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1 **Robust strategies to eliminate endocrine disruptive estrogens in water resources**

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19

20 **Abstract**

21 The widespread occurrence and ubiquitous distribution of estrogens, i.e., estrone (E1),
22 estradiol (E2), and estriol (E3) in our water matrices, is an issue of global concern. Public
23 and regulatory authorities are concerned and placing joint efforts to eliminate estrogens

24 and related environmentally hazardous compounds, due to their toxic influences on the
25 environmental matrices, ecology, and human health, even at low concentrations.
26 However, most of the available literature is focused on the occurrence of estrogens in
27 different water environments with limited treatment options. Thus, a detailed review to
28 fully cover the several treatment processes is needed. This review comprehensively and
29 comparatively discusses many physical, chemical, and biological-based treatments to
30 eliminate natural estrogens, i.e., estrone (E1), estradiol (E2), and estriol (E3) and related
31 synthetic estrogens, e.g., 17 α -ethinylestradiol (EE2) and other related hazardous
32 compounds. The covered techniques include adsorption, nanofiltration, ultrafiltration,
33 ultrasonication, photocatalysis of estrogenic compounds, Fenton, Fenton-like and photo-
34 Fenton degradation of estrogenic compounds, electro-Fenton degradation of estrogenic
35 compounds, ozonation, and biological methods for the removal of estrogenic compounds
36 are thoroughly discussed with suitable examples. The studies revealed that treatment
37 plants based on chemical and biological approaches are cost-friendly for removing
38 estrogenic pollutants. Further, there is a need to properly monitor and disposal of the
39 usage of estrogenic drugs in humans and animals. Additional studies are required to
40 explore a robust and more advanced oxidation treatment strategy that can contribute
41 effectively to industrial-scale applications. This review may assist future investigations,
42 monitoring, and removing estrogenic compounds from various environmental matrices.
43 In concluding remarks, a way forward and future perspectives focusing on bridging
44 knowledge gaps in estrogenic compounds removal are also proposed.

45 **Keywords:** Estrogens; Environmental impact; Occurrence; Toxicity; Elimination
46 strategies

47

48 **1. Introduction**

49 Estrogens are a group of hormones available in either form, i.e., synthetically developed
50 or natural. Among estrogens, i.e., estrone (E1), estradiol (E2), and estriol (E3) are three
51 primary endogenous forms that have estrogenic hormonal activity and play a crucial
52 health role. More specifically, these have been categorized as female sex hormones that
53 are mainly responsible for the development and regulation of the reproductive system of
54 females and the maintenance of secondary sexual characteristics. Besides natural
55 estrogens, several synthetic forms have been developed via the chemical synthesis route.
56 Synthetic estrogens are compounds that possess estrogenic activity, e.g., 17 α -
57 ethinylestradiol (EE2), estradiol valerate, estropipate, conjugate esterified estrogen, and
58 quinestrol. Whereas diethylstilbestrol, hexestrol, dienestrol, methestrol, and benzestrol
59 are some key examples of non-steroidal synthetic estrogens. Synthetic estrogens are
60 being extensively consumed alone or in combination with progestins as an oral
61 contraceptive pill, approved by the Food and Drug Administration (FDA) USA, treat
62 menopause symptoms, osteoporosis prevention, hypoestrogenism, and metastatic
63 breast and advance prostate prevention (Cobin and Goodman, 2017; Levin, 2018; Yang,
64 He, et al., 2021).

65 Due to the control or uncontrolled consumption of estrogens, their ultimate excretion in
66 urine and feces from humans and livestock in partially metabolized or relatively un-
67 metabolized form finds its way to water matrices (Combalbert and Hernandez-Raquet,
68 2010; Yang, He et al., 2021). Estrogens are metabolized in the liver, and cytochrome 450
69 is mainly responsible for initiating estrogens' metabolism. This enzyme is abundantly

70 expressed in the liver. Hydroxylation is the first step carried out by cytochrome 450 for
71 the metabolism of estrogens. Metabolism alters the intensity of estrogens' action and also
72 changes the profile of its effects on targeted organs (Tsuchiya, Nakajima, & Yokoi, 2005).
73 Steroidal estrogens are increasingly being noticed in the water environment around the
74 globe. Consequently, the hormonal compounds with estrogenic activities are unavoidably
75 merged into the water matrices through ineffectively treated wastewater and
76 runoff/disposal of animal wastes in freshwater or leakage into the groundwater bodies.
77 Besides human and livestock waste, continuous discharge of estrogen-residual-
78 containing industrial wastewater effluents adversely affects aquatic life. For instance,
79 endocrine disruption is one of such adverse effects as these bioactive compounds disrupt
80 the endocrine systems of aquatic organisms (Liang et al., 2020; Šauer et al., 2020),
81 revealing evolving disorders in animals and humans. Estrogens stimulate developmental
82 disorders in aquatic animals (Chiang et al., 2020; Wang et al., 2019). For instance, Šauer
83 et al. (2021) reported the effects of synthetic estrogens, i.e., progestins gestodene and
84 drospirenone, on the sex differentiation in common carp (*Cyprinus carpio*) from embryonic
85 to juvenile stage. Drospirenone, gestodene, and their mixture have anti-gonadotropic
86 activity in carp (*Cyprinus carpio*). Moreover, the exposure of test organisms to a binary
87 mixture of drospirenone and gestodene triggered an enlarged frequency of intersex (32%)
88 as compared to clean water and solvent control groups (both 3%). In an earlier study, Shi
89 et al. (2020) reported the effects of synthetic progestin medroxyprogesterone acetate
90 (MPA, 4.3 ng/L) or dydrogesterone (3.39 ng/L) on eye growth and the transcription of
91 associated genes in zebrafish. The environmental protection agency (EPA) finalized the
92 rule under the safe drinking water act necessitating water systems to monitor estrogen

93 hormones as part of an unregulated contaminant monitoring program (Agency, 2011).
94 The European Parliament has voted to add EE2 to the European Union watch list under
95 the water framework directive, the EU's main water policy instrument for setting anti-
96 pollution approaches (Chatain, 2013). Considering the above-mentioned scenarios,
97 estrogens and related pharmaceutically active residues are increasingly being detected
98 in our water bodies heavily consumed by humans, the livestock sector, and the
99 biomedical sector. Advance wastewater treatment technologies are being globally
100 focused on these days to reduce water pollution (Bilal, Rizwan, Adeel, & Iqbal, 2021; Gan
101 et al., 2022, Rizwan et al., 2022, Rasheed et al., 2019). Water resources are reducing
102 day by day, so challenges affiliated with the removal of estrogens have received great
103 attention due to their harmful effects. Simply changing the operational mode of
104 conventional wastewater treatment plants is not enough to mitigate estrogens. Advance
105 treatments like nanofiltration (NF), membrane bioreactors (MBRs), reverse osmosis (RO)
106 and ultrafiltration (UF), and advanced oxidation process (AOPs) are efficient to remove
107 estrogens. However, these processes have the drawback of high cost (Shahid, Kashif,
108 Fuwad, & Choi, 2021). Integrated treatment systems have been introduced recently to
109 overcome the limitations of conventional and advanced processes. These integrated
110 systems may combine physical, chemical, and biological treatments to obtain the
111 significant removal of estrogens (Ensano, Borea, Naddeo, de Luna, & Belgiorno, 2019).
112 Different biological treatments, including microbial strains and enzymes, have attained
113 great attention as cost-friendly, sustainable and significant tools for the removal of
114 estrogens (Bilal, Adeel, Rasheed, Zhao, & Iqbal, 2019; Ma, Shen, Tang, Li, & Dai, 2022;
115 Singh, Bilal, Iqbal, & Raj, 2021). However, optimization and enzyme stability remained

116 critical always in biological treatment systems. Several studies have reported the
117 widespread distribution and existence of estrogens and related compounds of
118 environmental concern (Combalbert and Hernandez-Raquet, 2010; Yang, He et al.,
119 2021). Several treatment methods (physical, chemical and biological) have been
120 proposed and deployed to tackle this issue of global concern. A detailed comparison was
121 made by considering physical, chemical, and biological treatments (Table 1). However,
122 no specific process exists to eliminate estrogens in wastewater treatment plants
123 (Camacho-Munoz et al., 2012; Huang et al., 2019; Aguilar-Pérez et al., 2021). The current
124 literature lacks a detailed overview of all possible methods in a single place. Thus, herein,
125 an effort was made to spotlight the potential of all treatment processes. Among many
126 reported methods, bio-based treatment systems that include the implementation of
127 microbes as natural factories and their unique enzyme-based catalytic systems have
128 governed a tremendous research consideration (Bilal et al., 2019; Singh et al., 2021a;
129 2021b; 2021c). So far, most of the available literature is focused on the occurrence of
130 estrogens and related hazardous compounds with limited treatment options. A detailed
131 review to fully cover the several treatment processes is needed. This paper
132 comprehensively reviewed many physical, chemical, and biological treatments to
133 eliminate estrogens and related environmentally hazardous compounds. The major
134 objective of this review article includes a brief knowledge about estrogens, operational
135 perspectives of various advanced physical, chemical, and biological processes to remove
136 estrogens, knowledge about mechanisms involved in estrogen removal, and recognition
137 of gaps for future developments.

138 **2. Estrogens – Sources and environmental fate**

139 Given a nonstop upsurge, discharge of estrogens and other pharmaceutically active
140 compounds is likely to increase the bioaccumulation of estrogens into various receiving
141 water systems (Jiang et al., 2013). Several steroidal estrogens, including E1, E2, E3, and
142 EE2, have been extensively disseminated in different water matrices. For instance, liquid
143 manure, feces, solid livestock waste, lagoon waste, and animal manure have been
144 reported as the primary source of estrogens detection (Biswas et al., 2013). A plethora of
145 literature is available that substantiates the occurrence of estrogens in groundwater,
146 freshwater, and wastewater, though in different quantities from different sources. For
147 example, Yang, He et al. (2021) reported the first large-scale assessment of 34 natural
148 and synthetic sex hormones, including five other estrogens in soils. In addition, the
149 occurrence and distribution of estrogenic substances in surface waters have been
150 reported by Deich et al. (2021). According to the results obtained, natural estrogens, i.e.,
151 E1, E2, and synthetic EE2 were the most abundant, with maximum concentrations up to
152 1.1 ng L^{-1} , 0.7 ng L^{-1} and 0.6 ng L^{-1} , respectively. Regardless of the concentration and
153 type, estrogens primarily enter the natural water matrices via sewage treatment plants
154 and are consequently transported into the oceans (Xu et al., 2014; Deich et al., 2021).
155 Routes of estrogens contamination to ground and surface water bodies are schematically
156 shown in Figure 1.

157 **3. Physical methods for removal of estrogenic compounds**

158 Removing estrogens from water bodies was imperative due to their toxic nature and
159 harmful effects. In this prospect, various physical, biological, and chemical methods have
160 been used in accordance with requirements and efficiency toward removal. Such as,
161 biological processes are beneficial for degrading many organic molecules but are limited

162 to toxic inorganic materials (Gao et al., 2020). In addition, chemical methods such as the
163 advanced oxidation process are useful for removing harmful estrogens with the formation
164 of some byproducts and limiting their applications. These limitations allow biological and
165 physiochemical characterization such as pH, the acid constant, the partition coefficient of
166 water, polarity, hydrophilicity, and degradability measurement at first (Grelska et al., 2020;
167 Simazaki et al., 2015). On the other hand, physical methods include adsorption, reverse
168 osmosis, membrane filtration, nanofiltration, and ultrafiltration, which are very useful for
169 removing inorganic molecules. However, membrane filtration shows limitations due to
170 pore blockage, which causes fouling problems (Si et al., 2019; Katibi et al., 2021).

171 **3.1. Adsorption of estrogenic compounds**

172 The removal of estrogens compound by using solid-phase (adsorbent) in an aqueous
173 phase is a useful technique due to its most efficient, effective and economic prospect.
174 Efficiently, the ability of adsorbent and removal of the particular compound first monitored
175 through specific sorption coefficient (ratio of adsorbed and dissolved targeted compound)
176 and isotherms equations, respectively. Several adsorbents, including activated sludge,
177 activated carbon (ACs), carbon nanotubes (CNTs), graphene oxide (GO), ion exchange
178 resin, molecularly imprinted polymers, chitin, carbonaceous material from waste, and
179 chitosan can be used for the removal of dissolved estrogens. Activated sludge removed
180 estrogens through sorption and degradation after removing excess sludge and became
181 a principal technique to purify sewage (Ren et al., 2007; Andersen et al., 2005; Johnson
182 and Sumpter, 2001; Khanal et al., 2006). The involvement of two processes made this
183 process unclear to understand which one is predominant in eliminating toxic estrogens
184 (Ren et al., 2007). The solubility of natural estrogens in water is less than synthetic ones

185 with a low Henry's law coefficient and less active toward biodegradability. This makes
186 sense that their removal is mainly based on the adsorption process compared primarily
187 to degradation, especially when the retention time with sludge is less (Ren et al., 2007).
188 On the contrary, some researchers concluded that removing estrogen by adsorption
189 process is less than 10 % (Andersen et al., 2003, 2005; Janex-Habibi et al., 2009; Joss
190 et al., 2004; Muller et al., 2008; Ternes et al., 2004). This can be happened due to different
191 experimental setups, as adsorption of estrogens mainly depends on particle size,
192 temperature, total organic carbon, ions content, salinity, and pH. Activated carbon (ACs)
193 is another efficient and effective adsorbent for removing many organic and inorganic
194 estrogens from water bodies. It is widely used for adsorbents due to its efficiency and
195 economical ease in availability (Babel and Kurniawan, 2003; Rivera-Utrilla et al., 2011).
196 In this direction, activated carbon granulate shows high adsorption capacity, and the
197 kinetically controlled process can be affected by environmental conditions (Zhang and
198 Zhou, 2005). The pore size of activated carbon can also affect adsorption capacity, i.e.,
199 an increase in pore size reduces the adsorption of estrogens due to their high hydrophobic
200 nature (Fukuhara et al. 2006). Another factor that controls the adsorption ability of ACs is
201 the organic matter present in water bodies irrespective of estrogens due to their high
202 speed of movement toward ACs as compared to estrogens and cover available sites for
203 adsorption (Fukuhara et al., 2006; Kumar et al., 2009; Zhang and Zhou, 2005).
204 In addition, carbon nanotubes (CNTs) are very useful adsorbents for estrogens due to
205 their excellent mechanical and electronic properties (Kurwadkar et al., 2019). A rolled
206 graphene sheet is present at the surface of CNTs for single-walled CNTs, whereas
207 multiple sheets of graphene are known as multiwalled CNTs (MWCNTs). Irrespective of

208 ACs, CNTs show considerably different properties as regardless of surface area and pore
209 size, their adsorption ability mainly depends on purity, functionalities, and adsorption sites
210 (Arora and Attri, 2020). The mechanism of estrogens removal is primarily based on pi-pi
211 bond formation and van der Waals interactions among CNTs and estrogens and is
212 responsible for adsorption performance (Kurwadkar et al., 2019). The adsorption ability
213 of CNTs can be enhanced by increasing interstitial space and with the attachment of
214 different functionalities according to the removal of specific estrogens. The application of
215 CNTs on a larger scale is still in the process due to their more toxic effects than ACs (Das
216 et al., 2018). Another family of adsorbent, molecularly imprinted polymers (MIPs) can be
217 used for water treatment to remove toxic waste materials. Several MIPs have been
218 synthesized for this purpose. MIPs adsorbent with an estradiol template is used to study
219 water purification by adding some estrogens in deionized water. These MIPs can be
220 reused after treatment with methanol/acetic acid followed by methanol (Meng et al.,
221 2005). These MIPs can also functionalize with different materials to prepare an adsorbent
222 with multiple adsorption sites. Gao and his coworkers functionalized MIPs with carbon
223 nanotubes and removed estrogens with selectively fast kinetics and high capacity for the
224 molecule's uptake with an adsorption capacity of 113.5 mmol/g. These CNTs
225 functionalized MIPs show efficiency of up to 95 to 98% and remain constant for different
226 water bodies (Gao et al., 2011; Joseph et al. 2011). Graphene oxide sheets can also be
227 used as an adsorbent for removing estrogens due to their excellent hydrophilicity, a more
228 significant number of oxygens containing functional groups, and high surface area. The
229 maximum adsorption capacity 149.4 mg/g of GO was reported by Jiang and coworkers
230 (Gupta et al., 2017; Jiang et al., 2016). Cellulose is another promising material used for

231 the adsorption of estrogen from water. Although cellulose shows less adsorption capacity
232 towards estrogen in aqueous media with low reproducibility, several modified cellulose
233 adsorbents have been used for water treatment applications (Adewuyi, 2020). Using
234 cellulose for estrogen removal opens a path for using natural polymers having good
235 sustainability and are highly effective at a low cost. These materials for water treatment
236 are only limited to general water contaminants and are not widely applied to remove
237 estrogens.

238 **3.2. Nanofiltration of estrogenic compounds**

239 Natural organic matter and micropollutants can be removed from wastewater and drinking
240 water through the nanofiltration (NF) membrane technique. The separation of estrogen
241 from water bodies through nanofiltration depends on steric hindrance and interaction of
242 charged toxic pollutants with membranes (Berg et al., 1997). Many studies employed
243 crossflow filtration and dead-end-cell filtration to remove estrogens by nanofiltrations. The
244 estrogens removal ability of NF membranes ranges from 1 to 1000 ng/L depending on
245 ionic strength, pH, charge, and contact angle of water with membranes materials (Nghiem
246 et al., 2004; Schafer et al., 2003; Kimura et al., 2003). Different parameters affect various
247 types of estrogens present in water bodies. Such as, the removal of hormones less
248 depends on ionic strength whereas more depends on pH due to repulsive interaction
249 between estrogens and membranes themselves. The mechanism of NF membranes
250 technique is based on multiple processes, including adsorption, charge repulsion and size
251 exclusion. These three mechanisms mainly decide estrogens' removal with different
252 efficiency depending on the type of compound to be removed. The wastewater contains
253 contaminants and estrogen forced toward the pores of the membrane (size 0.2 nm and

254 0.4 nm), and electrostatic charge and size help them attach to the membranes surface
255 through adsorption (Kingsbury et al., 2013; Sun et al., 2017). The surface of NF
256 membranes can be composed according to the required charge density in accordance
257 with contaminants. During purification, divalent and multivalent ions stuck at the surface
258 of the membrane and water with monovalent ions are allowed to pass (Dharupaneedi et
259 al., 2019). In practical studies, NF method has been used to remove selective estrogens
260 and achieve water with high quality and less organic matter. In this method, no chemical
261 disinfectant is needed to remove microbes and viruses (Li et al., 2020; Datel and
262 Hrabankova, 2020). The mechanism of selective removal of contaminants in NF
263 membrane method is fundamental to understand. Semiao and coworkers found that the
264 pore size plays a key role in the selective contaminant's reduction in NF membranes.
265 Nevertheless, removing many pollutants with a single purification method is still difficult.
266 Therefore, in many applications, the NF membranes method has been used in connection
267 with other water treatments to purify water bodies in multiple steps efficiently.

268 **3.3. Ultrafiltration of estrogenic compounds**

269 Ultrafiltration (UF) is another membrane technology widely used to treat water bodies
270 containing estrogens. Like nanofiltration, the contaminants are collected over a
271 membrane through the filtration process. The filtration process can be changed by
272 changing the pore size, surface charge density, and hydrophobic nature depending on
273 which type of contaminant must be removed by membranes (Schäfer et al., 2011;
274 Simmons et al., 2011). Similarly, hydrostatic pressure used to force water toward
275 membranes also plays an essential role in controlling the particles at the surface of
276 membranes. If the pore size of membranes is more significant, i.e., in micrometer,

277 ultrafiltration becomes known as microfiltration and works on a standard atmospheric
278 pressure (Zhou et al., 2010; Lu and Liu, 2010). On the other hand, membranes with pore
279 size 0.001-0.1 μ m are used for filtration, known as ultrafiltration, and its efficiency depends
280 on membranes type and the type of contaminants to be removed (Zhou et al., 2010; Lu
281 and Liu, 2010; Acero et al., 2010; Cissé et al., 2011; Gao et al., 2011; Lidén and Persson,
282 2016). In the ultrafiltration method, membranes operate at low pressure with a crossflow
283 separation. Due to the larger pore size of membranes, this method is mainly used to
284 remove larger particles known as macromolecules. This is a secondary treatment method
285 of considerable attention and extensive application with limited, targeted estrogens
286 removal (Aziz and Ojuma et al., 2020). Patel et al. (2019) used ultrafiltration membranes
287 to remove pharmaceuticals and found that only selected contaminants can be removed
288 using this method. This happened due to the different working mechanisms of NF
289 membranes and UF membranes. In NF membranes, the removal of contaminants is
290 based on hydrophobic adsorption and size exclusion, whereas only hydrophobic
291 adsorption occurs in UF membranes. Therefore, UF membranes are rare to use a single
292 treatment purification of water bodies. Such as, it is very useful in combination with the
293 reverse osmosis process as the contaminants allowed by UF membranes due to low
294 molecular weight can be removed by reverse osmosis membranes. Sun and his teams
295 concluded that particles of larger size could be rejected at the surface of membranes,
296 whereas smaller particles remain concentrated with water bodies and pass-through
297 reverse osmosis membranes with leachate effluents (Sun et al., 2015; Dharupaneedi et
298 al., 2019).

299 **3.4. Ultrasonication (US) of estrogenic compounds**

300 The removal of estrogens using different techniques mainly depends on their
301 physicochemical properties. Ultrasonication (US) method is now advantageous and is an
302 advanced method for removing complex organic/inorganic contaminants from water
303 bodies (Rao et al., 2016; Pokhalekar et al., 2016). This method is known as
304 sonodegradation and produces H_2O_2 , $OH\cdot$, $OH_2\cdot$, $OOH\cdot$ Species by sonolysis of water at
305 high temperature and pressure by nucleation collapse caused by ultrasound waves.
306 These US waves cause chemical and thermal breakage of contaminants into gaseous
307 and aqueous states (Naddeo et al., 2013). US treatment method is a clean, safe, and
308 energy-saving method without producing secondary pollutants compared to other water
309 treatment methods (Laxmi et al., 2010). During US treatment, the efficiency mainly
310 depends on the size, acidity, shape, hydrophobicity, and functionality of estrogens and
311 ultrasonication conditions. Another advantage of US treatment is the removal of
312 contaminants in a small concentration compared to other water treatment processes. Im
313 et al. (2014) performed the experiments to remove acetaminophen and naproxen by US
314 treatment and found an excellent efficiency of about 99 % and 86.1 %, respectively.
315 Further studies about the removal of estrogens revealed that efficiency mainly depends
316 on the chemical structure of compounds. Such as a pharmaceutical contaminant,
317 triclosan degraded 95 %, whereas iopromide and erythromycin removed about 50 %.
318 Several factors can affect estrogens' removal by the US method, including promoters,
319 catalysts background ions, water quality, pH, US frequency, temperature, reactor type,
320 US power, and properties of estrogens (Chu et al., 2017). To get high performance by US
321 treatment, all these factors need to be controlled. In this regard, low pH and high
322 temperature and pressure push radical formations in US treatment and increase their

323 efficiency. Similarly, many catalysts can also improve the cavitation bubbles for effective
324 US treatment (Chu et al., 2017).

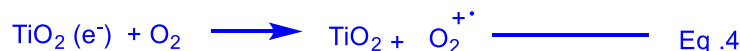
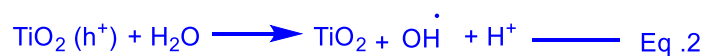
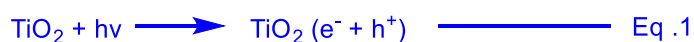
325 **4. Chemical methods for removal of estrogenic compounds**

326 Advanced oxidation processes are initiated through free radicals and various reactive
327 oxidative species to degrade organic pollutants. Here we have discussed different
328 Advance oxidation-based processes, including Fenton, Fenton like photo-Fenton, electro-
329 Fenton, photocatalytic and various ozone-based processes for degradation of estrogens
330 (E1, E2, E3, and EE2) (Figure 2).

331 **4.1. Photocatalysis of estrogenic compounds**

332 In photocatalytic treatment, the compound degradation is enhanced by activating a
333 catalyst through light energy (Figure 3) (González-González et al., 2022a; González-
334 González et al., 2022b). Catalyst plays a role in improving the reaction rate, and itself
335 remains unaltered after reaction completion. Photocatalyst consists of semiconductor
336 particles that contain a large surface area, valence band and conduction band. Between
337 the valence and conduction bands, there is an energy difference called a bandgap. The
338 photocatalytic reaction initiates when a photon of energy equal or greater to the band-gap
339 is absorbed by the catalyst and promotes the electron transfer from the valence band to
340 the conduction band, and this leads to the generation of a positive hole as well (Leyva,
341 Moctezuma, Baines, Noriega, & Zarazua, 2018). After absorption of light by the
342 photocatalyst, the valence band to conduction band electrons get to transfer, and holes
343 (h^+) are produced in the valence band (Eq. 1). Then hydroxyl radicals are formed by the
344 reaction of the positive hole with water molecules adsorbed on the catalyst's surface (Eq.
345 2). The target analyte that is adsorbed onto the catalyst's surface may react with the

346 generated hole or electron (Eq. 3), and recombination of charged species is also possible.
 347 Superoxide radicals may also cause by the reaction of conduction band electrons with
 348 oxygen (Eq. 4). The most used photocatalysts involve ferric oxide (Fe₂O₃), tin oxide
 349 (SnO₂), zinc oxide (ZnO), zinc sulfide (ZnS), silicon, and titanium dioxide (TiO₂). Titanium
 350 dioxide is considered one of the best photocatalysts because it is photo-stable,
 351 commercially available, low toxic, and can degrade various molecules under ambient
 352 conditions. TiO₂ exists across multiple mineral forms like rutile, anatase, and brookite.
 353 Among them, anatase is considered the best choice as a photocatalyst because of the
 354 transport of excitons to the surface (Luttrell et al., 2014). P25, which consists of almost
 355 85% anatase and 15% rutile, is the most widely employed TiO₂. Photocatalytic reactions
 356 involving TiO₂ have been described in Eq. 1-4, where h⁺ represents hole forms through
 357 missing electrons and P_{ad} represents the adsorbed pollutant on the catalyst surface.
 358 Previously TiO₂ has been used widely to degrade estrogens by many scientists
 359 (Sornalingam, McDonagh, & Zhou, 2016).



360
 361 Menon reported photocatalytic degradation of E2 (17β-estradiol) and EE2 (17α-
 362 ethinylestradiol) by using TiO₂-ZnO nanocomposite under UV and visible irradiations.
 363 100% degradation was obtained for both estrogenic compounds after 240 min@
 364 C₀ ≤ 1 mg/L, while 25% degradation was obtained at C₀ ≤ 10 mg/L under visible light.
 365 Fused rings of estrogens broke down into small molecules as byproducts which were

366 mineralized through the removal of TOC. While under UV light, 30% and 20%
367 mineralization was obtained for the E2 and EE2 respectively (@C₀ 10 mg/L) while in
368 visible light, 25%, 10% mineralization was obtained for E2 and EE2, respectively. The
369 estrogen receptor-positive MCF-7 breast cancer cells were used to determine the
370 estrogenicity through an E-screen assay. Analysis revealed that organic compounds
371 obtained after degradation were sorbed on nanocomposite. Desorption was carried out
372 following recalcination (@ 600 °C) for 1 hour, leading to the regeneration of nanocomposite
373 active sites. This enabled the reuse of nanocatalyst for 3 cycles under visible light without
374 visible light activity loss (Menon, George, Tatiparti, & Mukherji, 2021). In the latest report,
375 chemically activated g-C₃N₄ (graphitic carbon nitride) nanorods were utilized to degrade
376 EE2 in visible light in water and real hospital wastewater. Pure and hydrothermally
377 activated g-C₃N₄ photocatalysts were used, and activated g-C₃N₄ was found with an
378 enhanced surface area due to intra-layer delamination and depolymerization. Activated
379 g-C₃N₄ showed potential degradation of EE2 in water (96% @ 45 min) and hospital
380 wastewater (HWW 99.09%, 30 min) in the presence of visible light. This research
381 provides insight into the development of hetero-junction visible light photocatalyst. The
382 estrogenicity of treated water samples was evaluated through the performance of the
383 Yeast Estrogen Screen. Potential removal of estrogenic activity of EE2 was attained at
384 almost 99% (<45 min, 3 mg/l) under visible light-activated g-C₃N₄. After 5 cycles, the loss
385 of degradation potential (10%) was observed (Kumar, Avisar, Betzalel, & Mamane, 2020).
386 Zinc oxide ZnO has also been used to degrade estrogens (Sornalingam et al., 2016).
387 After titanium dioxide, the ZnO is the most common photocatalyst. The activity of ZnO
388 decreases due to dissolution and pollutes the water by releasing free Zn ions. Bayode,

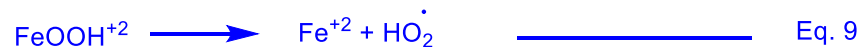
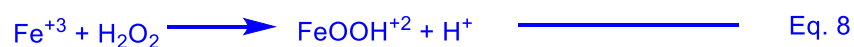
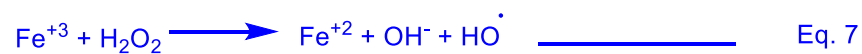
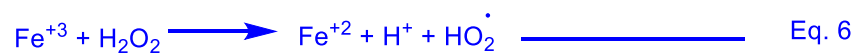
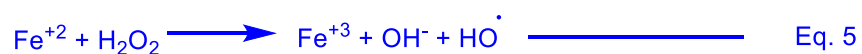
389 Vieira, et al. (2021) reported the potential of p-n heterostructured ZnO/GO photocatalyst
390 supported on clay to remove various steroids as E1, E2, E3, and EE2. The catalyst was
391 fabricated through a microwave-assisted strategy. The existence of carbon interlayer
392 (from *Carica papaya* seeds) and graphene oxide were important for the efficacy of
393 photocatalyst. In a single solute system, the removal of estrogen was found >89 and as
394 high as 98%, while in a mixed system, it was >91 and as high as 98 %. In real wastewater,
395 the efficiency of photocatalyst ZnO/GO/C was found 65.9, 66.7, 62.6 and 77.7% for the
396 removal of E1, E2, E3 and EE2, respectively. After 3 cycles of reuse, the efficacy of
397 photocatalyst was >80%. The existence of humic acid reduced the removal potential upto
398 >70% for tested estrogens, while the incorporation of 1 % hydrogen peroxide enhanced
399 the estrogens' photo-degradation up to 100% within 10 minutes. COD test revealed the
400 estrogen oxidation level after photo-degradation (51-77%). Photo-degradation was
401 mainly through hydroxyl radicals (OH·) via superoxide radical ($\cdot\text{O}_2^-$) and hole (h^+)
402 formation from the photocatalytic composite. In the ZnO and GO composite, the electron-
403 hole pairs are formed under light. The holes and electrons migrate between ZnO and GO
404 via a carbon interlayer (Figure 4). The carbon acts by hindering the quick recombination
405 of electron-hole pairs through the transference of electrons between ZnO and carbon,
406 which enhances the absorption of visible light and is responsible for increasing the
407 photocatalytic potential of composite catalyst (Sun et al., 2016). A toxicity test was
408 checked through *ceriodaphnia silvestrii*, and it was found that mild toxicity was found
409 (LC_{50} = ca. 0.89 mg/L) (Bayode, Vieira, et al., 2021). Another efficient photocatalyst
410 (Fe@PSK@GO) made from Kaolinite clay, Fe_2O_3 , and graphene oxide was used to
411 remove various estrogens (E1, E2, E3, EE2) under visible light. The synthesized catalyst

412 oxidized the estrogens at >80% in water. Mineralization of estrogens was obtained in the
413 range of 58-73%. The iron oxide presence in composite enhanced the photocatalytic
414 potential, and further incorporation of graphene oxide increased the efficiency of
415 photocatalyst. The incorporation of a carbon source doubled this potential. The holes and
416 superoxide radicals were the main sources of photocatalytic efficiency of the
417 nanocomposite. Toxicity assay showed that treated water proved safe for the use of
418 human beings. The catalyst could be recycled three times without potential loss (Bayode,
419 dos Santos, et al., 2021). CdS is a known photocatalyst that is responsive to light in a
420 good way and has been widely used (Zhao et al., 2020). The novel CdS nanorod/TiO₂
421 (B) nanobelt nanocomposite, through the hydrothermal process, was synthesized to
422 remove EE2 in visible light. TiO₂-B type was crosslinked with the Cds nanorods to
423 generate heterojunction with good removal (92%) than pure Cs nanorods within 120
424 minutes (Luo et al., 2019). Some commonly used photocatalysts for removing various
425 estrogens have been presented in Table 2.

426 **4.2. Fenton, Fenton like, and Photo Fenton degradation of estrogenic compounds**

427 Fenton's approach is promising to degrade organic contaminants. Iron-based salts are
428 utilized in this process to decompose hydrogen peroxide into hydroxyl radicals. Initially,
429 the hydroxyl radicals and Fe⁺³ ions are generated through iron salts (Fe⁺²) and hydrogen
430 peroxide. In the next step, the Fe⁺³ reacts with other reaction intermediates and forms
431 Fe⁺², and this step is enhanced in the presence of light. This process has the advantage
432 of the involvement of cost-friendly reagents (Eq. 5 & 6). The degradation potential of the
433 Fenton process depends on the concentration of the catalyst, temperature, and pH of the
434 reaction medium. In 2005, the first time degradation of estrogens was reported using UV-

435 Vis/Fe(III)/H₂O₂. Degradation efficiency was found dependent on the pH and
 436 concentration of Fe (III) and H₂O₂. E1 degradation was almost 98% after 160 min (Feng,
 437 Ding, Tu, Wu, & Deng, 2005). Iron (III) salts can replace iron (II) salts. This is known as
 438 the Fenton-like process, where Fe⁺³ and hydrogen peroxides are utilized. Hydroxyl
 439 radicals are produced due to the decomposition of H₂O₂, and Fe⁺³ is converted to Fe⁺²
 440 (Eq. 7) Fenton-like process also generates peroxy radicals (Eq 8, 9). The Fenton reaction
 441 performs efficiently at low pH values (2-5) to treat wastewater pollutants.



442
 443 Conventional Fenton reaction has limitations like narrow pH range, catalyst recycling, etc.
 444 Yaping & Jiangyong (2008) synthesized a heterogenous Photo-Fenton catalyst, α-
 445 FeOOH coated resin. This catalyst degraded E2 efficiently in the presence of H₂O₂ under
 446 UV light over a wide range of pH (3-11) (Yaping & Jiangyong, 2008). Various iron-based
 447 heterogeneous Fenton catalysts, such as supported and unsupported iron catalysts of
 448 zero-valent iron, and multi-metallic systems iron oxides, proved efficient for the
 449 degradation of various organic contaminants. The Fenton process is enhanced by
 450 electricity, UV, Visible light, and complexing agents (Scaria, Gopinath, & Nidheesh, 2021).
 451 A heterogeneous catalyst nano-zero valent iron Fenton system loaded with citrate ligand
 452 (C-NZVI) was fabricated to degrade estrogens. Citrate contains the hydroxyl fragments
 453 and covers the surface. This can overcome the defects of nZVI and deal with acid

454 corrosion, oxidation, and agglomeration. Hydroxyl and superoxides were formed and
455 degraded the E2. Ligand performed as the ion transport channel and enhanced the
456 excitation of formed radicals (hydroxyl and superoxide) and singlet oxygen (Figure 5)
457 (Yang et al., 2021). Catalyst efficiently degraded the E2, with the degradation potential
458 increasing by 4.59 folds (Yang et al., 2021). Mechanism of degradation of E2 exhibit that
459 four rings may be destroyed alternatively starting from the benzene ring. Silva and
460 coworkers carried out a study to degrade the various estrogens (E1, E2, E3, EE2) from
461 real wastewater treatment plant (WWTP) effluents in the presence of white LED Photo-
462 Fenton reaction. For the photo-Fenton response, the Fe^{+2} (Ferrous sulfate heptahydrate)
463 and hydrogen peroxide were used at various concentrations, and white LED light
464 ($\lambda = 455 \text{ nm}$) was used for irradiation purposes. To quantify the estrogenic activity, the
465 yeast estrogenic screen test was used. Under optimal conditions, the Photo-Fenton
466 process resulted in 62% removal of estrogenic activity and 59% mineralization. Treated
467 water samples were not found toxic to *Aliivibrio fischeri*, and almost 80% of estrogens
468 were removed. Thus results of the study showed that LED irradiated photo Fenton
469 reaction can be used to treat wastewater possessing estrogens (Silva et al., 2021). Some
470 commonly used Fenton, Fenton like, and Photo Fenton processes for degradation of
471 estrogenic compounds have presented in Table 3.

472 **4.3. Electro-Fenton degradation of estrogenic compounds**

473 The electro-Fenton process is based on the generation of H_2O_2 on the cathode, which
474 produces hydroxyl radicals after dissociation in the Fe^{+2} catalyst via the Fenton reaction.
475 This process occurs in acidic media due to the availability of more H^+ ions to produce
476 maximum hydrogen peroxide and avoid hydrogen peroxide decomposition into oxygen

477 and water (Poza-Nogueiras, Rosales, Pazos, & Sanroman, 2018). The electro-Fenton
478 process is a highly reported reaction for the degradation of various environmental
479 pollutants from an aqueous solution, but its applications are not widely reported for
480 estrogens degradation. In 2012, the Electro-Fenton process was reported to degrade
481 estrogen hormones. EF was used to degrade E2 in an aqueous-acetonitrile mixture.
482 Platinum anode and carbon felt cathode was used. The initial concentration effects of E2
483 were observed where 1, 5, and 10 mg/L of estrogen were completely degraded in 25, 30,
484 and 40 min, respectively. The efficacy of the Electro-Fenton process was confirmed for
485 high-rate degradation of E2 in an aqueous-acetonitrile mixture compared to the methanol
486 (co-solvent). The 100% degradation of 5 mg/L of E2 was optimized (@ 0.2mM Fe⁺² and
487 200 mA). In this study, the toxicity issue of obtained byproducts was not addressed (Naimi
488 & Bellakhal, 2012).

489 **4.4. Ozonation of estrogenic compounds**

490 Ozone can react with organic pollutants due to its strong oxidation potential. This
491 technique is highly employed for wastewater treatment. The ozonation process can be
492 coupled with UV, H₂O₂ or photocatalysis to enhance the potential to degrade organic
493 molecules. This may be attributed to an increase in the number of reactive radicals.
494 Hydrogen peroxide can be cost-friendly and highly soluble in water (Vieira, de Farias,
495 Spaolonzi, da Silva, & Vieira, 2021). Maniero et al. reported the degradation of 17β-
496 estradiol and 17α-ethinylestradiol in the presence of ozonation and O₃/H₂O₂ processes
497 under different pHs (3, 7 and 11). Through the YES assay, the estrogenic activity was
498 evaluated. Ozonation and O₃/H₂O₂ were found effective in removing both estrogens in an
499 aqueous medium. 17β-estradiol and 17α-ethinylestradiol were removed 100% and 99.5%

500 under pH 3, respectively. Estrogenic activity was removed completely in Ph for ozonation
501 and O₃/H₂O₂ (Guedes Maniero, Maia Bila, & Dezotti, 2008). According to a report, various
502 estrogens (E1, E2, E3 and EE2) were degraded in different oxidation processes as O₃,
503 O₃/UV and O₃/UV/H₂O₂. Significant degradation was obtained in both O₃/UV and
504 O₃/UV/H₂O₂, but it was comparatively low than O₃. And this may be attributed to presence
505 of hydroxyl radical scavengers as carbonates and bicarbonate ions (Pešoutová, Stříteský,
506 & Hlavínek, 2014). The clay was used as a catalyst in the ozonation process to degrade
507 EE2 in an aqueous solution. Acid-activated bentonites and ion-exchanged
508 montmorillonite (NaMt and Fe(II)Mt) exhibited catalytic potential during the ozonation of
509 EE2. In the presence of Fe(II)Mt, the degradation of EE2 was obtained at almost 96%
510 after one minute of ozonation. And after 15 minutes, under the same conditions, almost
511 99.9% degradation of EE2 was obtained. Acid-activated bentonites showed great
512 potential for providing total mineralization within a short time of ozonation. The catalytic
513 potential of clay correlates with its surface basic character and hydrophilicity. This study
514 provides insights into soil behavior in the oxidative degradation of organic compounds
515 (Boudissa et al., 2020). Perondi and his colleagues demonstrated the removal of E2
516 present in surface water under UVC (254 nm) light in the presence of various oxidants as
517 O₃, H₂O₂ and TiO₂. Pseudo-first order kinetics were followed:
518 $k_{UV/O_3} > k_{UV/TiO_2} > k_{UV/H_2O_2} > k_{UV}$. Efficient removal was obtained by photolysis (UV),
519 photo-peroxidation (UV/H₂O₂), photocatalysis (UV/TiO₂), and photo-ozonation (UV/O₃).
520 Eight transformation products were obtained. These may be obtained by ring-opening of
521 E2 and hydroxyl radical (Perondi et al., 2020). Table 4 presents various ozone-based
522 processes for the removal of estrogens.

523 **5. Biological methods for removal of estrogenic compounds**

524 **5.1. Microbial degradation of estrogenic compounds**

525 Biodegradation implicates the utilization of microbial strains like bacteria and fungi for the
526 degradation and transformation of endocrine-disrupting chemicals and many other
527 environmental contaminants. It has been considered a leading removal mechanism that
528 affects the fate of endocrine-disrupting chemicals in the environmental matrices. Plenty
529 of studies in the recent decade has investigated the biodegradation of various EDCs
530 through diverse microorganisms (Husain and Qayyum, 2012; Zhang et al., 2016;
531 Ahuactzin-Pérez et al., 2018). Lignin-degrading organisms, e.g., white-rot
532 basidiomycetes, have gained attention as efficient candidates for degrading emergent
533 micropollutants from the environment. Cabana and coworkers (2007) reported the WRF's
534 capability for eliminating EDCs in different environmental compartments and
535 recommended establishing reliable and robust biotechnological processes to treat EDC-
536 loaded water and wastewater systems. In another study, Cajthaml (2015) substantiated
537 the potentiality of ligninolytic fungi in degrading and transforming EDCs using a versatile
538 lignin-modification enzymatic system and cytochrome P-450. Ligninolytic fungi-assisted
539 EDCs degradation is likely via micropollutants polymerization or degradation by enzymes.
540 It is worth noting that ligninolytic fungi are among the few microorganisms that efficiently
541 degrade PCBs and EE2 (Cajthaml, 2015).

542 Ligninolytic bacteria are robust microbes for EDC degradation due to their ability to
543 secrete lignin-degrading enzymes, including manganese peroxidase and laccase. Due to
544 high tolerance in diversified environmental matrices and genomic maneuverability,
545 bacteria possess a profound ability to catalyze EDCs biodegradation. In addition,

546 integration of bacteria with physical approaches, i.e., adsorption, offers unique
547 advantages with the excellent aptitude to eliminate EDCs in aqueous environments
548 (Zhang et al., 2016). Nonetheless, the introduction of harmful microbes into the
549 surroundings is the primary concern of adopting biodegradation approaches for removing
550 EDCs, which may lead to antibiotics and multiple drug emergence issues. Thus, the
551 enzymatic process involving extracellular ligninolytic enzymes is likely a more appropriate
552 substitute.

553 **5.2 Removal of estrogenic compounds by ligninolytic enzymes**

554 The environmental risk of EDC exposure to humans and the inadequate efficiency of
555 classical treatment strategies for complete EDC removal have rekindled the interest of
556 researchers in exploring alternative options for EDC removal in wastewater. The
557 exploitation of ligninolytic oxidative enzyme-based treatment processes has lately
558 garnered much attention as a promising and eco-friendly alternative for eliminating EDCs.
559 Ligninolytic enzymes, including manganese peroxidase (MnP), Lignin peroxidase (LiP)
560 laccase, and versatile peroxidase (VP), have recently been used to degrade EDCs in
561 water matrices efficiently. Becker et al. (2017) investigated the biocatalytic transformation
562 of endocrine disruptors in real wastewater and artificial mixtures by laccases derived from
563 *Myceliophthora thermophila* and *Trametes versicolor*. Results revealed the feasibility
564 EDCs and hormones degradation even at a very low concentration of laccases.
565 Adsorption onto support matrices was the effective elimination mechanism in the artificial
566 mixtures. In binary solutions, immobilized biocatalyst presented high performance for
567 EDCs removal in the initial 6 h (87% for *M. thermophila* and 83% for *T. versicolor*);
568 however, adsorption superimposed this degradation during the experimental course (99%

569 after 24 h). A comparable removal pattern was detected in real wastewater but low
570 adsorption efficiency and enzyme activity. The immobilized laccase-based biocatalytic
571 system displayed the maximum removal efficiency of androgenic (99% removal after 6 h)
572 and estrogenic (82% after 24 h) activity. Efficient degradation of EE2 by *Lentinula*
573 *edodes* was attributed to the laccase since the induced production of laccase increased
574 the degradation efficacy of tested pollutants from 50% to 80% (Eldridge et al., 2017).

575 A commercial laccase from *M. thermophila* was used to design an enzymatic membrane
576 reactor to remove E1 and E2 continuously. The effects of hydraulic residence duration (1-
577 4 h), oxygen frequency supply, and laccase activity (500-1,000 U/L) were also assessed.
578 Interestingly, high degradation efficiencies between 64-100% were achieved with a
579 corresponding degradation rate of 0.96-2.92 mg/(L·h). The estrogenic activity of effluent
580 was diminished to over 95%, proving the practicality of laccase-assisted bioreactor for
581 estrogenic compounds removal (Lloret et al., 2012). To improve the enzyme stability and
582 degradation efficiency in a packed bed reactor, Eupergit supports immobilized laccase
583 was operated in a fluidized bed reactor to remove estrogens. In addition to a significant
584 improvement in biocatalyst stability (over 16 days), removal levels increased to 76 and
585 90%. The estrogenic activity of the effluent was also decreased up to 90%. In conclusion,
586 immobilized laccase-based technology appears a promising tool for extending the
587 exploitation of laccases in environmental bioremediation.

588 LiP is an emerging enzyme class produced by various fungal strains, i.e., *Phanerochaete*
589 *chrysosporium*. These fungi also have veratryl alcohol (VA) as a secondary metabolite
590 that regulates the functions of LiP. Mao et al. (2010) reported the effective removal and
591 transformation of E2 via LiP-mediated catalytic reactions, which can be substantially

592 boosted using VA. They evaluated enzyme reaction kinetics and LiP activity in the
593 systems with or without VA. Experimental findings substantiated higher removal and
594 transformation of E2 by VA through two effects, including mitigation of LiP inactivation
595 and modification of enzymatic catalytic kinetics. MnP is another important lignin-modifying
596 enzyme for the removal and degradation of EDCs. Tamagawa et al. (2006) treated
597 steroidal estrone with *P. sordida* YK-624 in high-carbon and low-nitrogen culture medium
598 ligninolytic conditions. After 5 days of treatment, E1 concentration was reduced by 98%.
599 The disappearance of E1 was ascribed to the activities of ligninolytic enzymes MnP and
600 laccase, which were detected during the treatment. HPLC analysis revealed the complete
601 disappearance of E₁ in the reaction following 1 h exposure to either laccase or MnP. The
602 yeast two-hybrid assay system evidenced that the estrogenic activity of E₁ was also
603 completely eliminated by 2 h of enzymatic treatment.

604 VP has emerged as a promising biocatalyst to eliminate EDCs in the environment. VP
605 integrates the substrate preference of LiP and MnP (Ruiz-Duenas et al., 2009). The
606 catalytic cycle of VP relies on the concerted action of various compounds, like cofactors,
607 substrates, and mediating agents that initiate and take part in the biocatalytic cycle.
608 Taboada-Puig et al. (2011) formulated cross-linked enzyme aggregates (CLEAs) from
609 glucose oxidase and VP to evaluate their potential to remove different EDCs, triclosan,
610 NP, BPA, E2 and EE2. The as-synthesized CLEAs could remove all tested pollutants
611 except for triclosan, and the elimination of estrogenicity was over 55% for all EDCs.
612 Therefore, incorporating glucose oxidase with any other ligninolytic enzyme is suitable to
613 remove EDCs. In another study, the same group used VP-induced Mn³⁺-malonate
614 oxidant for continuous elimination of triclosan, BPA, E1, E2, and EE2 from real and

615 synthetic wastewaters in a two-stage (TS) system and observed a degradation efficiency
616 in the range of 28 to 58 $\mu\text{g/L min}$, with negligible enzyme deactivation. Results revealed
617 a 14-fold improvement in the VP-assisted removal efficiency of EDC in a TS system than
618 enzymatic membrane reactor (EMR). Moreover, the functional challenges faced during
619 EMR system were also circumvented. VP-coupled TS enzymatic system presented a
620 100% degradation rate for all studied EDCs, thus substantiating the feasibility of this
621 system for efficiently eliminating EDC even at high concentrations (Taboada-Puig et al.,
622 2015).

623 **6. Transformation mechanisms and products**

624 The disappearance of the parent molecule of estrogen is used to assess the removal.
625 Estrogen-derived molecules have a significant impact on the environment. Therefore, it
626 is important to understand the removal mechanism of estrogens and their formed by-
627 products and intermediates. Determination of degradation routes identifies the hurdles of
628 the process (Rathner, Petz, Tasnádi, Koller, & Ribitsch, 2017). Evaluation of final
629 products may facilitate an increase in the purity of the final stream of products through an
630 appropriate separation approach. Conclusively, the operational process and removal
631 potential of estrogens may be enhanced. Different degradation methods provide various
632 end products. Photocatalytic degradation of E2 and EE2 through $\text{TiO}_2\text{-ZnO}$
633 nanocomposite under UV and visible irradiations was determined, and both estrogens
634 were degraded. Fused estrogens rings were broken down into simple small compounds
635 that were further mineralized by removing TOC (Total organic carbon) (Menon et al.,
636 2021). Bisphenol A (BPA) was degraded using $\text{S}_2\text{O}_8^{2-}/\text{UV-C}$ and $\text{H}_2\text{O}_2/\text{UV-C}$ treatments.
637 LC-MS/MS was used to evaluate the degradation products. A bioassay revealed that

638 obtained by-products after degradation were more toxic and harmful than BPA at the
639 initial point of H₂O₂/UV-C. After degradation, various ring-opening products like oxalic
640 acid, succinic acid, fumaric acid, 3-methyl benzoic acid and 4-sulfobenzoic acid were
641 obtained after degradation (Olmez-Hanci et al., 2015). Laccase and peroxidases oxidized
642 the phenolic group and produced phenoxy radicals (Beck et al., 2018). Sometimes, the
643 phenoxy radicals combine to create a dimer of the initial molecule. In the next step, these
644 formed dimers may get oxidized again through oxidoreductase enzymes and form
645 trimmers up to oligomers (Xia et al., 2014). At the initial stage of the reaction, the
646 production of phenoxy radicals demonstrates the disappearance of the molecule
647 monomer. The radical oxidation mechanism is involved in the enzymatic transformation
648 of 17 β -estradiol and this leads to the formation of dimers and trimmers. Xia and
649 colleagues reported the removal of 17 β -estradiol by using the laccase enzyme in the
650 existence of organic matter. This happened because of electron resonance on phenoxide
651 oxygen to benzene and the production of promising active sites on the estrogenic
652 molecule (Xia et al., 2014). The mass spectrum showed the m/z peak at 271, showing
653 the residue of starting of 17 β -estradiol. The degradation pathway of 17a-ethinylestradiol
654 showed the same mechanism as that of 17 β -estradiol. Laccase was used to remove the
655 17a-ethinylestradiol and the main product of the reaction was found to dimer of 17a-
656 ethinylestradiol with m/z peak at 591 after 60 minutes of reaction. However, at later stages
657 of reaction the concentration of dimer decreased. This was because the dimer oxidation
658 directly through laccase and indirect oxidation may occur with the help of monomer
659 radicals. This may lead to 17a-ethinylestradiol oligomeric derivatives (Tanaka et al.,
660 2009). Golveia et al. (2018) used laccase for the removal of 17a-ethinylestradiol.

661 Mass spectrum results combined with the electrospray ionization and detector (time of
662 flight) exhibited a dimer as the main product of the transformation reaction. While mass
663 spectrum obtained after 6 h has only one peak at m/z of 311, which shows the presence
664 of a new degradation product. This new product was produced because of hydroxyl
665 radical attack on 17 α -ethinylestradiol, leading to dehydration and attack of hydroxyl
666 radical, further providing 17 α -ethinylestradiol semiquinone (Golveia et al., 2018). Trudeau
667 et al. (2011) demonstrated the almost complete transformation of E1 into lumiestrone
668 under illumination within 5 minutes. More radiation exposure leads to 100% E1
669 degradation (Trudeau et al., 2011). Mboula et al. (2015) carried out the photocatalytic
670 degradation of E2 in solar light by employing various titanium dioxide catalysts. The
671 obtained photo-products were examined. Intermediates possessed the phenolic moiety,
672 which caused the estrogenic activity of the solution. Incorporation of hydroxyl moiety was
673 suggested as a mass difference of 16 m/z was found of parent and intermediate ion.
674 Intermediate having a mass of 288 g/mol was ascribed as 2-hydroxyestradiol or its
675 resonating structure (10 ϵ -17 β -dihydroxy-1,4-estradien-3-one) (Mboula et al., 2015).

676 **7. Toxicity and estrogenic activity assessment of transformed byproducts**

677 AOPs are potential processes for removing a wide range of environmental pollutants from
678 water bodies. But due to high operational costs and various studies, different byproducts
679 are generated from parent compound degradation. The formed b-products are sometimes
680 more toxic than parent molecules. Identifying and quantifying byproducts is still a
681 challenge due to their low concentration and solution complexity (H.-C. Chen, Kuo, &
682 Ding, 2009). Analysis of byproducts is necessary for the safety of living beings. Different
683 analytical techniques and methods have been developed to identify photo-products

684 qualitatively and quantitatively. Hyphenated techniques as GC/MS, LC/MS, LC/MS/MS,
685 GC/MS/MS are employed to detect pollutants and their degradation byproducts. Several
686 studies focused on photo-degradation of parent molecules, which is not enough to find
687 the estrogenic effect of photo-products. Few intermediates show high estrogenicity
688 compared to their parent molecules (Mboula et al., 2015), which forced them to evaluate
689 their biological potential. It is also necessary to assess the estrogenicity of aquatic
690 systems to check the synergistic effects caused by different photo-transformed
691 byproducts (Yan & Song, 2014). Various methods are used to determine estrogenic
692 activity and Yeast estrogen screen assay (Yes) is the most commonly used technique to
693 measure the potency of estrogen hormones. Photocatalytic activity of TiO₂ removed the
694 estrogenic potential of E1, E2 and E3 in 30 minutes, while only 37% removal was
695 observed in the photolysis process. Removal of estrogenic activity followed the pseudo-
696 first-order kinetics for photolysis and photocatalysis with the constant rate values of
697 0.0069/min and 0.0461/min, respectively (W. Zhang, Li, Wu, & Hu, 2012). Whidbey and
698 his colleagues carried out the YES assay and photoproducts of E2 and EE2 showed no
699 estrogenic activity under indirect photolysis. At the same time, E1 degradation formed the
700 limestone, which showed moderate activity under direct photolysis. Degradation of the
701 phenolic group of hormones is the cause of loss of activity (Whidbey et al., 2012).
702 Photoproducts of E2 showed no estrogenic activity in yeast estrogen-based assay (Ohko
703 et al., 2002). Dimitroula et al. (2012) determined the removal of estrogenicity of a mixture
704 of phenol, bisphenol-A and EE2 spiked in secondary treated STP effluent and observed
705 loss inactivity (30%). Degradation of EE2 in wastewater exhibited 27% reduction in
706 estrogenicity evaluated by YES assay. Degradation of E1, E2 and EE2 in wastewater

707 provided 30% removal of estrogenicity even parent molecule has completely removed in
708 120 minutes, showing that few intermediates generated were also estrogenic (Frontistis
709 et al., 2012). Estrogenicity of E2 photodegraded products was evaluated and it was found
710 that estrogenic activity was not removed. Sample of reaction taken at 15 min exhibited
711 high estrogenic effect in comparison of initial E2 solution. This may be due to the
712 formation of byproducts that were highly estrogenic compared to E2 or this may also be
713 due to the combined estrogenic potential of E2 and its photo-degraded products (Mboula
714 et al., 2015).

715 **8. Way forward and future perspective**

716 The presence of pharmaceuticals in our water resources poses adverse effects on the
717 environment. Treated wastewater is used for irrigation worldwide, but if it is not
718 appropriately treated, the risk of estrogenic contamination increases. This study focuses
719 on removing estrogens from water sources using various physical, chemical, and
720 biological approaches. We have established multiple areas in which potential
721 interventions may impact change. Firstly, there is a need to develop drugs based on green
722 chemistry to be quickly metabolized by the body and undergo facile environmental
723 biodegradation. Previously designed treatment plants had low capacities, so there is a
724 need for their upgradation by integrating them with novel structures, disposal approaches,
725 and monitoring systems, keeping in view the demands and focusing on the recovery of
726 resources. The literature shows that treatment plants based on chemical (photocatalysis,
727 Fenton, Fenton-like and Photo Fenton, electro-Fenton and ozonation-based degradation)
728 and biological (microbial and enzymatic degradation) approaches can be potent cost-
729 friendly for removal of estrogenic pollutants. Secondly, there is a need to properly monitor

730 and dispose of the usage of estrogenic drugs in humans and animals, because
731 sometimes drugs are disposed of intentionally in flushes and enter the sewer system.
732 Industries involved in production and health care units that dispose of their wastes in the
733 ecosystem are also significant sources of estrogenic waste.

734 The practices can be collecting expired and unused estrogenic products and regulating
735 disposal norms. Previous developments regarding biosensors for estrogen detections
736 have been reported. Electrochemical, cell-based, optical, and organism-based
737 biosensors have been introduced for sensing estrogens in environmental matrices (Lu,
738 Sun, & Sun, 2020). Dehalogenase enzymes have been an important tool for sensing
739 halogenated molecules. Various biosensors based on dehalogenation enzymes have
740 been reviewed in detail to sense halogenated molecules (Gul et al., 2021). Gul and
741 colleagues also developed a 3-dimensional platform integrated with smart phone ambient
742 light sensor to detect halogen-based pollutants (Gul et al., 2021). Therefore, the
743 attachment of different sensors with treatment plants can be helpful to find these
744 compounds.

745 Thirdly, the provision of guidelines about the allowed limit of estrogens in wastewater
746 treatment plant effluents may facilitate the researchers for practical applications of
747 chemical and biological-based treatments and their potential to degrade these effluents
748 in useful products. Finally, attention should be given to using microbes with hybrid
749 treatment systems, keeping in more capacity of microbes to bear environmental stress.
750 This review proposes that hybrid technologies can be a good choice for wastewater
751 treatment for removing estrogens. These processes will help remove the drugs and
752 protect the environment from damage efficiently.

753 **9. Conclusions**

754 Endocrine disruptive chemicals (EDCs) have been known to pose potential menaces to
755 the environmental matrices and living entities. Considering the complexity and
756 concentrations of estrogenic compounds, the development of appropriate treatment
757 approaches is indispensable to ensure the effective elimination of disruptive endocrine
758 estrogens in water resources. Various physical, biological, and chemical methods have
759 been applied following the requirement and efficiency for removal. Physical processes,
760 including adsorption, reverse osmosis, membrane filtration, nanofiltration, and
761 ultrafiltration, have been widely adopted to remove EDCs. Applying adsorption-based
762 treatments for eliminating EDCs are mounting using many adsorbents. Nevertheless, the
763 assessment for continuous use at a large scale remains to be implemented. In addition,
764 the membrane filtration showing good efficiency is restricted to only a few membranes.
765 Additional studies are required to explore a robust and more advanced oxidation
766 treatment strategy that can contribute effectively to industrial-scale applications.
767 Exploiting ligninolytic oxidative enzyme-based treatment processes has recently gained
768 prodigious attention as a promising and eco-friendly alternative for eliminating EDCs.
769 Ligninolytic enzymes, including MnP, laccase, and VP, have efficiently degraded various
770 EDCs in water matrices. However, a relentless improvement to overcome the limitations
771 of all physicochemical techniques might outspread the EDCs removal performance,
772 treatment, and applicability.

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777 **Conflict of Interest**

778 The authors declare that they have no conflict of interest.

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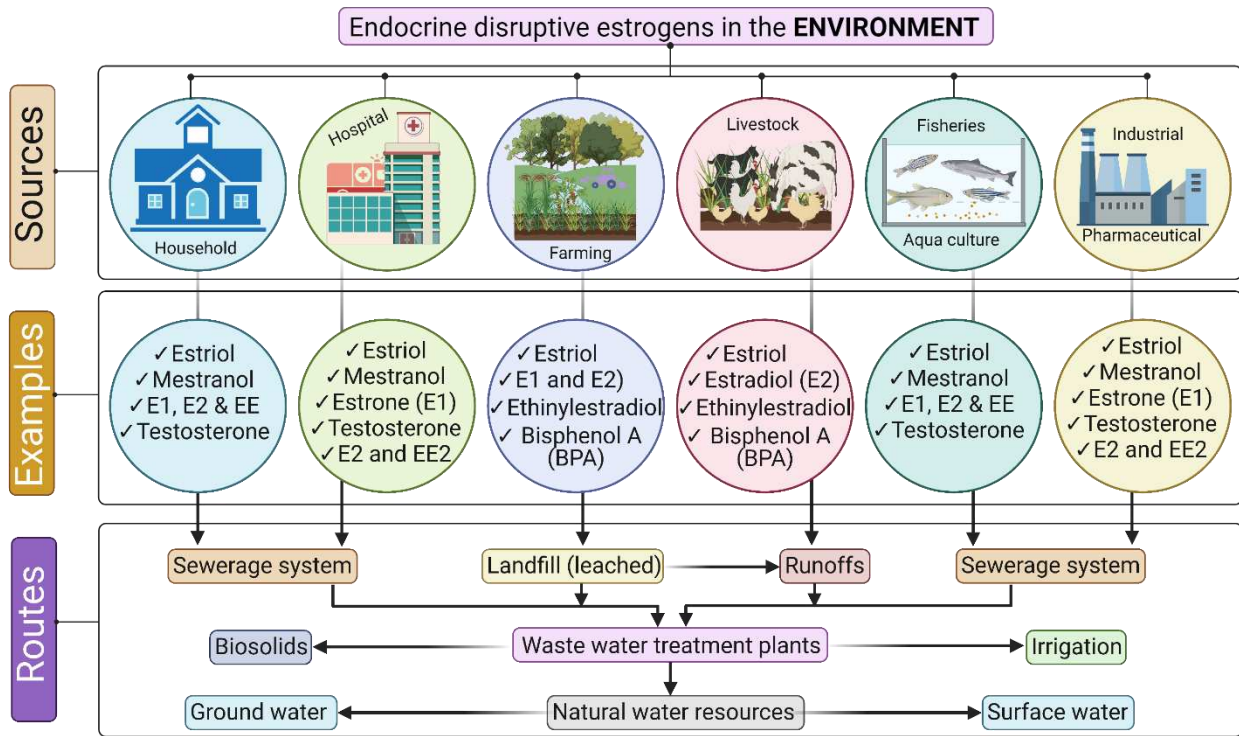
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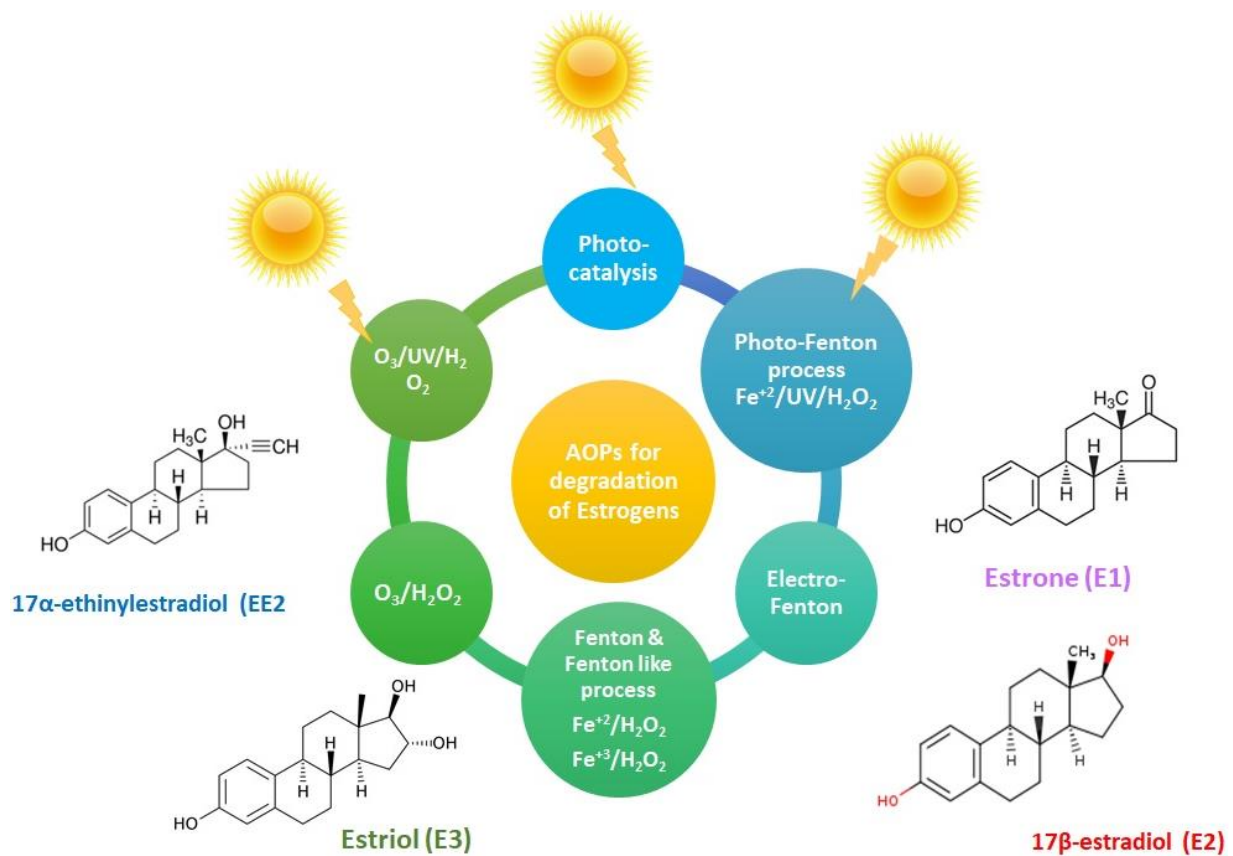
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1294 **List of Figures**



1295
 1296 **Figure 1** Routes of steroidal estrogens contamination to ground and surface water
 1297 bodies. *Created with BioRender.com and extracted under premium membership.*

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1309 **Figure 2** Advance oxidation processes for degradation of Estrogens.

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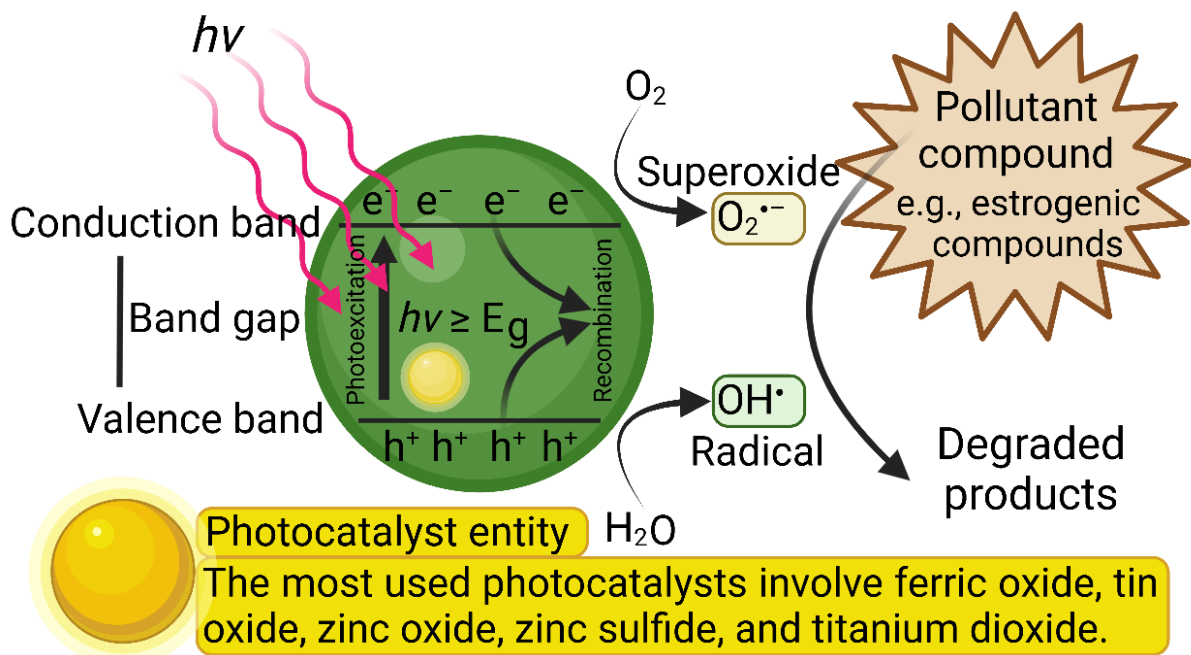
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1321 **Figure 3** Schematic illustration of photocatalysis mechanism via the generation of free
 1322 radicals and superoxide to catalytically degrade hazardous pollutants, e.g., estrogenic
 1323 compounds. *Created with BioRender.com and extracted under premium membership.*

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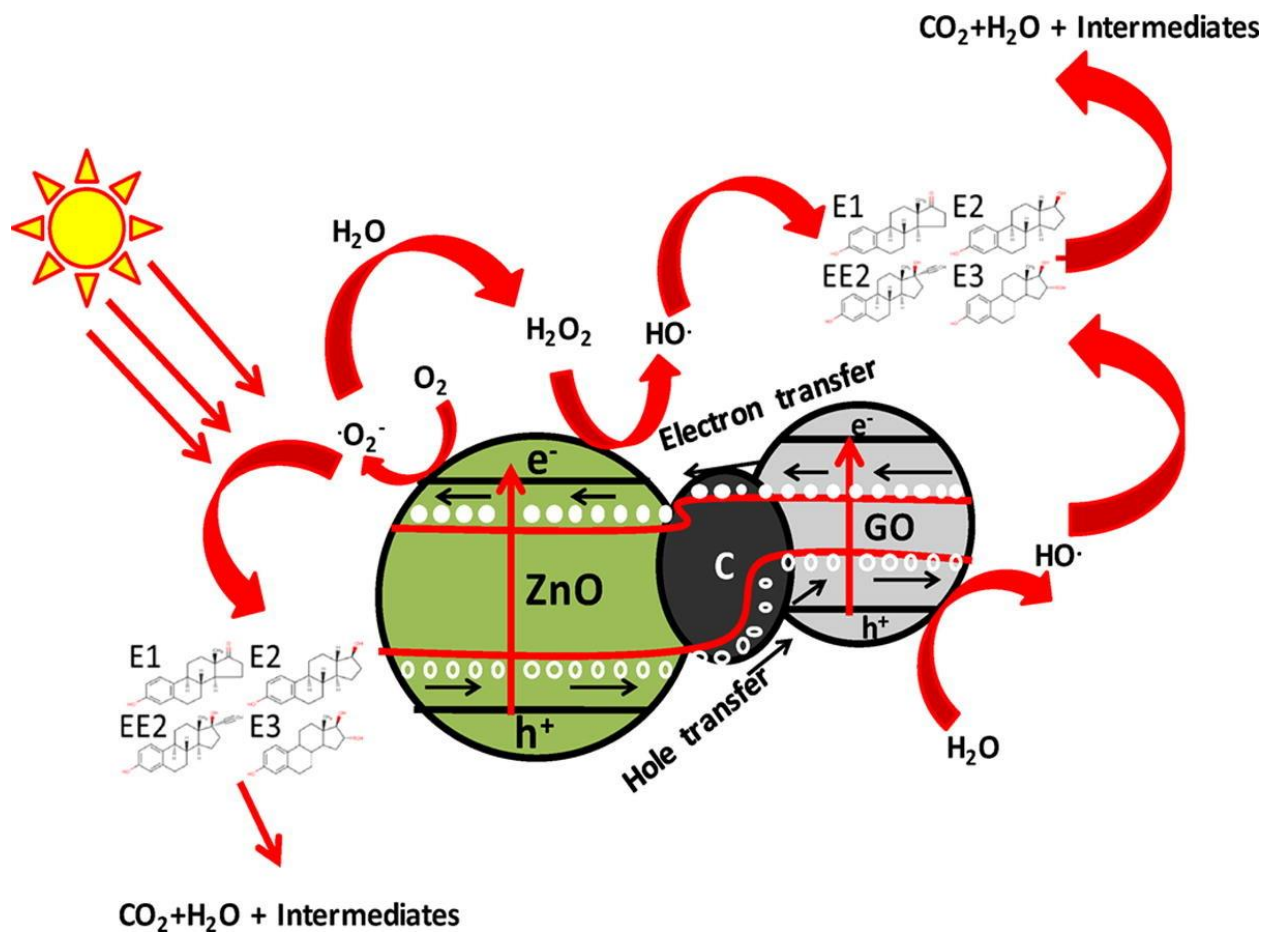
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1335 CO₂+H₂O + Intermediates

1336 **Figure 4** Proposed visible light photo-degradation mechanism of steroid estrogens over

1337 ZnO/C/GO composite. Reprinted from Ref. (Bayode, Vieira, et al., 2021) with permission

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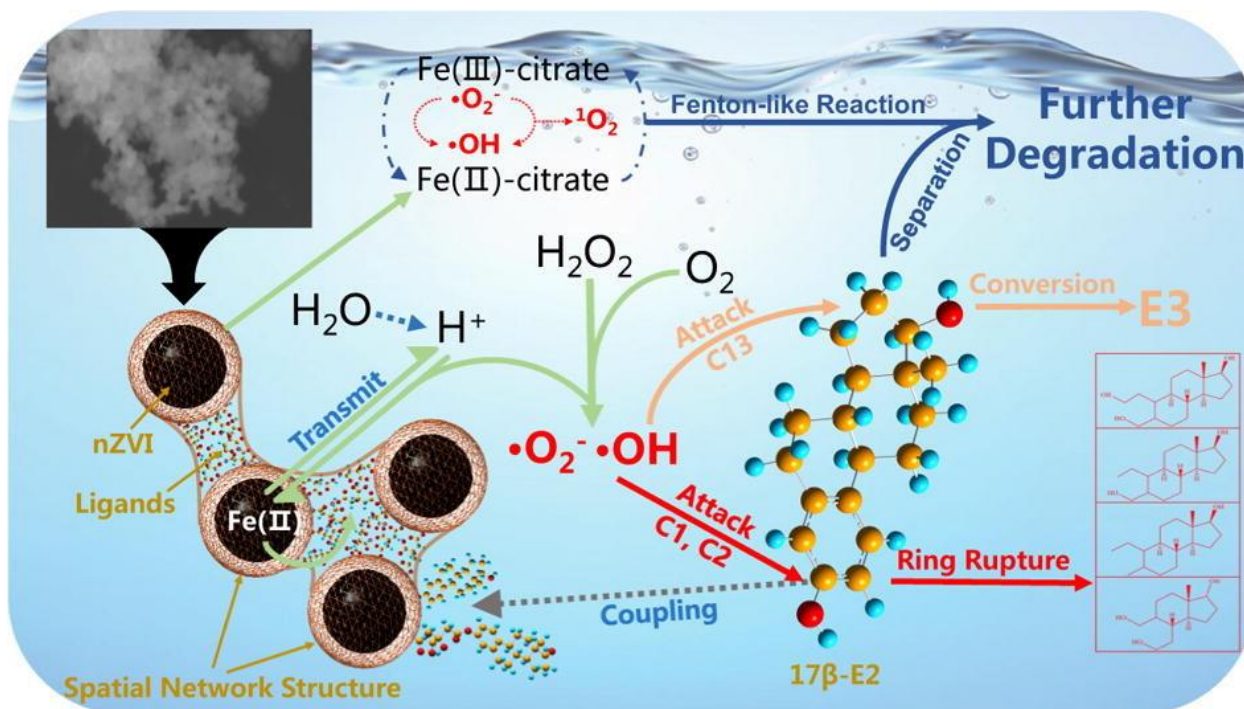
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1348 **Figure 5** Proposed degradation mechanisms of C-nZVI to 7β-E2, Reprinted from Ref.

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Table 1 Comparison of the physical, chemical, and biological based treatments.

Treatment methods	Advantages	Disadvantages	References
Physical methods			
Adsorption	Facile process, old technique, cost friendly, high surface area provision for removal	Poor selectivity, clogging issues, waste products formation, 50% removal of estrogens	(Brienza et al., 2014; Grover, Zhou, Frickers, & Readman, 2011)
Nanofiltration	Nanofiltration technique performs well in combination with other techniques, cost effective for small scale use, requires less room, facile, quick and highly effective even at high conc.	At inappropriate pressure the fouling of nanofilter membranes, maintenance cost high, removal of contaminants based on hydrophobic adsorption and size exclusion only	(Crini & Lichtfouse, 2019)
Ultrafiltration	secondary treatment method of considerable application with limited targeted estrogens removal leachate effluents, requires less space	only selected contaminants can be removed by using this method, removal based on hydrophobic adsorption only, useful in combination with other techniques like reverse osmosis	(Aziz and Ojuma et al., 2020, Patel et al., 2019, Sun et al., 2015)
Ultrasonication	Advance technique, Safe, energy saving, less formation of secondary contaminants in comparison of other tecniques, removal of contaminants even at smaller concentration, estrogen removal	Efficiency depends on specific ultrasonication conditions, efficiency of method is dependent on various factors like temp, Ph , catalyst US power and characteristics of estrogens	Pokhalekar et al., 2016, Laxmi et al., 2010, Chu et al., 2017).
Chemical Methods			

Photocatalysis	Intermediates are not formed, efficiently removes all types of estrogenic compounds	Applications of this process is limited, duration of process is long, requires high energy	(Mayer et al., 2019; Ohko et al., 2002)
Fenton, Fenton like and Photo fenton degradation	Highly used for removal of estrogenic compounds, works under wide pH range (3-11), Efficient degradation of estrogens, almost 80% degradation of estrogens	Use of hydrogen peroxide and UV light	(Yaping & Jiangyong 2008, (Silva et al., 2021).
Electro-Fenton degradation	Electro-Fenton process was confirmed for high-rate degradation of E2 in an aqueous-acetonitrile mixture compared to the methanol (co-solvent)	Process occurs in acidic media, process is less reported for degradation of estrogens,	(Poza-Nogueiras et al., 2018). (Naimi & Bellakhal, 2012).
Ozonation	Less time consuming, removal rate is 90%	High treatment cost, by-products of process may be toxic	(Li, Zhang, Liang, & Yediler, 2013; Qi et al., 2020)
Biological methods			
Microbial degradation	Environmentally safe, less harmful chemicals are involved	Depends on different factors as specific microbes required, temp. and pH	(Brienza et al., 2014)
Enzymatic degradation	Efficient for removal of estrogens	Expensive process, stability of enzyme matters a lot, selective enzymes are required for every pollutant	(Becker et al., 2017; Logar, Brouwer, Maurer, & Ort, 2014)

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Table 2 Photocatalytic degradation of estrogenic compounds.

Matrix	Compound	Catalyst	Light source	% Degradation	Reference
-	E2 EE2	TiO ₂ -ZnO nanocomposite	UV/Vis	At C ₀ <1 mg/L, 100%, at C ₀ ≤ 10 mg/L, ≥25%, under visible irradiation (240 min)	(Menon et al., 2021)
Water and Hospital wastewater (HWW)	EE2	activated g- C ₃ N ₄	Solar light	Water = 96% (45 min) HWW = 99.09% (30 min)	(Kumar et al., 2020)
Real wastewater	E1, E2, E3, EE2	p-n ZnO/GO/C	Visible light	E1 = 65.98, E2 = 66.76, E3 = 62.66, EE2 = 77.72%	(Bayode, Vieira, et al., 2021)
Real wastewater, Tap water, Rainwater	E1, E2, E3, EE2	Fe@PSK@GO	Visible light	Real water; E1 = 66%, E2 = 67% E3 = 63% EE2 = 78% Tap water; E1 = 75%, E2 = 79%, E3 = 79%, EE2 = 89% Rainwater; E1 = 77, E2 = 84, E3 = 77, EE2 = 91	(Bayode, dos Santos, et al., 2021)
Aqueous system	17α- methyltestost erone	doped TiO ₂ with Sm ³⁺ or Gd ³⁺ @0.3 and 0.5 %wt	Solar light	-	(Arévalo-Pérez et al., 2020)
-	EE2	CdS nano- rod/TiO ₂ (B) nano-belt composite	Visible light	92% (120 min)	(Luo et al., 2019)
Synthetic water	E2, EE2	TiO ₂ -P25/H ₂ O ₂	UVC	E2 = 51/32% EE2 = 48/30%	(Castellanos, Paulo Bassin, Dezotti, Boaventura, & Vilar, 2020)
Aqueous system	E2	Pd(II)- porphyrin- coated membra nes	Visible light	98%	(Lyubimenko, Gutierrez Cardenas, Turshatov, Richards, & Schäfer, 2021)
Aqueous system	E2	TiO ₂	UV	85% (60 min)	(Orozco-Hernández et al., 2019)

Synthetic solution	E3, EE2	Au-TiO ₂	11 W UVA (λ = 365 nm) or Cool White (λ > 410 nm) LEDs	Potential results	(Sornalingam et al., 2020)
-	E2	Carbon fibers/TiO ₂ /MIL-101(Fe)	Visible light	87.4% (60 min)	(Y. Zhang et al., 2021)
Blended system	E2	E2-TiO ₂ -NR photoanode	300 W-lamp equipped with UV reflector in photoelectrochemical quartz reactor	64%	(Cai et al., 2021)
-	EE2	Zn-doped titanium oxides	UVA-LED, λ = 365 nm,	-	(Engelhardt, Schmitz-Stöwe, Schwarz, & Stöwe, 2020)
Real discharged water	EE2	Persulpahte	UV/heat	Significant degradation obtained	(Chainarong, Chanut, Tunlawit, Tharisara, & Athaphon, 2020)
-	E1, EE2	Ti-0.5wt%W alloy (NT/Ti-0.5W)	UV/Vis	UV light; E1 = 54, EE2 = 66% (2 min) Visible light; E1 = 50% (1.8 min), EE2 = 50% (4.6 h)	(Escudeiro de Oliveira, Barroso, de Almeida, Moraes, & de Arruda Rodrigues, 2020)

Abbreviations: estrone = E1; 17 β -estradiol = E2; estriol = E3; 17 α -ethinylestradiol = EE2; PSK = Crushed *carica papaya* seeds, kaolinite, ZnCl₂.6H₂O, 20 mL 0.1 M NaOH

Table 3 Various advanced oxidation processes (AOPs) for removal of different estrogens.

Matrix	Compound	AOPs	Catalyst	Light source	% Degradation	Reference
WWTPE	E1, E2, E3, EE2	Photo-Fenton	Fe ⁺² / H ₂ O ₂	White LED (λ = 455 nm)	Significant degradation obtained	(Silva et al., 2021)
Wastewater	E2	Fenton	<i>cis</i> -[Fe(cbc)Cl ₂] ⁺ (Fe(L1)/ H ₂ O ₂)	-	Approx. 90%	(M. Chen et al., 2019)
-	E2	Fenton	Fe/HC- H ₂ O ₂	-	Significant degradation obtained	(Yu et al., 2019)
Aqueous system	E2	-	nZVI/PGBC/PS	-	100% (45 min) by activating sodium persulfate (PS)	(Ding et al., 2021)
Wastewater	E2	-	Fe/GBC/PS	-	100% (90 min)	(P. Zhang et al., 2019)
Wastewater	EE2	Coupled Fenton process	up-flow anaerobic sludge blanket reactors (UASB) coupled with Fe ⁺² / H ₂ O ₂	-	Significant degradation obtained	(López-Velázquez, Villanueva-Rodríguez, Mejía-González, & Herrera-López, 2020)

Abbreviations: estrone = E1; 17β-estradiol = E2; estriol = E3; 17α-ethinylestradiol = EE2; kaolinite, ZnCl₂.6H₂O, 20 mL 0.1 M NaOH; HC = hydrochar; PGBC = porous graphitized biochar

Table 4 Various ozone-based processes for degradation of estrogens.

Matrix	Compound	Catalyst	Light source	% Degradation	Reference
Aqueous medium	E2, EE2	O ₃ /H ₂ O ₂		100%, 99.5 (pH 3)	(Guedes Maniero et al., 2008)
Municipal wastewater	E1, E2, E3, EE2	O ₃ /H ₂ O ₂ /UV	(λ = 455 nm)	99.4% E1, 98.2% E2, 98.7% E3, 99.6% EE2	(Pešoutová et al., 2014)
Solvent free process	EE2	O ₃ /benonite (Fe(II)Mt,	-	96% (1 minute)	(Boudissa et al., 2020)
Surface water	E2	UVC/O ₃	UVC (λ = 254 nm)	Significant degradation obtained	(Perondi et al., 2020)

Abbreviations: estrone = E1; 17β-estradiol = E2; estriol = E3; 17α-ethinylestradiol = EE2; kaolinite, ZnCl₂.6H₂O, 20 mL 0.1 M NaOH