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The autonomic nervous system from a morphofunctional perspective : historical overview and current concepts over the last two centuries highlighting contributions from Eastern Europe

Reference:

Timmermans Jean-Pierre.- The autonomic nervous system from a morphofunctional perspective : historical overview and current concepts over the last two centuries highlighting contributions from Eastern Europe

The anatomical record: advances in integrative anatomy and evolutionary biology - ISSN 1932-8494 - Hoboken, Wiley, (2023), p. 1-8

Full text (Publisher's DOI): https://doi.org/10.1002/AR.25169

To cite this reference: https://hdl.handle.net/10067/1942530151162165141

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The Anatomical Record

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Journal:	Anatomical Record
Manuscript ID	AR-SI-Auto-22-0456
Wiley - Manuscript type:	Introduction
Date Submitted by the Author:	24-Nov-2022
Complete List of Authors:	Timmermans, Jean-Pierre; Universiteit Antwerpen Laboratorium voor Celbiologie en Histologie
Keywords:	Eastern Europe, Autonomic nervous system, enteric nervous system, gut, heart



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Special issue intro/commentary

The autonomic nervous system from a morphofunctional perspective: historical overview and current concepts over the last two centuries highlighting contributions from Eastern Europe.

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From its earliest days, the Anatomical Record (AR) has paid ample attention to the autonomic nervous system by publishing multiple seminal papers on this particular field of research. Definitely worth mentioning in this respect are Martin R. Chase's pioneering article on sensory ganglia (1909) and the works of Albert Kuntz (Kuntz, 1909; Kuntz et al. 1956, Hoffman and Kuntz, 1957) on the histogenesis of the ortho- and parasympathetic nervous system as well as the latter author's article in collaboration with John W. Hamilton on the afferent innervation of the skin (Kuntz and Hamilton, 1938). In 1958, the journal dedicated an entire volume comprising no less than 4 issues (Volume 130, January 1958) in honor of Albert Kuntz' widely acclaimed work. In the 30s, the AR published further seminal articles on the autonomic nervous system, for example by Stephen W. Ranson and Peter Mihalik, who provided a detailed description on the structure of the vagal nerve (Ranson and Mihalik, 1932) and by David Bodian, who introduced the famous 'Bodian staining' to stain nerve fibers and nerve endings in mounted paraffin section (Bodian, 1936). This trend continued during and shortly after WWII, with highly relevant papers, for instance, on the anatomy of the lumbosacral and posterior sacral nerve plexus (Horwitz, 1939), the origin and structure of the sympathetic and parasympathetic ganglia in chicken (Smith Jones 1941, 1942). Noteworthy, the journal also published reviews on 'approved films in anatomy and biology', including a 3-hour movie dedicated to the studies of Ivan Pavlov, produced by Techfilm Studio and recorded in the Physiological Laboratory of the Institute of Pediatrics in Saint-Petersburg (Leningrad) (Addison, 1941). In the post WWII period, numerous important AR contributions by Russell T. Woodburne (on the sacral parasympathetic innervation of the colon, 1956), John S. Dixon (on the fine structure of parasympathetic cells in the otic ganglia, 1966), Alan M Laties and David Jacobowitz (on the autonomic innervation of the eye, 1966), Keith C. Richardson (on the ultrastructure of autonomic nerve fibers, 1966, 1969), and Per Freitag and Milton B. Engel (on the autonomic innervation of salivary glands, 1970), further attest to the importance and justified recognition attributed by the journal to this field of research. Somewhat surprisingly, the autonomic innervation of the urogenital system remained largely underexposed in the journal's earlier history. A first extensive report on this topic that I managed to trace back in the historical AR collection, is a paper by DJ Gray of Stanford University (not to be confused with Henry Gray of the Gray's Anatomy textbook) dealing with the intrinsic nerves of the testis (Gray, 1947). Later on, Roberts and Taylor (1970) reported in the AR on the relation of the autonomic nerve supply to the visceral pelvic fascia, in particular the parasympathetic supply to the bladder musculature and the importance to make a clear distinction between the presacral fascia and the fascial sheath of the rectum proper during pelvic surgery in order to avoid functional damage. Using for that time modern neurohistochemical techniques such as

acetylcholinesterase staining, Klück (1980) provided an extensive description of the autonomic innervation of the human urinary bladder and urethra. A seminal AR paper on the hepatic innervation by Robert McCuskey and his lab was published in 1978 (Reilly et al. 1978). More recently, the AR devoted several special issues to subjects dealing with the autonomic nervous system, i.e. "The enteric nervous system and its targets" (Volume 262(1), 2001), "Morphology, cell biology, physiology and pathology of airway receptors" (Volume 270A, 2002), and the "Hepatic nervous system" (Volume 280A, 2004) (fig 1).

Strikingly, the vast majority of the reviews covering historical and conceptual views on the autonomic nervous system, including those published in the Anatomical Record, are very much marked by biased referencing to pioneering studies in this scientific domain. With very few exceptions (e.g. Carlson BM, 1968), references to studies from colleagues in Eastern Europe in the 19th and 20th centuries are not included, which is largely due to both language barriers and lack of access to scientific sources, but definitely also to a number of geopolitical events such as both world wars and the associated postwar period. Therefore, I am grateful to our guest editor, Prof. Dr. Petr Masliukov from the Yaroslavl State University, who despite the very difficult current geopolitical situation, succeeded in engaging a substantial number of colleagues from Eastern Europe in providing a detailed view of the past and current research achievements in the field of the autonomic nervous system. I am also greatly indebted to all contributors to this special issue for their willingness to participate and in this way let science prevail over today's geopolitical issues and blocs.

The groundwork for earlier and, by extension, many of the current concepts on anatomical organization and function of the autonomic nervous system (ANS) was laid by a number of pioneering morphological, pharmacological and physiological papers by well-known researchers, such as Gaskell (1886), Dogiel (1895, 1896, 1899) Bayliss and Starling (1899), Pavlov (1902) Elliot (1905), Langley and Orbeli (1910), Langley (1921), Loewi (1921), Von Euler and Gaddum (1931), Orbeli (1932), Lawrentjew (1934) and Dale (1935). For a more comprehensive overview of all peripheral and central components of the autonomic nervous system I highly recommend reading all the volumes of the Autonomic Nervous System Book Series, edited by the late Geoffrey Burnstock and published at Harwood Academic Publishers.

The first contribution to this special issue by Prof. Alexandr Nozdrachev of the State University of Saint Petersburg (Nozdrachev, 2023) provides a comprehensive review on the pioneering achievements of researchers from Eastern Europe in the field of the autonomic nervous system since the 19th century and also includes an overview of the recent advances in this domain by researchers from this geographical region. Particularly the earlier works were published in either Russian or German and therefore remained somewhat unacknowledged or unknown until translated editions became available.

In contrast to what is still stated in many modern textbooks, the autonomic nervous system extends beyond the classic division between the ortho- and parasympathetic system and should in fact be subdivided into three anatomically distinct parts, i.e. the orthosympathetic system (OS), the parasympathetic system (PS) and the enteric nervous system (ENS). Indeed, already 100 years ago, Langley (1921) – and later on many other authors - provided arguments that the ENS cannot just be considered as a postganglionic component of the PS. All three components in the peripheral nervous system communicate and interact with the central

nervous system and comprise both motoric and sensory pathways. In recent decades, growing evidence has accumulated that there is also an intense bidirectional interaction with the endocrine and immune systems. The first anatomical descriptions of the enteric nerve plexuses by the German anatomist and physiologist George Meissner (1857), the German physician and biologist Leopold Auerbach (1862) and the Russian anatomist Arnold Schabadach (1930), and the subsequent morphological typing of the enteric neurons based on silver impregnation by the Russian histologist and neuroscientist Alexander Stanislavovich Dogiel (1899) and further refined by the East German anatomist Werner Stach (1980, 1981, 1982a, 1982b, 1985, 1989) (fig 2), were later on complemented by studies that exploited then novel techniques, such as intracellular electrophysiological recordings, immunocytochemical stainings and neuronal tracings, allowing scientists to link morphological features to electrophysiological subtyping (S-neurons and AH-neurons) (Nishi and North 1973; Hirst at al. 1974, for a comprehensive review see Wood, 1987) and to a detailed neurochemical coding of these morphologically classified enteric neurons in several mammalian species including human (see for instance Furness and Costa 1987; Bornstein and Furness, 1988; Furness et al 1988; Timmermans et al 1997, 2001; Brehmer et al 1999; Hens et al. 2000; Nurgali et al 2004; Furness 2006; Brehmer 2006, 2021; Lindig et al. 2009). Even today, these early studies continue to serve as a solid base for a plethora of exciting new studies applying fascinating novel techniques that involve optical recording and optogenetic activation (Spencer et al 2020) or the simultaneous application of live cell calcium imaging and whole cell patch clamp even at different topographical levels (Li et al 2022), in attempts to further unravel the complex enteric neurocircuits. Similarly, and in much more detail, single-cell sequencing opens up new perspectives in further exploring the molecular taxonomy of enteric neuronal classes as elegantly shown recently by the Marklund lab (Morarach et al 2021). The guest editor of this special issue, Prof. Dr. Petr Masliukov from Yaroslavl State Medical University, in collaboration with several national and international colleagues, studied for more than 20 years the sympathetic innervation of the gastrointestinal tract at the developmental, neonatal, adult and aged stages of life. His main findings, including neuroplasticity of the sympathetic gastrointestinal innervation, are nicely summarized in a review in this issue (Masliukov et al 2023, this issue), while the original contribution of Proseva and Preobrazhensky (2023, this issue) and those of Budnik and Masliukov (2023, this issue) and Kirov and Lazarov (2023), this issue) focus on the neurochemical profiles of sympathetic preganglionic neurons and myenteric neurons, respectively.

Sympathetic and parasympathetic nerves are the main modulators of the cardiovascular system, regulating heart rate, blood vessel caliber and the contractile behavior of atrial and ventricular myocardiocytes. Chemoreceptors sensitive to a decrease in arterial PO₂, a rise in PCO₂, or a drop in pH as well as baroreceptors measuring blood pressure are located in the carotid and aortic bodies. A first detailed historic overview of the innervation of the carotid body in this journal was provided by Gerard and Billingsley (1923). In the November issue of 1929 and the June issue of 1930 Ralph F Shaner reviewed the then current knowledge of the development of the conduction system in the heart and the development of the innervation of the mammalian heart. In the special AR issue dedicated to Albert Kuntz, AM Rappaport and WD Wilson (1958) published their observations on the ultrastructure of the carotid body. In the late sixties and early seventies, McKibben and Getty (1969), RB Smith (1971) and Walter C. Randall and colleagues (1971) reported in the AR their first observations on the distribution and functional morphology of the intrinsic cardiac innervation of the human neonate,

primates and domestic cattle, respectively. In this special issue, Dainius Pauza and colleagues from the Lithuanian University of Health Sciences in Kaunas (Lithuania) share their longstanding expertise and further elaborate on the comparative anatomy of the cardiac and epicardiac innervation as well on the cardiac conducting system validating several models for cardiac research and cardiac pathology (Saburkina et al, 2023; Pauziene et al, 2023; Inokaitis et al 2023). This part on the autonomic innervation of the heart is further complemented by a review article from Filipovic and collaborators from the University of Split (Croatia) on our current knowledge of the pathophysiological mechanisms and diagnostic approaches related to cardiac autonomic neuropathy (Filipovic et al, 2023), by a joint study from the Bulgarian Academy of Sciences, the Trakia University in Stara Zagora and the Medical University of Sofia demonstrating the neuronal plasticity displayed by the carotid body in hypertension (Atanasova et al, 2023) and by a contribution of Vakhrushev et al (2023) dealing with the significant contribution of ANS dysfunction to the development of cardiovascular diseases such as arterial hypertension, coronary artery disease and chronic heart failure. In most cases the observed symptoms are due to an increase in sympathetic activity and a decrease in parasympathetic activation. Standard treatment consists of catheter-based renal artery denervation by means of radiofrequency ablation. Considering the large variability in outcome of this technique, these authors from the Almazov National Medical Research Center in Saint-Petersburg propose transcatheter laser renal denervation as a valid alternative approach and demonstrate that this technique is as effective as radiofrequency ablation while causing much less damage to the intimal layer.

The last two contributions to this special issue deal with neurochemical features of specific areas in the brain (Romanova et al, 2023) and the spinal cord (Veshchitskii et al, 2023), known to be part of the central nervous system control of the autonomic outflow.

While I do recognize that this special issue has inevitably been subject to some degree of selectivity and hence does cover only some aspects of the autonomic nervous system, I am convinced that the contributions in this special issue provide a good picture of how the achievements of researchers in Eastern Europe and the former USSR have contributed to our current insights into the morphofunctional organization of the autonomic nervous system. and would hence like to offer my heartfelt thanks to our Guest Editor Prof. Dr. Petr Masliukov for his time and efforts in making this exciting special issue possible and to all contributing authors from distinct countries in Eastern Europa illustrating that science and unity between scientists from Russia and colleagues from independent democratic countries can still surmounts threathening geopolitical situations.

References

-Addison WHF (1941) Reviews of approved films in anatomy and biology. Anat Rec 79(4):549-

-Atanasova D, Dandov A, Lazarov N (2023) Neurochemical plasticity of the carotid body in hypertension. Anat Rec, this issue

2 3 -Auerbach L (1862) Ueber einen Plexus myentericus, einen bisher unbekannten ganglio-4 nervösen Apparat im Darmkanal der Wirberlthiere. In: Morgenstern, Breslau, pp 1-13 5 -Bayliss WM, Starling EH (1899) The movements and innervation of the small intestine. J 6 Physiol 24: 99-143 7 8 -Bodian D (1936) A new method for staining nerve fibers and nerve endings in mounted 9 paraffin sections. Anat Rec 65 (1): 89-97 10 -Bornstein JC, Furness JB (1988) Correlated electrophysiological and histochemical studies of 11 submucous neurons and their contribution to understanding enteric neural circuits. J Auton 12 Nerv Syst 25 (1): 1-13 13 14 -Brehmer et al (1999) Morphologial classification of enteric neurons – 100 years after Dogiel. 15 Anat Embryol 200 (2): 125-135 16 Brehmer A (2006) Structure of enteric neurons. Adv Anat Embryol Cell Biol 186:1-94 17 -Brehmer A (2021) Classification of human enteric neurons. Histochemistry and Cell Biology 18 19 156: 95-108 20 -Budnik A, Masliukov P (2023) Postnatal development of the enteric neurons expressing 21 neuronal nitric oxide synthase. Anat Rec, this issue 22 -Carlson BM (1968) Regeneration research in the Soviet Union. Anat Rec 160(4):665-674 23 -Chase MR (1909) A histological study of sensory ganglia. Anat. Rec. 3: 121-140 24 25 -Dale H (1935) Pharmacology and nerve endings. Proc Roy Soc Med 28: 319-332 26 -Dixon JS (1966) The fine structure of parasympathetic nerve cells in the otic ganglia of the 27 rabbit. Anat Rec 156(3): 239-251 28 -Dogiel AS (1895) Zur Frage über den feineren Bau des sympathischen Nervensystems bei den 29 30 Säugethieren. Archiv für Mikroskopische Anatomie 46 (1): 305-344 31 -Dogiel AS (1896) Zwei Arten sympathischer Nervenzellen. Anat Anz 11 (22) 679-687 32 -Dogiel AS (1899) Ueber den Bau der Ganglien in den Geflechten des Darmes und der 33 Gallenblase des Menschen und der Säugethiere. Arch Anat Physiol Anat Abt (Leipzig) 1899: 34 130-158 35 36 -Elliot TR (1905) The action of adrenalin. J Physiol 32: 401-467 37 -Filipovic N, Marinovic Guic M, Kosta V, Vukojevic K (2023) Cardiac innervations in diabetes 38 mellitus – anatomical evidence of neuropathy. Anat Rec, this issue 39 -Freitag P, Engel MB (1970) Autonomic innervation in rabbit salivary glands. Anat Rec 167(1): 40 41 87-105 42 -Furness JB (2006) The enteric nervous system; Blackwell, Oxford 43 -Furness JB, Costa M (1987) The enteric nervous system. Churchill Livingstone, Edinburgh 44 -Furness JB, Bornstein JC, Trussell DC (1988) Shapes of nerve cells in the myenteric plexus of 45 the guinea-pig small intestine revealed by the intracellular injection of dye. Cell Tissue Res 46 47 254(3): 561-571 48 -Garner CM, Duncan D (1958) Observations on the fine structure of the carotid body. Anat Rec 49 130(4): 691-709 50 -Gaskell WH (1886) On the structure, distribution and function of the nerves which innervate 51 52 the visceral and vascular structures. J Physiol 7: 1-80 53 -Gerard MW, Billingsley PR (1923) The innervation of the carotid body. Anat Rec. 25 (6): 391-54 400 55 -Gray DJ (1947) The intrinsic nerves of the testis. Anat Rec 98 (3):325-335 56 -Hens J, Schrödl F, Brehmer A, Adriaensen D, Neuhuber W, Scheuermann DW, Schemann M, 57 58 Timmermans JP (2000) Mucosal projections of enteric neurons in the porcine intestine. J Comp 59 Neurol 421: 429-436 60

4

5

6 7

8

9

-Hirst GDS, Holman ME, Spence I (1974) Two type of neurons in the myenteric plexus of duodenum in the guinea pig. J Physiol 236: 303-326 -Hoffman HH, Kuntz A (1957) Vagus nerve components. Anat Rec 127 (3):551-567 -Horwitz MT (1939) The anatomy of (A) the lumbosacral nerve plexus – its relation to variations of vertebral segmentation, and (B), the posterior sacral nerve plexus. Anat Rec 74(1): 91-107 10 -Inokaitis H, Pauziene N, Pauza D (2023) The distribution of sinoatrial nodal cells and their 11 innervation in the pig; Anat Rec, this issue 12 -Jones DS (1941) Further studies on the origin of sympathetic ganglia in the chick embryo. 13 14 Anat Rec 79(1): 7-15 15 -Jones DS (1942) The origin of the vagi and the parasympathetic ganglion cells of the viscera 16 of the chick. Anat Rec 82(2): 185-197 17 -Kirov T, Atanasova D, Lazarov N (2023) Neurochemical profile of the myenteric plexus in the 18 19 rat colorectal region. Anat Rec, this issue 20 -Klück P (1980) The autonomic innervation of the human urinary bladder, bladder neck and 21 urethra: a histochemical study. Anat Rec. 198(3) :439-447 22 -Kuntz A (1909) A contribution to the histogenesis of the sympathethic nervous system. Anat 23 Rec. 3: 458-465 24 25 -Kuntz A, Hamilton JW (1938) Afferent innervation of the skin. Anat Rec 71 (4): 387-400 26 -Kuntz A, Hoffman HH, Jacobs MW (1956) Nerve fiber components of communicating rami 27 and sympathetic roots in man. Anat Rec. 126 (1): 29-41 28 -Langley JN, Orbeli LA (1910) Observations on the sympathetic and sacral autonomic system 29 30 of the frog. J Physiol 41 (5): 450-482 31 -Langley JN (1921) The autonomic nervous system. Part 1. Cambridge W. Heffner 32 -Laties AM, Jacobowitz D (1966) A comparative study of the autonomic innervation of the eye 33 in monkey, cat and rabbit. Anat Rec 156(4): 383-395 34 -Lawrentjew BJ (1934) Einige Bemerkungen über Fortschritte und Aufgaben der Erforschung 35 36 des autonomen Nervensystems. Z Mikrosk Anat Forsch 36: 651-659 37 -Li Z, Boesmans W, Kazwiny Y, Hao MM, Vanden Berghe P (2022) Simultaneous whole-cell 38 patch-clamp and calcium imaging on myenteric neurons. Am J Physiol Gastrointest Liver 39 Physiol 323 (4): G341-G347 40 41 -Lindig ML, Kumar V, Kikinis R, Pieper S, Schrödl F, Neuhuber WL, Brehmer A (2009) Spiny 42 versus stubby: 3D reconstruction of human myenteric (type I) neurons. Histochem Cell Biol 43 131:1-12 44 -Masliukov P, Emanuilov A, Budnik A (2023) Sympathetic innervation of the development, 45 maturity, and aging of the gastrointestinal tract. Anat Rec, this issue 46 47 -McKibben JS, Getty R (1969) A study of the cardiac innervation in domestic animals: cattle. 48 Anat Rec 165(2): 141-151 49 -Meissner G (1857) Über die Nerven der Darmwand. Z Ration Med N F 8: 364-366 50 -Morarach K, Mikhailova A, Knoflach V, Memic F, Kumar R, Li W, Ernfors P, Marklund U (2021) 51 52 Diversification of molecularly defined myenteric neuron classes revealed by single-cell RNA 53 sequencing. Nat Neurosci 24(1): 34-46 54 -Nishi S, North RA (1973) Intracellular recording from the myenteric plexus of the guinea-pig 55 ileum. J Physiol 231: 471-491 56 -Nozdrachev A (2023) A brief retrospective of Russian research of the autonomic nervous 57 58 system. Anat Rec, this issue 59 60

2	
3	Nurgali K Stephing MI Eurness IB (2004) Correlation of electrophysiological and
4	rungan k, stebbing wis, runess 3D (2004) conclution of cleetrophysiological and
5	morphological characteristics of enteric neurons in the mouse colon. J Comp Neurol 468(1):
5	112-124
0	
7	-Orbell LA (1932) Review of sympathetic innervation of skeletal muscle, sensory organs and
8	central nervous system. Sechenov Physiol J USSR 15: 1-22
9	
10	-Paviov I (1902) The work of the digestive glands (translation by WH Thompson, Charles Griffin
10	& Company, Ltd. London
	Deutine N. Deneguine K. Ducqueite K. Inckeitic II. Schurking L. Dickhangur K. Schocking D.
12	-Pauziene N, Ranceviene K, Rysevalle K, mokallis H, Saburkina I, Piekhanova K, Sabeckiene D,
13	Sabeckis I, Martinaityte R, Pilnikovaite E, Pauza D (2023) Comparative analysis of intracardiac
14	noural structures in the aged rate with essential hypertension. Anat Rec. this issue
15	neural structures in the agent rats with essential hypertension. Anat Net, this issue
16	-Porseva V, Preobrazhensky N (2023) Neuronal nitric oxide synthase and calbindin expression
10	in sympathetic preganglionic neurons following capsaisin treatment. Anat Rec. this issue
17	
18	-Randall WC, Armour JA, Randall DC, Smith OA (1971) Functional anatomy of the cardiac
19	nerves in the baboon. Anat Rec 170 (2): 183-198
20	Panson SM/ Mihalik $D(1022)$ The structure of the views name. Anot $Paper E4(2) 2EE 260$
21	-Ranson Sw, Minalik P (1932) The structure of the vagus herve. Anat Rec 54 (3) 355-360
21	-Rappaport AM, Wilson WD (1958) The structural and functional unit in the human liver (liver
22	acinus) Anat Roc 120 (4): 672 689
23	acinus/ Anat nec 150 (4). 075-089
24	-Reilly FD, McCuskey PA, McCuskey RS (1978) Intrahepatic distribution of nerves in the rat.
25	Anat Rec 191(1): 55-67
26	
27	-Richardson KC (1966) Electron microscopic identification of autonomic nerve endings. Nature
20	210: 756
20	Richardson KC (1060) The fine structure of autonomic nerves ofter vital staining with
29	-Richardson RC (1909) The line structure of autonomic nerves after vital staming with
30	methylene blue. Anat Rec 164(3): 359-377
31	-Roberts WH, Taylor WH (1970) The presacral component of the visceral pelvic fascia and its
32	relation to the netwise relation of the bladder. Anot Dec 100(2): 207,212
33	relation to the pelvic splanchnic innervation of the bladder. Anat Rec 166(2): 207-212
34	-Romanova IV, Mikhailova EV, Mikhrina AL, Shpakov AO (2023) Type 1 melanocortin receptors
25	in pro oniomologocortin vacoprossin and ovytacin immunopositivo pourons in different
35	in pro-optometanocortine, vasopressine and oxytocine initiatiopositive neurons in different
30	areas of mouse brain. Anat Rec, this issue
37	-Saburkina L Pauziene N, Solomon OL Rysevaite K, Pauza D (2023) Comparative gross anatomy
38	Subarticle in the second of th
39	of epicardiac ganglionated nerve plexi on the human and sheep cardiac ventricles. Anat Rec,
40	this issue
41	Schabadasch A (1020) Intramurale Nervengeflechte des Darmrehrs, 7 Zeilfersch Mikrosk Anat
40	-schabadasch A (1950) intrainurale Nervengenechte des Darmonns. 2 Zeinorsch Wikrosk Anat
42	10: 320-385
43	-Shafer RE (1929) The development of the atrioventricular node, bundle of His, and sino-atrial
44	
45	node in the calf; with a description of a third embryonic node-like structure. Anat Rec. 44(2):
46	85-99
47	Shafer RE (1020) On the development of the nerves to the mermedian heart. Anot Res. 46
19	-shale KF (1950) On the development of the nerves to the manimalian heart. Anat Kec. 46
40	(1): 23-39
49	-Smith RB (1971) The occurrence and location of intrinsic cardia ganglia and nerve pleyuses in
50	Sinti No (1371) The occurrence and location of intrinsic cardia galigina and herve prevases in
51	the human neonate. Anat Rec 169(1): 33-40
52	-Spencer NJ, Travis L, Hibberd T, Kelly N, Feng J, Hu H (2020) Effects of optogenetic activation
53	of the optomic nervous system on gestraintestinal matility is reason anallistating. Automore
54	or the enteric hervous system on gastrointestinal motility in mouse small intestine. Autonomic
5-	Neuroscience: Badic and Clinical 229: epub 102733
22	-Stach W (1980) Neuronal organization of the myenteric plevus (Auerbach) in the small
56	Stach w (1900) weutonal organization of the myenteric piexus (Auerbach) in the Shall
57	intestine of the pig (in German). I. Type I neurons. Z Mikrosk Anat 94 (5): 833-849
58	-Stach W (1981) Neuronal organization of the myenteric plexus (Auerbach) in the small
59	interting of the nig (in Cormon) II. Type II neurone, $7 M_{\rm excel}$ And $0 \Gamma(2)$, $4 C I (402)$
60	intestine of the pig (in German). II. Type II neurons. 2 Wikrosk Anat 95(2): 161-182

-Stach W (1982a) Neuronal organization of the myenteric plexus (Auerbach) in the small intestine of the pig (in German). III. Type III neurons. Z Mikrosk Anat 96(3): 497-994

-Stach W (1982b) Neuronal organization of the myenteric plexus (Auerbach) in the small intestine of the pig (in German). IV. Type IV neurons. Z Mikrosk Anat 96(6): 972-994

-Stach W (1985) Neuronal organization of the myenteric plexus (Auerbach) in the small intestine of the pig (in German). V. Type V neurons. Z Mikrosk Anat 99(4): 562-582

-Stach W (1989) A revised morphological classification of neurons in the enteric nervous system. In: Singer MV, Goebell H (eds) Nerves and the gastrointestinal tract. Kluwer, Lancaster, pp. 29-45

-Timmermans JP, Adriaensen D, Cornelissen W, Scheuermann DW (1997) Structural organization and neuropeptide distribution in the mammalian enteric nervous system, with special attention to those components involved in mucosal reflexes. Comp Biochem Physiol 118A (2): 331-340

-Timmermans JP, Hens J, Adriaensen D (2001) Outer submucous plexus: an intrinsic nerve network involved in both secretory and motility processes in the intestine of large mammals and humans. Anat Rec 262: 71-78

-Vakhrushev AD, Condori Leandro HI, Goncharova NS, KorobchenkoLE, MitrofanovaLB, Makarov IA, Andreeva EM, Lebedev DS, Mikhaylov, EN (2023) Laser renal denervation: a comprehensive evaluation of microstructural renal artery lesions and impact on hypertension inducibility. Anat Rec, this issue

-Veshchitskii AA, Kirik OV, Korzhevskii DE, Merkulyeva N (2023) Development of neurochemical labelling in the intermediolateral nucleus of cats' spinal cord. Anat Rec, this issue

-Von Euler US, Gaddum JH (1931) An unidentified depressor substance in certain tissue extracts. J Physiol 72: 74-87

-Wood JD (1987) Physiology of the enteric nervous system. In: Physiology of the Gastrointestinal Tract, 2nd Ed, Ed. Johnson LR, PP. 367-398, New York, Raven Press

-Woodburne RT (1956) The sacral parasympathetic innervation of the colon. Anat Rec 124(1): 67-76

Figure Legends:

Fig 1: Covers of AR special issues dealing with different aspects of the autonomic nervous system.

Fig 2: (a, c) Images taken from the historical collection of a silver-impregnated intestinal whole-mounts of Prof. Em. Werner Stach (Rostock). (a) Myenteric ganglion of the porcine small intestine showing Dogiel type I, II and III neurons. Inset: picture of Werner Stach taken this year at the age of 87 (left) and Alexandr Dogiel (right). (b) Outer submucous ganglion immunostained with an antibody raised against neurofilaments showing the typical type II morphology seen with silver-impregnation. (c) Detail of a myenteric Dogiel type I neuron after silverimpregnation. (d) confocal 3D reconstruction of a similar myenteric Dogiel type I neuron after immunostaining for the tachykinin receptor, neurokinin 1. (e) Additional morphological classification of enteric neurons according to Stach (*modified from Timmermans et al, 1997*).





Figure 1. 171x72mm (300 x 300 DPI)

Dogiel Type I ne

Dogiel Type II neurons

type I neurons

type V neurons

dendritic type II neurons

50 µm



John Wiley & Sons, Inc.



type III neurons

50 µm

2a



type IV neurons

type VII neurons



type VI neurons

type II neurons

extended classification by W. Stach 2e

Figure 2.

171x265mm (300 x 300 DPI)