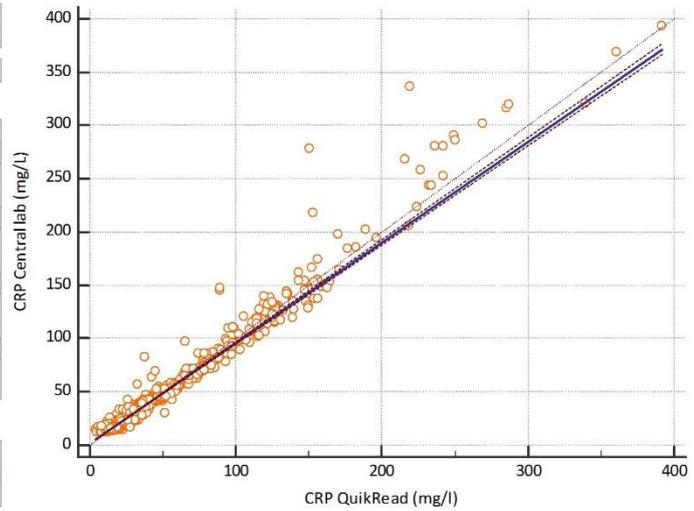
272 **Table 3:** : Number of serum samples found positive by the QuikRead CRP assay or the
 273 central lab assay using a cut-off of ≥ 100 mg/L (n = 2922)

| | | CRP central lab | | TOTAL |
|-----------------|-----------------|-----------------|-----------------|-------|
| | | < 100 mg/L | ≥ 100 mg/L | |
| CRP QuikRead | < 100 mg/L | 2822 | 7 | 2829 |
| | ≥ 100 mg/L | 5 | 88 | 93 |
| TOTAL | | 2827 | 95 | 2922 |

274

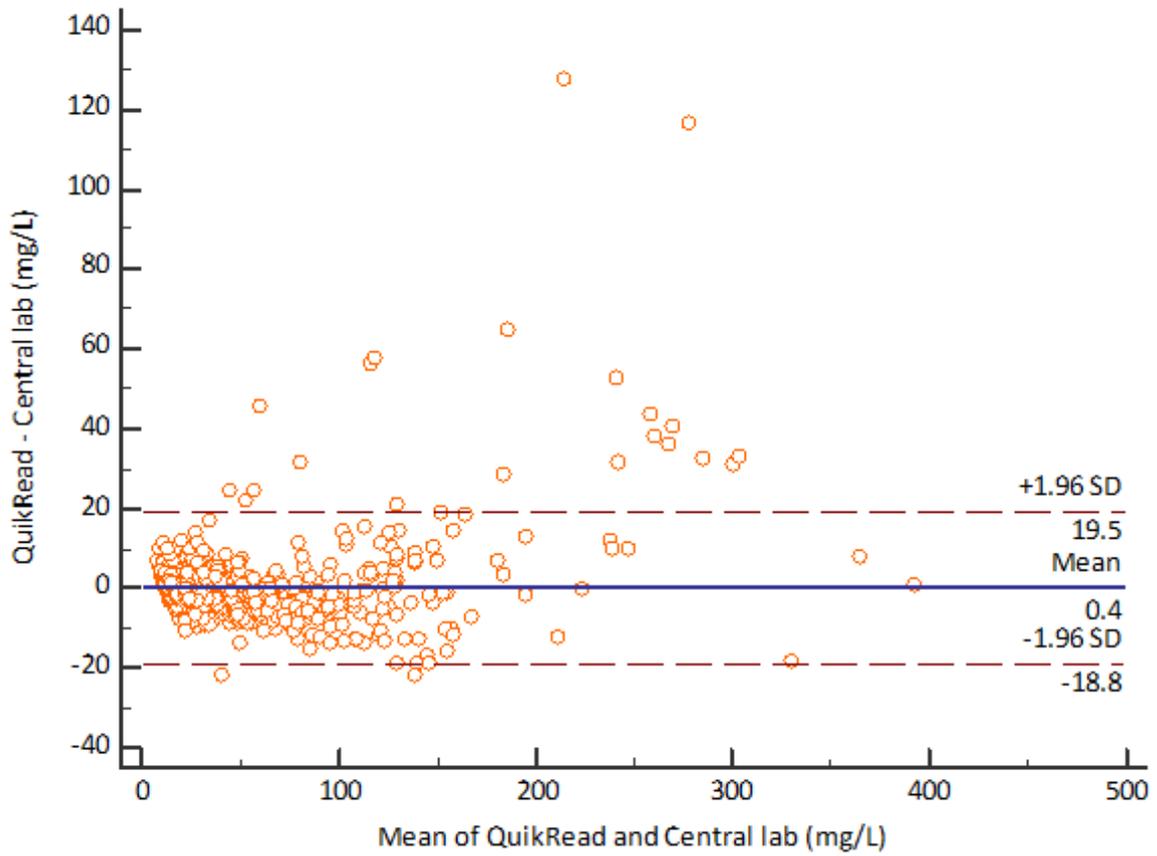
275 **Figure 1:** Comparison of QuikRead CRP test with central laboratory CRP values using
 276 Passing-Bablok regression analysis (n = 854)

| | |
|--|---|
| Variable X | CRP_Central_lab |
| Variable Y | CRP_QuikRead |
| Sample size | 854 |
| Regression Equation | |
| $y = 1.513640 + 0.943102 x$ | |
| Systematic differences | |
| Intercept A | 1.5136 |
| 95% CI | 1.1141 to 1.8700 |
| Proportional differences | |
| Slope B | 0.9431 |
| 95% CI | 0.9319 to 0.9544 |
| Random differences | |
| Residual Standard Deviation (RSD) | 7.8369 |
| ± 1.96 RSD Interval | -15.3603 to 15.3603 |
| Linear model validity | |
| Cusum test for linearity | Significant deviation from linearity (P<0.01) |
| Spearman rank correlation coefficient | |
| Correlation coefficient | 0.976 |
| Significance level | P<0.0001 |
| 95% CI | 0.973 to 0.979 |



277

278 **Figure 2:** Comparison of QuikRead CRP test with central laboratory CRP values using the
279 Bland-Altman plot (n = 854). Mean differences \pm 1.96 SD for the difference are given



280