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1 **The sensitivity of ginkgo leaf unfolding to the temperature and**
2 **photoperiod decreases with increasing elevation**

3 Zhaofei Wu^{1#}, Chen-Feng Lin^{2#}, Shuxin Wang¹, Yufeng Gong¹, Yun-Peng Zhao^{2*}, Jing Tang^{3,4,5},

4 Hans J. De Boeck⁶, Yann Vitasse⁷, Yongshuo H. Fu^{1*}

5 **Affiliations**

6 ¹ College of Water Sciences, Beijing Normal University, Beijing, China

7 ² Systematic & Evolutionary Botany and Biodiversity Group, MOE Key Laboratory of Biosystems
8 Homeostasis & Protection, College of Life Sciences, Zhejiang University, Hangzhou 310058, China

9 ³ Terrestrial Ecology Section, Department of Biology, University of Copenhagen,
10 Universitetsparken 15, DK-2100, Copenhagen Ø, Denmark

11 ⁴ Center for Permafrost (CENPERM), University of Copenhagen, Øster Voldgade 10, DK-1350,
12 Copenhagen K, Denmark

13 ⁵ Department of Physical Geography and Ecosystem Science, Lund University, Sölvegatan 12, SE-
14 223 62, Lund, Sweden

15 ⁶ Plants and Ecosystems, Department of Biology, University of Antwerp, Antwerp, Belgium

16 ⁷ Swiss Federal Institute for Forest, Snow and Landscape Research (WSL), Birmensdorf,
17 Switzerland

18 **Corresponding authors:**

19 Yongshuo H. Fu

20 Phone: +86-10-58802736

21 Email: yfu@bnu.edu.cn

22 Yun-Peng Zhao

23 Phone: +86-571-88206463

24 Email: ypzhao@zju.edu.cn

25 Any figure does not need to used color in print.

26 **Abstract**

27 Climate change substantially affects plant phenology, resulting in earlier vegetation
28 onset across temperate and boreal regions. Phenological shifts caused by warming may
29 alter species interactions across trophic levels because of species-specific responses,
30 and influence the reproductive success of dioecious species if the phenological
31 sensitivity to warming (S_T) differs between genders. We used twigs collected from male
32 and female ginkgo trees at three elevations on Tianmu Mountain in eastern China. The
33 twigs were cultivated in climate chambers to determine the effects of three temperatures
34 (10, 15, and 20 °C) and two photoperiods (8 and 16 h). We observed slightly earlier leaf
35 unfolding dates in male twigs (1 day), and a higher heat requirement (growing degree
36 hours) for leaf unfolding in female ($14,334 \pm 588$ °C) compared to male twigs ($13,874$
37 ± 551 °C). Similar responses to temperature ($S_T = 3.7$ days °C⁻¹), photoperiod and
38 elevation were observed across genders. The long photoperiod treatment shortened the
39 time to leaf unfolding by 9.1 days, but temperature and photoperiod effects on leaf
40 unfolding differed significantly depending on the elevation of the donor trees.
41 Specifically, S_T was higher (4.17 days °C⁻¹) and the photoperiod effect on S_T was larger
42 (decreased by 1.15 days °C⁻¹) at the lowest elevation than at the higher elevations (S_T
43 $= 3.26$ days °C⁻¹; decreased by 0.48 days °C⁻¹). This may be related to environment-
44 induced local adaptations and self-protection mechanisms of trees at high elevations to
45 avoid frost damage. Our results indicate that the photoperiod and genetic adaptations to
46 local environments influenced the warming-induced phenological responses in ginkgo,
47 but these responses were generally similar between the genders. For a given species,

48 individuals in different climates may exhibit different phenological responses to higher
49 temperatures, with individuals in warmer regions likely becoming increasingly limited
50 by the photoperiod as the climate warms further.

51

52 **Keywords:** climate change, temperature sensitivity, photoperiod, elevation, twig
53 cutting experiment, gymnosperm

54 **1. Introduction**

55 The timing of vegetation onset is a crucial regulatory factor affecting the terrestrial
56 carbon sequestration in temperate and boreal regions (Keeling *et al.*, 1996; Richardson
57 *et al.*, 2013; Wu *et al.*, 2019; Piao *et al.*, 2020). It also influences the regional and global
58 climate by providing biophysical and biogeochemical feedback to the climate system
59 (Peñuelas *et al.*, 2009; Piao *et al.*, 2019). Ground monitoring data and satellite-based
60 vegetation indices have revealed significant tree phenological shifts in response to
61 global warming worldwide (Peñuelas & Filella, 2001; Fu *et al.*, 2015b; Menzel *et al.*,
62 2020; Piao *et al.*, 2020; Wu *et al.*, 2021). For a given species, these phenological
63 changes are generally greater at high latitudes and/or elevations (Vitasse *et al.*, 2018;
64 Gao *et al.*, 2020). This phenomenon may be explained by the warming-induced
65 environmental constraints affecting warm-edge species distributions. More specifically,
66 the relatively short photoperiod and lack of chilling conditions prevent the complete
67 release of winter dormancy. However, this possibility requires experimental verification.

68

69 Spring phenological sensitivity to warming (S_T ; advancement of leaf unfolding in days
70 per degree warming, days °C⁻¹) has declined significantly over the past few decades
71 (Fu *et al.*, 2015b). This may be partially related to a shortening photoperiod, during
72 which buds become sensitive to increasing temperatures and to the increasing thermal
73 requirements for bud burst resulting from decreased chilling during winter (Fu *et al.*,
74 2019). Although chilling is essential for releasing tree buds from dormancy (Chuine *et al.*,
75 *et al.*, 2010; Yu *et al.*, 2010; Baumgarten *et al.*, 2021), the effects of a shortened

76 photoperiod on leaf unfolding and phenological sensitivity to warming remain unclear
77 (but see Fu *et al.*, 2019). Indeed, for photoperiod-sensitive species, a shortened
78 photoperiod may increase the amount of heat required for leaf unfolding, thereby
79 delaying cell development. This so-called “photoperiod limitation” represents a
80 protective mechanism enabling trees to avoid being damaged from frost events after
81 warm spells in late winter and early spring (Korner & Basler, 2010; Fu *et al.*, 2019).
82 Additionally, there are genetic differences underlying the diverse phenological
83 responses among populations growing in contrasting climates (Vitasse *et al.*, 2013).
84 There is very little available information regarding the possible genetic adaptations to
85 specific photoperiods and whether populations growing in regions with the same
86 photoperiod, but varying temperatures (i.e., along an elevational gradient), have
87 evolved to adapt to differences in the interactive effects of temperature and photoperiod.
88 How elevation alters spring phenological responses to climate change, especially under
89 different photoperiods have received less attention and were urgently needed to be
90 illustrated.

91

92 To predict future phenological shifts, manipulative experiments are particularly
93 relevant as they allow us to investigate phenological responses beyond the range of
94 historical temperature conditions and to change one environmental variable at a time in
95 a controlled manner. Therefore, studies using twig cuttings under different chilling,
96 forcing, and photoperiod conditions in climate chambers have recently been conducted
97 (Flynn & Wolkovich, 2018; Baumgarten *et al.*, 2021). Twig cuttings have been proven

98 to be a viable alternative to donor trees for phenological studies (Vitasse & Basler, 2014;
99 Du *et al.*, 2019).

100

101 Although most of the previous related studies focused on the phenological responses of
102 European and North American angiosperms to climate change (Basler & Körner, 2012;
103 Fu *et al.*, 2019; Vitasse *et al.*, 2009; Flynn & Wolkovich, 2018), there has been some
104 research on deciduous gymnosperms and Asian species (Matsumoto *et al.*, 2003;
105 Kumar & Sati, 2016). Ginkgo (*Ginkgo biloba* L.), which represents a gymnosperm
106 lineage, is a tall deciduous dioecious tree native to China, but it is currently distributed
107 worldwide, including at various latitudes (Zhao *et al.*, 2019). The morphology of this
108 ‘living fossil’ species has essentially not changed for at least 200 million years (Major,
109 1967; Zhou & Zheng, 2003) and its ecological traits have been conserved since the late
110 Cretaceous period (Royer *et al.*, 2003). Its survival during the Quaternary glaciations
111 depended on range shifts as well as environmental adaptations, some of which were
112 related to robust responses to abiotic and biotic stresses (Zhao *et al.*, 2019). Furthermore,
113 a previous study suggested that the responsiveness of leaf unfolding to temperature may
114 vary between male and female individuals (Sofi, 2018). Investigating the difference of
115 ginkgo leaf unfolding responses to climate change between genders and the interplay
116 with photoperiod is important for comprehensively characterizing the phenological
117 responses of gymnosperms to ongoing climate change.

118

119 In this study, we conducted a manipulative experiment on the twigs of male and female

120 ginkgo to evaluate the effects of two photoperiods (8 and 16 h) and three temperatures
121 (10, 15, and 20 °C) on leaf unfolding, as well as the response differences along elevation
122 gradients. We aimed to address the following scientific questions: 1) how the timing of
123 leaf unfolding response to warming and photoperiod between male and female
124 individuals, and 2) how the photoperiod affects the temperature sensitivity of leaf
125 unfolding among different elevations. We tested the hypotheses that short photoperiod
126 affects the phenological sensitivity to temperature (Flynn & Wolkovich, 2018; Fu *et al.*,
127 2019; Zohner *et al.*, 2016) and this effect may be larger in the provenances at lower
128 elevations as a protective mechanism against warm events in late winter (Vitasse *et al.*,
129 2013); and 2) leaf unfolding dates and phenological sensitivity are similar between male
130 and female individuals to safeguard reproductive success under different climatic
131 conditions, but may differ among provenances (elevations).

132 **2. Materials and Methods**

133 *2.1 Sample collection sites*

134 Twigs were collected from adult ginkgo trees on 17–19 January 2021 on Tianmu
135 Mountain (119° 23' 47"–119° 28' 27" E, 30° 18' 30"–30° 24' 55" N) in Zhejiang
136 province in eastern China. More specifically, samples were collected at the following
137 three elevations: E1: 344 ± 23 m, E2: 826 ± 39 m, and E3: 1,098 ± 41m. The aerial
138 temperature 15 cm above ground and soil water content 8 cm below ground were
139 recorded with an interval of 15 min using Temperature-Moisture-Sensor loggers (TMS-
140 5; TOMST, Prague, Czech; Wild *et al.*, 2019). The mean values of both parameters for

141 the three elevation classes were 15.75 ± 0.09 °C and $25 \pm 1\%$, 13.43 ± 0.73 °C and 23
142 $\pm 6\%$, 11.94 ± 0.21 °C and $29 \pm 3\%$, respectively (Table S1). The dominant vegetation
143 type on Tianmu Mountain is a subtropical evergreen and deciduous broad-leaved mixed
144 forest at an elevation of 300–1,506 m. The occurrence of ginkgo in China, mostly
145 cultivated, and the location of Tianmu Mountain is presented in Figure S1 (Gu *et al.*,
146 2021). According to our observation, cultivated ginkgo trees are bigger than 10 cm for
147 trunk diameter.

148 *2.2 Experimental design*

149 At each elevation site, we selected 11 or 12 adult trees, ideally with an equal number of
150 female and male individuals (Table S1). For each donor tree, we collected 12 healthy
151 twigs (30–40 cm long) from the south-facing side of the crown. The twigs were pruned,
152 cleaned, and disinfected with a commercial hypochlorite solution as previously
153 described (Basler & Körner, 2012; Du *et al.*, 2019). The pruned twigs were placed in
154 395-ml plastic bottles, which were filled with tap water (up to 2/3 of the total volume),
155 and then incubated in three climate chambers at Beijing Normal University. The water
156 was changed every 2 weeks during the experiment. Additionally, the twigs were pruned
157 2 cm at the base to avoid the occlusion of vessels.

158

159 The climate chambers were set at 10 °C (T10), 15 °C (T15), and 20 °C (T20) for the
160 duration of the experimental period. We used a constant temperature above 10 °C to
161 avoid any confounding effects due to chilling (Baumgarten *et al.*, 2021). In each climate

162 chamber (i.e., temperature treatment), two photoperiod treatments were set up by
163 covering half of the samples with shade cloth daily for 16 h (P16; long photoperiod
164 treatment) and 8 h (P8; short photoperiod treatment). Because we selected twigs from
165 male and female trees at three elevations, analyses involved 36 combinations (three
166 temperatures \times two photoperiods \times two genders \times three elevations) with 11–12 twigs
167 per treatment (one per tree). In each treatment, temperature sensors (HOBO MX2202)
168 were used to record the air temperature every 30 min. The photoperiod treatment did
169 not affect the temperature (Fig. S2). The photon flux in all chambers was approximately
170 $161 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$, which was consistent with that in an earlier investigation (Du *et al.*,
171 2019).

172 *2.3 Phenology monitoring*

173 We monitored the leaf unfolding of each twig every 3 days. The leaf unfolding stage
174 was defined as the period in which at least one leaf was fully unfolded (Vitasse, 2013).
175 During the phenological analysis, we also recorded the number of unopened buds and
176 calculated the percentage of unopened buds per twig (i.e., number of unopened buds
177 divided by the total number of buds on a twig).

178 *2.4 Data analysis*

179 The growing degree hours (GDH) were calculated as follows:

$$180 \quad GDH = \sum_t^{LU} \left(\sum_1^{24} (T_{hour} - T_{base}) \right) \text{ if } T_{hour} > T_{base}$$

181 where T_{hour} is the hourly temperature and T_{base} is the base temperature, which was set

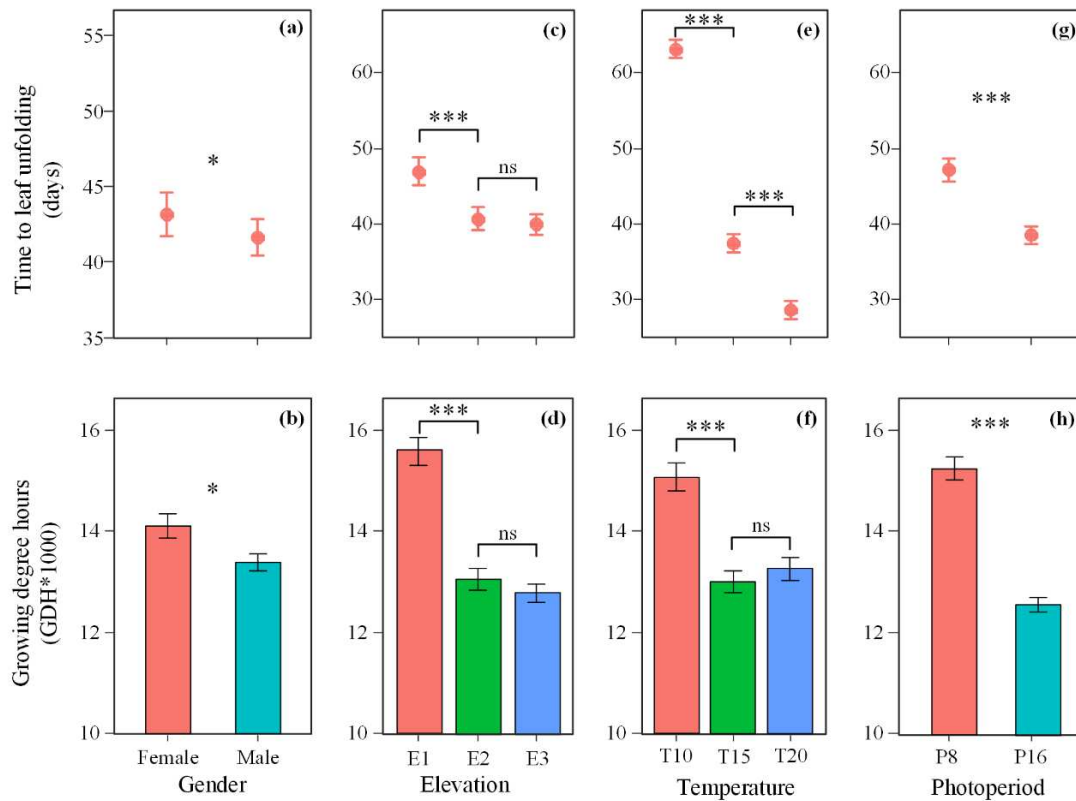
182 at 0 °C on the basis of a previous study (Fu *et al.*, 2016). The hourly accumulation of
183 GDH was calculated from the start of the climate chamber experiment (23 January) to
184 the date of leaf unfolding (LU). The treatment time to reach the leaf unfolding stage
185 (i.e., the number of days from the start of the experiment until at least one leaf had
186 unfolded) was recorded. Additionally, the requirement for the GDH of twigs during this
187 period was calculated for each treatment. Moreover, the paired *t*-tests were used to
188 evaluate the differences between the mean time of leaf unfolding, the GDH required for
189 leaf unfolding, and the percentage of unopened buds under different treatments.
190 Furthermore, as previous studies revealed that a certain amount of low temperatures
191 during winter (higher than the freezing point, but lower than a temperature threshold),
192 or called chilling, is required to break endodormancy (Cannell & Smith, 1983; Chuine
193 *et al.*, 2013; Fu *et al.*, 2016), which occurred before January (Beil *et al.*, 2021). After
194 checking the temperature at the collection site, i.e. lower than 8 °C in January at Tianmu
195 mountain, we therefore selected the 8 °C as the threshold following previous studies
196 (Chuine, 2000; Fu *et al.*, 2019), and estimated the chilling hours when the site hourly
197 temperature was between 0 and 5 °C starting from 1 September to 31 December 2020.
198 The temperature sensitivity of leaf unfolding (S_T) was calculated according to linear
199 regression analyses of the leaf unfolding date of each twig and the temperature recorded
200 by the HOBO sensors (Onset Computer Corporation, USA) for each treatment during
201 the experimental period. Differences in the temperature sensitivity between the genders,
202 elevations, and the photoperiod treatments were evaluated according to ANCOVAs,
203 which can test the significance of both the slope and the intercept of the linear

204 relationships (Fu *et al.*, 2019). All statistical analyses were conducted using R (version
205 3.5.2).

206 **3. Results**

207 *3.1 Differences in leaf unfolding between genders and provenance elevations*

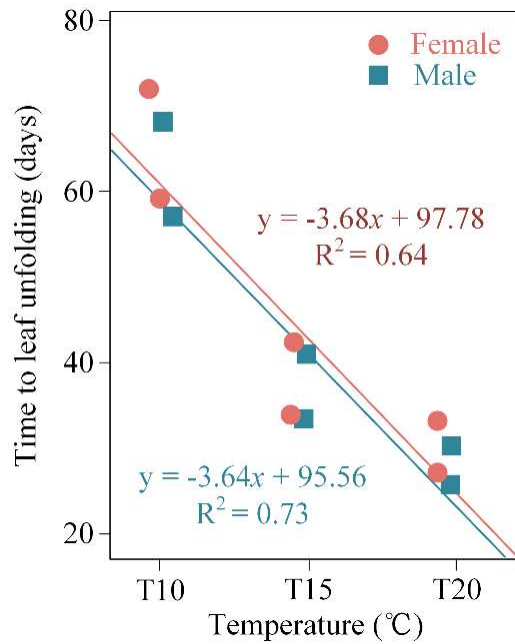
208 Leaf unfolding occurred slightly earlier on male twigs than on female twigs (1 day
209 earlier) (Fig. 1a). The heat requirement (GDH) for leaf unfolding was also mildly
210 greater for female twigs ($14,334 \pm 588$ °C) than for male twigs ($13,874 \pm 551$ °C) (Fig.
211 1b). In addition, there was no significant difference in S_T between female and male
212 samples (i.e., 3.68 ± 0.22 days °C⁻¹ and 3.64 ± 0.17 days °C⁻¹, respectively; Fig. 2). We
213 also found that the responses of leaf unfolding to photoperiod and elevation were same
214 between male and female (Fig. S3). These results suggest that the leaf unfolding and its
215 response to environment changes between male and female ginkgo trees are well
216 synchronized. Therefore, in the following analysis of the effects of temperature,
217 photoperiod, and elevation on leaf unfolding, we used the mixed dates across male and
218 female samples.



219

220 **Fig. 1** Time to leaf unfolding for twigs (mean \pm se, the upper panel), and heat
 221 requirement required for leaf unfolding (defined as the growing degree hours, the lower
 222 panel) for different gender (a, b), elevation (c, d), temperature (e, f) and photoperiod (g,
 223 h) treatments. “ns” indicates that no significant differences exist between two
 224 treatments, * and *** indicate $P < 0.05$ and $P < 0.001$, respectively.

225



226

227 **Fig. 2** Time to leaf unfolding for both male and female trees. The temperature
 228 sensitivity of leaf unfolding was determined as the slope of linear regression between
 229 temperature treatments and the time to leaf unfolding.

230

231 The time required to reach the leaf unfolding stage was significantly longer for the twigs
 232 collected at the lowest elevation (E1 = 50.5 days) than for the twigs collected at the two
 233 higher elevations (E2 = 41.3 days and E3 = 40.8 days; no significant difference) (Fig.
 234 1c). The requirement of GDH at the leaf unfolding stage was greater for the E1 twigs
 235 (16,128 °C) than for the twigs from the two higher elevations (E2 = 13,161 °C and E3
 236 = 13,024 °C; no significant difference) (Fig. 1d).

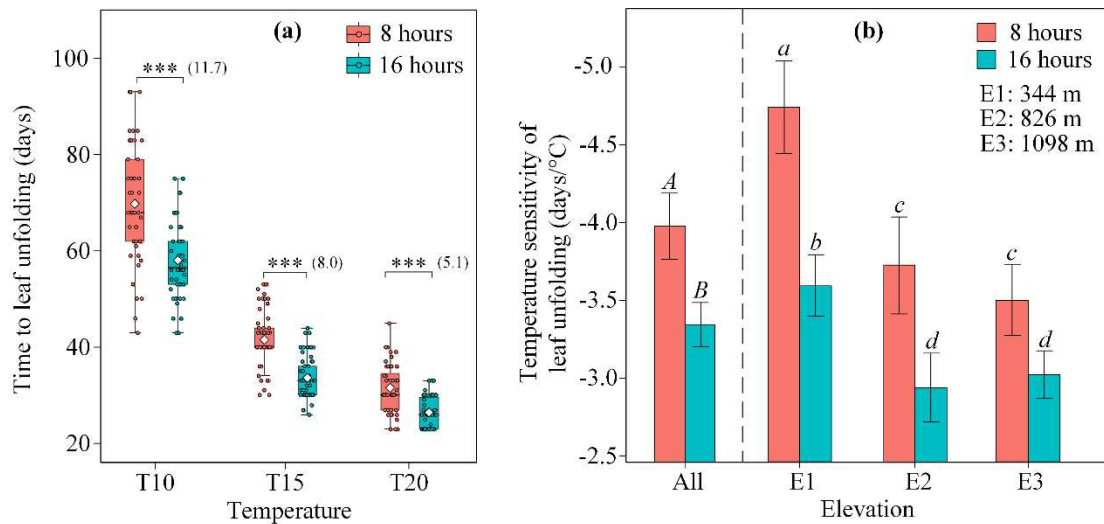
237 *3.2 Temperature and photoperiod effects on leaf unfolding*

238 Increases in the temperature and photoperiod significantly accelerated the leaf
 239 unfolding of all samples (Fig. 1e and g). Leaf unfolding occurred 36.1 and 26.7 days

240 earlier for the twigs that underwent the T20 and T15 treatments, respectively, compared
241 with the twigs that underwent the T10 treatment (Fig. 1e). Additionally, the heat
242 requirement for leaf unfolding was significantly lower for the T15 and T20 treatments
243 than for the T10 treatment (Fig. 1f). More specifically, the heat requirements were
244 13,345 °C and 13,445 °C at 15 °C and 20 °C, respectively, whereas it was 15,523 °C at
245 10 °C (Fig. 1f).

246

247 The time to leaf unfolding was shortened by 9.1 days following the long photoperiod
248 treatment, which corresponds to a 2,837 °C decrease in the heat requirement (Fig. 1g
249 and h). Interestingly, the long photoperiod significantly accelerated leaf unfolding for
250 all temperature treatments, but this photoperiod effect decreased as the temperature
251 increased. Compared with the effects of the P8 treatment, the P16 treatment shortened
252 the time to leaf unfolding by 11.7, 8.0, and 5.1 days during the T10, T15, and T20
253 treatments, respectively (Fig. 3a). Consequently, the temperature sensitivity of leaf
254 unfolding decreased from 3.98 days °C⁻¹ (P8 treatment) to 3.34 days °C⁻¹ (P16
255 treatment). Similar responses to the photoperiod were observed at each provenance (Fig.
256 3b). However, the photoperiod-related decrease in S_T was more pronounced for the E1
257 twigs (1.15 days °C⁻¹) than for the E2 (0.79 days °C⁻¹) and E3 (0.48 days °C⁻¹) twigs.
258 This suggests the constraining effect of the short photoperiod increased as the elevation
259 decreased.



260

261 **Fig. 3** Time (a) and temperature sensitivity (b) to leaf unfolding in the different
 262 temperature treatments under short (8 hours, P8) and long photoperiod (16 hours, P16)
 263 treatments. *** indicates significant difference between the two photoperiods ($P <$
 264 0.001). The numbers in the brackets refer to the differences of time to leaf unfolding
 265 between the two photoperiod treatments. The different letters in (b) denote significant
 266 differences ($P < 0.05$) in the temperature sensitivity between the two photoperiods.

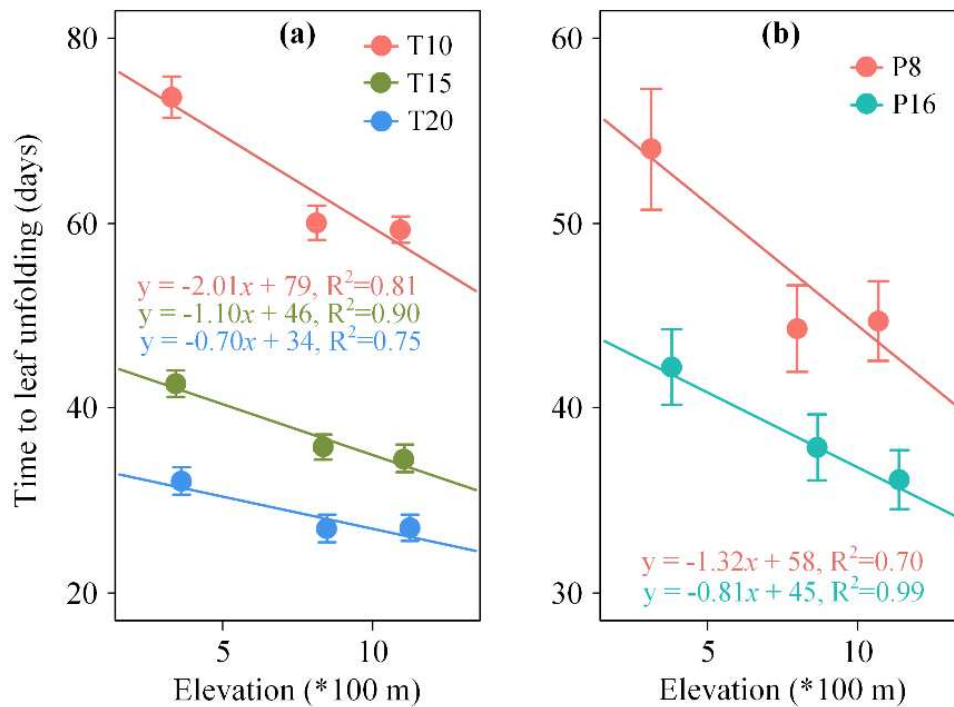
267

268 To clarify the potential mechanism mediating the leaf unfolding process, we examined
 269 the relationship between the forcing requirement (GDH) and chilling conditions
 270 (grouped according to the elevation). The chilling accumulation increased as the
 271 elevation increased, which further decreased the GDH required to induce leaf unfolding
 272 under both photoperiod treatments (Fig. S4).

273 3.3 Effect of the original elevation

274 The time to leaf unfolding was shortened by 2.0 days per 100 m increase in elevation
 275 for the T10 treatment, but by only 1.1 and 0.7 days for the T15 and T20 treatments,

276 respectively (Fig. 4a). Similarly, the long photoperiod also reduced the advanced rate
 277 per 100 m increase in elevation from 1.3 days to 0.8 days for P8 and P16, respectively
 278 (Fig. 4b).

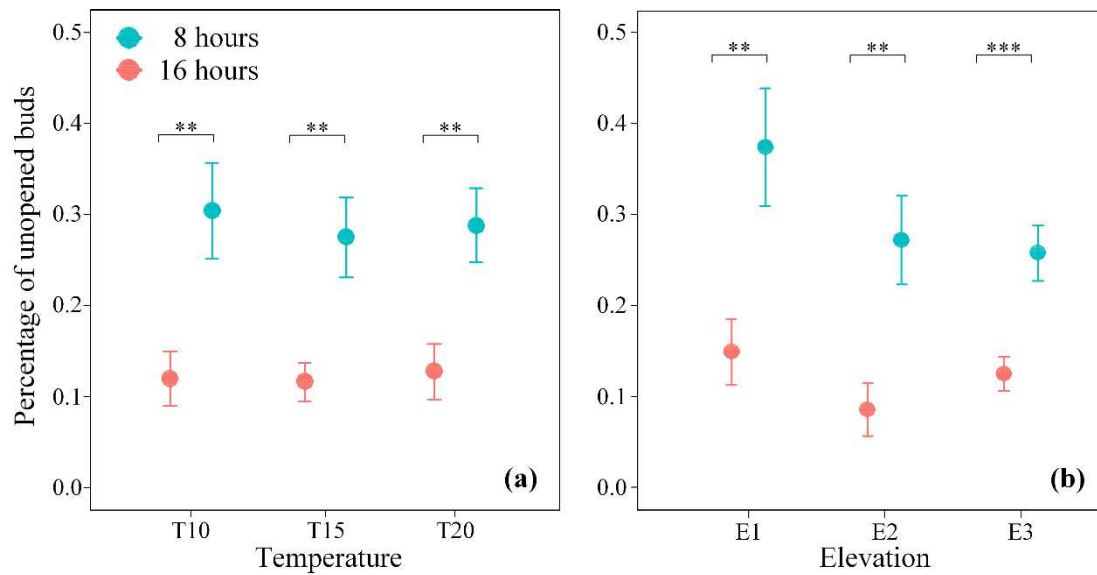


279
 280 **Fig. 4** Time to leaf unfolding under the different temperature and photoperiod
 281 treatments for the three elevation provenances. T10, T15 and T20 indicate the
 282 temperature treatments of 10 °C, 15 °C and 20 °C, respectively. P8, P16 represent the
 283 photoperiod treatments of 8 hours and 16 hours, respectively.

284

285 3.4 Effect of the photoperiod on twig vitality

286 The percentage of unopened buds per twig was significantly higher for the P8 treatment
 287 (29%) than for the P16 treatment (12%), irrespective of the temperature treatments and
 288 provenances (Fig. 5).



289

290 **Fig. 5** Unopened bud percentage under different temperature (a) and elevation (b)

291 treatments. ** and *** indicated significant difference existed between the two

292 photoperiods at $P < 0.01$ and $P < 0.001$ level, respectively.

293 4. Discussion

294 4.1 Effects of the temperature and photoperiod on leaf unfolding

295 Consistent with the findings of previous studies, we observed that temperature increases

296 significantly shortened the time to leaf unfolding by accelerating the accumulation of

297 the required heat (Flynn & Wolkovich, 2018; Fu *et al.*, 2019). We also determined that

298 the required GDH decreased with increasing temperatures, which is in accordance with

299 the findings of an earlier large-scale investigation involving a remote sensing dataset

300 (Fu *et al.*, 2015a). However, our results challenged the linearity between the GDH

301 requirement and leaf unfolding. Indeed, the heat accumulation rate may increase

302 exponentially in the low-temperature range, and plateau above a certain threshold, as

303 reflected by some phenological models (e.g., the sigmoid model; Chuine *et al.*, 2010).

304 Our data imply that the likelihood of such a relationship increases for ginkgo trees with
305 a stable heat requirement between 10 and 15 °C.

306

307 In addition to the temperature, the photoperiod reportedly regulates the spring
308 phenology of trees in temperate and boreal regions (David & Christian, 2014; Fu *et al.*,
309 2019; Way & Montgomery, 2015), with shorter photoperiods delaying bud development
310 and longer photoperiods accelerating leaf unfolding, especially under low chilling
311 conditions (Du *et al.*, 2019; Flynn & Wolkovich, 2018; Heide, 1993; Laube *et al.*, 2013;
312 Caffarra *et al.*, 2011). However, most of these studies were conducted using European
313 or American species. Accordingly, relatively little is known about photoperiod effects
314 on Asian species. In this study, consistent with the findings of the earlier related
315 research, we revealed that increases in the photoperiod shortened the time to leaf
316 unfolding of ginkgo. Interestingly, we also proved that the photoperiod effect differs
317 substantially among populations along an elevational gradient, with the greatest
318 photoperiod effect at the lowest elevation. This spatial variation in the photoperiod
319 effects on spring phenology is consistent with that reported by Zohner *et al.* (2016).
320 The differences in the responses of twigs to the photoperiod might be related to the
321 winter conditions at the collection sites, with the most photoperiod-sensitive species
322 generally originating from the regions with mild winters (Zohner *et al.*, 2016).
323 Additionally, the phenological sensitivity to temperature was highest for the twigs
324 collected at the lowest elevation, regardless of the photoperiod treatment, which is
325 consistent with the results of an earlier reciprocal transplant experiment involving seven

326 tree species in the European Alps (Vitasse *et al.*, 2013). Environment-induced local
327 adaptations may explain this phenomenon. Trees at relatively high elevations may have
328 adapted to be less responsive to temperature changes, enabling them to avoid the
329 adverse effects of a late frost in the spring (Firmat *et al.*, 2017; Vitasse *et al.*, 2010).

330

331 Our study provides evidence that bud development is promoted by increases in the
332 photoperiod, with an increase in the leaf unfolding rate, irrespective of the temperature,
333 which is in line with the results of previous studies (Du *et al.*, 2019; Zhang *et al.*, 2021;
334 Baumgarten *et al.*, 2021). For example, Zhang *et al.* (2021) demonstrated that a long
335 photoperiod can accelerate dormancy release and increase the bud burst rate, although
336 the photoperiod effect varies considerably among species (Zhang *et al.*, 2021; Fu *et al.*,
337 2019). If the temperature continues to increase, the restrictive effects of daylength on
338 leaf unfolding will likely increase, especially among plants distributed in the warm
339 region, as revealed by a recent remote sensing-based investigation (Moon *et al.*, 2021).
340 Therefore, the interaction between the photoperiod and temperature should be analyzed
341 more precisely and quantitatively to clarify ecosystem responses to climate change
342 (David & Christian, 2014; Flynn & Wolkovich, 2018; Zhang *et al.*, 2021; Fu *et al.*,
343 2019).

344 *4.2 Effects of gender and elevation on spring phenology*

345 We detected leaf unfolding dates are slightly earlier on the male twigs than on the
346 female twigs. Previous studies have found that compared with male ginkgo trees, the

347 spring phenological events of female ginkgo trees are usually delayed by about 3–5
348 days (Sofi, 2018; Wang *et al.*, 2009). To increase the chances of a successful pollination,
349 pollen maturation and dispersal should ideally occur after the ovules of female trees
350 have formed (Wang *et al.*, 2009). Since the timing of flowering and leaf unfolding is
351 synchronized, earlier leaf unfolding may lead to a more efficient pollination and
352 reproduction. Additionally, the allocation of resources for reproductive activities in
353 females (Lloyd & Webb, 1977) may result in delayed leaf development. However, we
354 observed that almost all of the buds on the examined twigs lacked ‘flowers’ (strobiles),
355 which may explain the small difference (only one day) in the leaf unfolding between
356 the two genders. Interestingly, we found same response of leaf unfolding to warming,
357 photoperiod and elevation between male and female, suggesting that leaf unfolding
358 response to environment change are synchronized between male and female. These
359 phenomena are likely related to the reproduction strategy of dioecious plants to ensure
360 a higher overlap in the male and female reproductive periods. Because this study
361 involved only one dioecious species, future investigations will need to include more
362 species to analyze the gender-related phenological responses to ongoing climate
363 changes.

364

365 Previous studies demonstrated that leaf unfolding is enhanced as the elevation decreases
366 under natural conditions, primarily because of the temperate lapse rate (Dittmar &
367 Elling, 2006; Vitasse *et al.*, 2009). In our climate chamber experiments, leaf unfolding
368 occurred earlier on the twigs collected at higher elevations than on the twigs collected

369 at the lowest elevation. This may be related to the insufficient chilling conditions at the
370 lowest elevation when samples were collected. Indeed, sufficient chilling can
371 effectively decrease the GDH requirement in temperate and subtropical regions (Du *et*
372 *al.*, 2019; Flynn & Wolkovich, 2018; Laube *et al.*, 2013; Baumgarten *et al.*, 2021). It is
373 likely that the relatively cold conditions at high elevations are sufficient for decreasing
374 the amount of heat required for flushing. Moreover, to maximize the growing season,
375 the environmentally induced adaptive plasticity of spring phenology at high elevations
376 may further decrease the GDH requirement (Firmat *et al.*, 2017; Vitasse, Bresson,
377 Kremer, Michalet, & Delzon, 2010). Therefore, plants growing at high elevations are
378 likely to have a relatively long growing season in response to a warming climate
379 (Vitasse *et al.*, 2010).

380

381 Interestingly, the temperature sensitivity decreased as the elevation increased, which is
382 consistent with the findings of an earlier study (Vitasse *et al.*, 2013), but in contrast to
383 the results of other studies (Chapman, 2013; Gao *et al.*, 2020). We assumed that the
384 chilling requirement can be fulfilled more easily at high elevations, leading to an
385 increase in the sensitivity of leaf unfolding to the temperature (Du *et al.*, 2019; Flynn
386 & Wolkovich, 2018; Laube *et al.*, 2013). However, we observed that although ginkgo
387 leaf unfolding responded quickly to the temperature, the temperature sensitivity
388 decreased along the elevation gradient, similar to the results of earlier research in
389 eastern Europe (Vitasse *et al.*, 2013; Gao *et al.*, 2020). This negative relationship may
390 be attributed to the decrease in spring radiation as the elevation increases (Gao *et al.*,

391 2020), but this obviously cannot explain our experimental results. Alternatively,
392 increasing temperature variances at high elevations may help to explain our results
393 (Lechowicz, 1984; Wang *et al.*, 2014). Trees growing in area with larger magnitude of
394 temperature variance such as high-elevation may have adapted to the unstable
395 temperature conditions and therefore have lower temperature sensitivity to warming
396 (Lechowicz, 1984; Wang *et al.*, 2014). In addition, the temperature sensitivity is
397 dependent on both the changes in heat accumulation and the time, although the heat
398 requirement for budburst is larger at the low elevation, the warm temperature may fulfill
399 the requirements with a short time period, and thus result in a higher temperature
400 sensitivity at the low elevation. Furthermore, an active selection for reducing
401 temperature sensitivity or a more stringent interaction with photoperiod in high
402 elevation might further minimize the damage risk induced by late spring frosts (Vitasse
403 *et al.*, 2013). Because populations from the margins of warm and cold regions may
404 respond differently to environmental cues and warming, predicting phenological events
405 for these populations might be challenging. A limitation of the present study was the
406 use of twigs for the controlled analysis of different temperature and photoperiod
407 gradients. The constant temperature and photoperiod treatments during our experiments
408 do not accurately reflect natural conditions. This high level of experimental control did
409 enable better attribution of plant responses to specific environmental cues, however, but
410 should be supplemented in the future with experiments under more natural
411 environmental conditions to also reveal natural feedbacks (De Boeck *et al.*, 2015).

412 **5. Conclusions**

413 Our study revealed that the sensitivity of ginkgo leaf unfolding to the temperature and
414 photoperiod decreases as the elevation increases, possibly because of local adaptations
415 that protect against a late frost in the spring. Our experiment indicated that bud
416 development is mainly affected by the temperature, but other climatic conditions (e.g.,
417 chilling) and the photoperiod can significantly change the heat requirement for leaf
418 unfolding. Therefore, phenological models should account for population-specific
419 responses to the temperature by considering local adaptations, especially along
420 bioclimatic gradients (e.g., across elevations). Additionally, although the differences
421 between genders were small in this study, our results demonstrated the gender-
422 synchronous response of leaf unfolding under changing environments and also provide
423 new insights into the phenological responses of dioecious plants to climatic variations.
424 Future research should focus on the interactive effects of the photoperiod and
425 temperature, the differential responses of genders, as well as gymnosperms and
426 angiosperms, and the environmental plasticity of plants to elucidate leaf phenological
427 responses to climate change.

428

429 **Author Contribution Statement**

430 Y.H.F. and Y.P.Z conceived the ideas and designed methodology; Y.P.Z and C.F.L
431 collected the twig at Tianmu Mountain; Z.W., S.W and Y.G collected the data of leaf
432 phenology; Z.W. and Y.H.F. analyzed the data and led the writing of the manuscript, in

433 corporation with Y.P.Z and C.F.L; J.T., H.D.B. and Y.V. advised on experimental
434 method, research design and assisted in writing the manuscript; Z.W. and C.F.L
435 contributed equally to this work. All authors contributed critically to the drafts and gave
436 final approval for publication.

437 **Data availability statement**

438 The datasets that support the findings of the current study are available from the
439 corresponding author on reasonable request.

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