

**This item is the archived peer-reviewed author-version of:**

Soil microbes and community coalescence

**Reference:**

Rillig Matthias C., Lehmann Anika, Aguilar-Trigueros Carlos A., Verbruggen Erik, et al.- Soil microbes and community coalescence

Pedobiologica - ISSN 0031-4056 - (2015), p. 1-4

Full text (Publishers DOI): <http://dx.doi.org/doi:10.1016/j.pedobi.2016.01.001>

## Soil microbes and community coalescence

Matthias C. Rillig<sup>1,2\*</sup>, Anika Lehmann<sup>1,2</sup>, Carlos A. Aguilar-Trigueros<sup>1,2</sup>, Janis Antonovics<sup>3</sup>,  
Tancredi Caruso<sup>4</sup>, Stefan Hempel<sup>1,2</sup>, Johannes Lehmann<sup>5</sup>, Kriszta Valyi<sup>1,2</sup>, Erik Verbruggen<sup>6</sup>,  
Stavros D. Veresoglou<sup>1,2</sup>, Jeff R. Powell<sup>7</sup>

<sup>1</sup>*Freie Universität Berlin, Institut für Biologie, Plant Ecology, D-14195 Berlin, Germany*

<sup>2</sup>*Berlin-Brandenburg Institute of Advanced Biodiversity Research (BBIB), D-14195 Berlin, Germany*

<sup>3</sup>*Department of Biology, University of Virginia, Charlottesville, VA 22904, USA*

<sup>4</sup>*School of Biological Sciences and Institute for Global Food Security, Queen's University of Belfast, Belfast BT9 7BL, Northern Ireland, UK*

<sup>5</sup>*Atkinson Center for a Sustainable Future, Department of Crop and Soil Sciences, Cornell University, Ithaca, New York 14853, USA*

<sup>6</sup>*Department of Biology, University of Antwerp, PLECO (Plant and Vegetation Ecology), 2610 Wilrijk, Belgium*

<sup>7</sup>*Western Sydney University, Hawkesbury Institute for the Environment, Penrith NSW, 2751 Australia*

\*Author for correspondence. Matthias C. Rillig, Freie Universität Berlin, Institut für Biologie, Plant Ecology, Altensteinstr. 6, D-14195 Berlin, Germany. Email [matthias.rillig@fu-berlin.de](mailto:matthias.rillig@fu-berlin.de).

**Running title:** Microbial community coalescence

**Word count:** 1724 (without references); **Figures count:** 1

## 26 **Abstract**

27 Community coalescence is a recently introduced term describing the interaction of entire  
28 communities and their environments. We here explicitly place the concept of community  
29 coalescence in a soil microbial context, exploring intrinsic and extrinsic drivers of such  
30 coalescence events. Examples of intrinsic events include the action of earthworms and the  
31 dynamics of soil aggregates, while extrinsic events are exemplified by tillage, flooding, litter-  
32 fall, outplanting, and the addition of materials containing microbial communities. Aspects of  
33 global change may alter the frequency or severity of coalescence events. We highlight  
34 functional consequences of community coalescence in soil, and suggest ways to  
35 experimentally tackle this phenomenon. Soil ecology as a whole stands to benefit from  
36 conceptualizing soil biodiversity in terms of dynamic coalescent microbial assemblages.

37

38 **Keywords:** community coalescence, metacommunity, soil biodiversity, global change,  
39 community, spatial structure, disturbance

40

## 41 **1. Introduction**

42 Community coalescence is a recently coined term (Rillig et al., 2015) describing situations  
43 where two or more entire communities (and their environments) interact because pieces of the  
44 environment that are large relative to the size of the organisms they contain can be  
45 translocated by a variety of forces. While such interactions of whole communities are hard to  
46 envisage in the normal context of plant and animal ecology, community coalescence among  
47 microbes, especially in the soil, is likely an ever-present feature. Community coalescence is  
48 only partially encompassed by existing metacommunity theory (Rillig et al., 2015; Fig. 1; also  
49 see there for a pertinent discussion of microbial biogeography), which captures the idea of  
50 connectedness, but not of wholesale exchange of environments and communities. To  
51 understand the latter in a soil context where community coalescence is likely to be common,  
52 thus necessitates a fresh look at features of such exchanges in order to develop suitable theory  
53 and experimental approaches. The purpose of this contribution is to more explicitly place the  
54 concept of community coalescence in a soil microbial context.

55 Many examples of wholesale exchanges between microbial communities come from the  
56 aquatic literature (Livingston et al., 2013; Adams et al., 2014; Souffreau et al., 2014), where

57 flows and confluence of water bodies are the natural force driving such mixing. Here we  
58 highlight such coalescence events in soils, and explore how they may help explain the large  
59 microbial biodiversity and its spatial and temporal organization. Soils are uniquely suited for  
60 thinking about community coalescence, because coalescent phenomena are likely to be  
61 commonplace there. Soil microbial soil communities are likely to provide the major systems  
62 in which coalescent processes are both functionally important and where they can be  
63 empirically investigated. This is in part because soil microbes are at the base of the soil food  
64 web, and play key roles for ecosystem processes including interactions regulating plant  
65 communities (Bever et al. 2010).

66 In the following, we differentiate between intrinsic (naturally occurring via ecological  
67 interactions) and extrinsic (as a result of external influences and disturbance) sources of  
68 coalescent events in soil. We separate between these events to illustrate how commonly  
69 occurring soil processes can be understood in the light of community coalescence. Both cases  
70 have in common the initial development of separate communities, with different abiotic  
71 conditions and community composition, which are abruptly mixed by those events.

72

## 73 **2. Soil-intrinsic coalescence events**

74 Here, we focus on soil-intrinsic coalescence events, and consider external drivers of such  
75 events in the next section (see also Fig. 1). First we also need to ask: where (and what) are the  
76 microbial communities in soil? Assemblages of organisms can be described at various spatial  
77 and temporal scales, and for our purposes we explicitly take a microbial vantage point. Beare  
78 et al. (1995), for example, designated various arenas of activities in soil, acknowledging the  
79 large physiochemical and community differences that exist in soil, sometimes in very close  
80 proximity. Importantly, these fine-scale differences in microbial assemblages potentially  
81 provide the source communities engaging in coalescence events. We illustrate this for two  
82 examples, earthworms and soil aggregates.

83 *Earthworms.*- The activity of earthworms is a particularly intriguing case of microbial  
84 community coalescence, as assemblages from various microhabitats are literally mixed up  
85 (Barois et al., 1993), e.g. by breaking up aggregates, then experiencing a passage through the  
86 gut, there encountering the earthworm gut microbial assemblage, before being finally released  
87 back into the soil (Fig. 1c). The pervasive effects of earthworms on various soil and  
88 ecosystem parameters have long been recognized. The perspective we offer here is that the

89 continuous mixing of various soil microbial communities (for example breaking up of soil  
90 aggregates), together with mixing of environments, e.g., earthworm gut physicochemical  
91 conditions, organic matter pieces, will generate persistent “non-equilibrium” environmental  
92 heterogeneity. Similar effects will also occur in other animals inhabiting the soil, such as  
93 microarthropods or nematodes, but effects will be much larger with earthworms.

94 *Soil aggregates.* -Soil aggregates are the building blocks of soil structure, creating pore spaces  
95 and providing microbial microhabitats of extreme difference at a very fine spatial scale. For  
96 example, aggregate interiors can be anaerobic, differing in a number of physicochemical  
97 properties, and as a consequence harbor microbial assemblages quite different from those in  
98 the exterior surfaces (Tiedje et al., 1984; Sexstone et al., 1985; Mummey et al., 2006).  
99 Disintegration of an aggregate exposes the interior to quite different microbial assemblages  
100 (Fig. 1g).

101 The sum of these short-term and local encounter (and re-encounter) events determines the  
102 total microbial assemblage at broader scales, including spatial and temporal heterogeneity in  
103 the distribution of the assemblage. Even though these processes must be common in soils,  
104 and have been documented phenomenologically in the extensive literature on soil disturbance,  
105 no study has investigated how community diversity changes during these events, what novel  
106 interactions are generated, or how diversity is increased or decreased (i.e. local extinctions;  
107 Veresoglou et al. 2015) by such coalescence.

108

### 109 **3. Human-mediated or externally driven community coalescence events**

110 Many external influences and disturbances would be expected to trigger fine-scaled  
111 coalescence events in the soil (Fig. 1). Examples include tillage (Fig. 1h), flooding, litter-fall,  
112 outplanting, and the addition of materials containing microbial communities, e.g. compost,  
113 stored biochar, or manure (Fig. 1d). While the net effects of all these events have been well  
114 studied, the microbial community coalescence aspect remains unquantified and poorly  
115 understood; for example, the consequences of material additions to agricultural fields are  
116 frequently examined, yet these studies do not disentangle microbial community encounters  
117 from the effects of organic matter addition. Flooding is also an interesting case where subsoil  
118 communities, which differ in microbes and carbon dynamics (Fierer et al., 2003), would  
119 encounter the topsoil community.

120 Global change can change the frequency and severity of these external drivers and thus also of  
121 coalescence events. For example, any global change factor affecting primary production (e.g.  
122 warming, elevated atmospheric carbon dioxide, artificial light at night) can have ripple-on  
123 effects on litter composition, thus creating different sized pulses of microbial inputs entering  
124 the soil system. Human-associated pathways also include accidental transport of soil (Hughes  
125 et al., 2010; Cowan et al., 2011; MacNeill et al., 2011), yet the extent to which such  
126 community coalescence events promote or hinder the dispersal of different types of microbial  
127 components is not known.

128

#### 129 **4. Functional consequences**

130 Given that community coalescence may be a common feature in soils, what does this mean  
131 for soil microbial community composition, biodiversity and ecosystem function?

132 Microbial communities in close proximity, such as in root-associated habitats or those  
133 inhabiting interiors or exteriors of soil aggregates, will have contrasting trait distributions.  
134 Depending on the frequency and mixing ratios of community coalescence events, community  
135 encounters will result in very different average trait distributions compared to the initial  
136 communities before the coalescence events. These new configurations of functional traits will  
137 in turn affect plant productivity, decomposition and nutrient cycling. As ecosystem processes  
138 encompass quite different habitats within the root-soil system, coalescence will play a major  
139 role as a mechanism underlying the relationship between soil microbial diversity and  
140 ecosystem functioning (e.g. Bell et al., 2005).

141 The coalescence of very divergent microbial communities from different soil compartments,  
142 and even their temporary coexistence, may also offer enhanced opportunities for horizontal  
143 gene transfer (Cruz and Davles, 2000), which itself will have important evolutionary and  
144 functional consequences, e.g. antibiotic resistance (Rillig et al., 2015). While such co-  
145 occurrence could also be caused by regular dispersal events, the sheer scale of exchange  
146 during coalescence is likely to be much larger and involve a greater diversity of species.

147

#### 148 **5. The way forward - how to study this phenomenon?**

149 A long term research goal is to identify the contribution of community coalescence to the  
150 composition and function of the microbial soil biota. This is now possible because of high-

151 throughput sequencing, but even with the advances in such methods, there are huge  
152 challenges, given the fine-scale heterogeneity and complexity of the soil environment, not to  
153 mention the huge diversity of microbial taxa. Hence it will be necessary to initially focus on  
154 clearly delineated, experimentally tractable compartments, for which soil ecology has already  
155 amassed a solid background. The litter-soil interaction is one such system, as is the study of  
156 community coalescence in soil aggregate turnover. The former has easily defined and  
157 experimentally realizable assemblages, the latter addresses the very essence of soil  
158 complexity and structure. Dedicated experiments to disentangle the effects of the movement  
159 of the communities from additions of other material during coalescence will be important. For  
160 example, in litter-soil interactions, experimentally uncoupling the effects of organic matter  
161 addition from the microbial community addition is a challenge. Studies (Koide et al., 2005;  
162 Osono, 2005), using culture-based methods, showed that adding sterilized leaves (i.e. without  
163 endophytes) to soil led to distinct fungal communities in the leaf litter compared to litter  
164 where the endophyte community was not excluded. Similar studies, with a focus on other soil  
165 compartments, are feasible and could shed light on the magnitude of effects on both  
166 community assembly and functions.

167 Many organisms have adaptations to exploit regular "meeting" of or re-exposure to different  
168 communities and environments. Because of their economic and medical significance, the best  
169 examples we have come from parasites, such as gut nematodes and protozoans, many of  
170 which rely on the regular contact of soil and gut communities in order to complete their life-  
171 cycles. Extending the focus from specific parasites to other "free-living" members of the  
172 community around them would be a well worthwhile endeavor. For example identifying  
173 which microbes are regularly found in the plant and the soil, or in the gut microbiome and the  
174 soil, and asking how they differ in their traits from those that are confined to each  
175 compartment alone would be a useful starting point for such research (compare Bai et al.  
176 (2015), studying *Arabidopsis* root and leaf microbiome overlap). Parasites with life-cycle  
177 components in host and soil compartments are evidence that coalescence events can  
178 contribute soil microbial diversity, but we need further studies to quantify this for the whole  
179 community. Focusing on these parasitic microbes themselves, and identifying other microbes  
180 with correlated distributions and therefore potentially moving with them could provide a  
181 useful community module on which to focus for the study of coalescence.

182 Many innovations in design and approach await study of coalescing soil assemblages.  
183 Observational approaches and directed sampling may be very productive in addressing

184 recurrent examples of coalescence such as leaf-litter fall. Equally, it is possible to envisage  
185 numerous and imaginative experimental approaches to understanding the impact of  
186 coalescence. For example, how would preventing microbial entry through leaf fall change the  
187 endophytic microbiome of a tree? How much does the functioning of this endophytic  
188 microbiome depend on recurring re-exposure to the leaf-inhabiting community? Importantly,  
189 soil ecology as a whole stands to benefit from conceptualizing soil biodiversity in terms of  
190 dynamic coalescent microbial assemblages.

191

192

193 **References**

- 194 Adams HE, Crump BC, Kling GW. Metacommunity dynamics of bacteria in an arctic lake:  
195 the impact of species sorting and mass effects on bacterial production and  
196 biogeography. *Frontiers in Microbiology* (2014) **5**:82. doi: 10.3389/fmicb.2014.00082.
- 197 Bai Y, Müller DB, Srinivas G, Garrido-Oter R, Potthoff E, Rott M, Dombrowski N,  
198 Münch PC, Spaepen S, Remus-Emsermann M, Hüttel B, McHardy AC, Vorholt JA,  
199 Schulze-Lefert P. Functional overlap of the Arabidopsis leaf and root microbiota.  
200 *Nature* (2015) **528**:364–369.
- 201 Barois I, Villemin G, Lavelle P, Toutain F. Transformation of the soil structure through  
202 *Pontoscolex corethrurus* (Oligochaeta) intestinal tract. *Geoderma* (1993) **56**:57-66.
- 203 Beare MH, Coleman DC, Crossley DA, Hendrix PF, Odum EP. A hierarchical approach to  
204 evaluating the significance of soil biodiversity to biogeochemical cycling. *Plant and*  
205 *Soil* (1995) **170**:5-22.
- 206 Bell T, Newman JA, Silverman BW, Turner SL, Lilley AK. The contribution of species  
207 richness and composition to bacterial services. *Nature* (2005) **436**:1157–1160.
- 208 Bever JD, Dickie IA, Facelli E, Facelli JM, Klironomos JN, Moora M, Rillig MC, Stock WD,  
209 Tibbett M, Zobel M. Rooting theories of plant ecology in microbial interactions.  
210 *Trends in Ecology & Evolution* (2010) **25**:468-478.
- 211 Cowan DA, Chown SL, Convey P, Tuffin M, Elughes K, Pointing S, Vincent WF. Non-  
212 indigenous microorganisms in the Antarctic: assessing the risks. *Trends in*  
213 *Microbiology* (2011) **19**:540-548.
- 214 de la Cruz F, Davies J. Horizontal gene transfer and the origin of species: lessons from  
215 bacteria. *Trends in Microbiology* (2000) **8**:128-133.
- 216 Fierer N, Allen AS, Schimel JP, Holden PA. Controls on microbial CO<sub>2</sub> production: a  
217 comparison of surface and subsurface soil horizons. *Global Change Biology* (2003)  
218 **9**:1322-1332.
- 219 Hughes KA, Convey P, Maslen NR, Smith RIL. Accidental transfer of non-native soil  
220 organisms into Antarctica on construction vehicles. *Biological Invasions* (2010)  
221 **12**:875-891.
- 222 Koide K, Osono T, Takeda H. Colonization and lignin decomposition of *Camellia japonica*  
223 leaf litter by endophytic fungi. *Mycoscience* (2005) **46**:280-286.
- 224 Livingston G, Jiang Y, Fox JW, Leibold MA. The dynamics of community assembly under  
225 sudden mixing in experimental microcosms. *Ecology* (2013) **94**:2898-2906.

226 McNeill M, Phillips C, Young S, Shah F, Aalders L, Bell N, Gerard E, Littlejohn R.  
227       Transportation of nonindigenous species via soil on international aircraft passengers'  
228       footwear. *Biological Invasions* (2011) **13**:2799-2815.

229 Mummey DL, Rillig MC, Six J. Endogeic earthworms differentially influence bacterial  
230       communities associated with different soil aggregate size fractions. *Soil Biology &*  
231       *Biochemistry* (2006) **38**:1608-1614.

232 Osono T. Colonization and succession of fungi during decomposition of *Swida controversa*  
233       leaf litter. *Mycologia* (2005) **97**:589-597.

234 Rillig MC, Antonovics J, Caruso T, Lehmann A, Powell JR, Veresoglou SD, Verbruggen E.  
235       Interchange of entire communities: microbial community coalescence. *Trends in*  
236       *Ecology & Evolution* (2015) **30**:470-476.

237 Sexstone AJ, Revsbech NP, Parkin TB, Tiedje JM. Direct measurement of oxygen profiles  
238       and denitrification rates in soil aggregates. *Soil Science Society of America Journal*  
239       (1985) **49**:645-651.

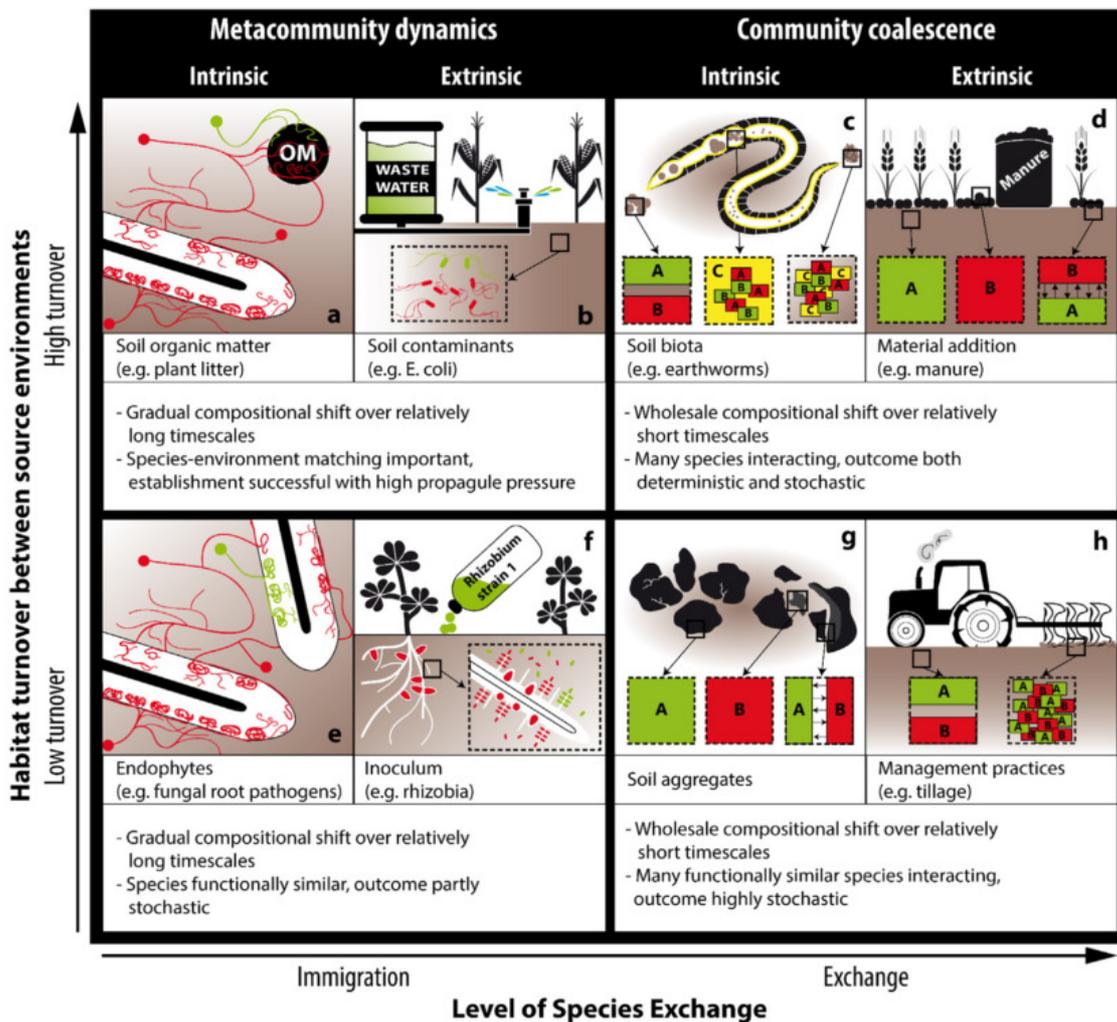
240 Souffreau C, Pecceu B, Denis C, Rummens K, De Meester L. An experimental analysis of  
241       species sorting and mass effects in freshwater bacterioplankton. *Freshwater Biology*  
242       (2014) **59**:2081-2095.

243 Tiedje JM, Sexstone AJ, Parkin TB, Revsbech NP, Shelton DR. Anaerobic processes in soil.  
244       *Plant and Soil* (1984) **76**:197-212.

245 Veresoglou SD, Halley J, Rillig MC. Extinction risk of soil biota. *Nature Communications*  
246       (2015) **6**: 8862.

247

248



250

251 Fig. 1. Conceptual overview depicting metacommunity and coalescence phenomena occurring  
 252 in soils, divided into intrinsically (a, c, e, g) and extrinsically (b, d, f, h) driven events (see  
 253 text). The distinction between metacommunity dynamics and community coalescence is  
 254 depicted as the degree of species (and environmental) exchange along the x-axis, in reality a  
 255 continuum ranging from individual species dispersing to the wholesale interchange  
 256 characteristic of community coalescence. The y-axis shows the degree of difference between  
 257 the source environments for either metacommunity or community coalescence events. Picture  
 258 inserts illustrate examples and text describes the nature and outcome of exchanges among  
 259 communities. In the metacommunity examples, red colored organisms represent established  
 260 and green colored the newly arriving species. For the coalescence examples, the red, green  
 261 and yellow colored shapes display different communities.