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1 **Occurrence and patterns of toxic metals in Mangrove Forests from the Oman Sea, Iran**

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14

15 **Abstract**

16 Concentrations of selected toxic metals were investigated in roots, stems, leaves and sediments
17 from mangrove forests situated along the coast of the Oman Sea, Iran. Results showed that the
18 overall average concentrations of lead, nickel, copper, and zinc in sediments were 47.90, 54.12,
19 42.13 and 44 µg/g dry weight (dw) and 3.81, 16.41, 29.23 and 25 µg/g dw in plant tissues,
20 respectively. In addition, the bioconcentration factors (BCFs) of root, stem and leaf ranged from
21 0.5 to 1.7, 0.2 to 1.5, and 0.4 to 1.3, respectively. Calculated bioconcentration factors showed that
22 all plant tissues were able to uptake copper from the sediments, making them suitable biological
23 indicators for this metal. Similarly, the roots were found a suitable indicator for nickel and lead,
24 while leaves and stems were better indicators for zinc contamination. Pollution indices showed
25 that the sediments of mangrove forests along the coast of the Oman Sea were in the low ecological
26 risk category (risk index < 150), and that all investigated sites were in the category of low to
27 moderate pollution (pollution load index: 1.5-0.11), with a 21% probability of biological toxicity.

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29 **Key words:** Mangrove ecosystem, Toxic metals, geochemical indicators, bioavailability, Iran

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39 **1. Introduction**

40 Toxic metals are natural earth elements; in trace amounts, certain metals are necessary to support
41 life but in larger amounts they may build up in biological systems and become a significant health
42 hazard. As non-degradable and widespread contaminants, toxic metals can enter water bodies and
43 consequently the food web, causing adverse effects on the aquatic environment and finally human
44 health (Das et al., 2014; Shah, 2021).

45 Mangroves are plants or shrubs typically growing on low-sloping banks with fine-grained
46 sediments. Mangrove trees have been shown to be able, to some extent, to accumulate certain
47 (toxic) metals. This was evidenced in a previous study on toxic metals in the mangrove trees of
48 Gowater bay, Chabahar, southeastern coast of Iran, where nickel was found in mangrove plant
49 roots, likely resulting from the accumulation of this metal naturally present in ophiolite stones in
50 the beach bed. Conversely, the presence of other toxic metals, like cadmium, copper and zinc, in
51 other plant tissues was the result of accumulation from anthropogenic sources like upstream runoff
52 (Einollahipeer, 2012). Due to this accumulation potential, mangrove forests play an important role
53 in aquatic ecosystems, making them suitable to be used as biological indicators (Smical et al.,
54 2008). To a certain extent, mangroves can tolerate an uptake of metals. This is because they possess
55 aerial roots (i.e. pneumatophores), which allow for gas exchange in soggy soils, and facilitate the
56 uptake of oxygen, helping to alleviate potential negative effects of metal toxicity. In addition,
57 mangroves can sequester and store metals within certain tissues (e.g. roots) which helps protecting
58 the vital metabolic processes and physiological functions of the trees. Mangroves also possess
59 several detoxification mechanisms to mitigate the harmful effects of metals, e.g. by producing
60 metal-binding ligands or through their enzymatic system (Kumari& Rathore, 2021; Alongi, 2021;
61 Sruthi et al., 2017), and are known to benefit from symbiotic associations with microorganisms in
62 the root systems which contribute to metal tolerance and detoxification by facilitating metal
63 immobilization, precipitation, or transformation into less toxic forms (Harguinteguy et al., 2014;
64 Sawidis et al., 2011; Sarwar et al., 2017; Salam et al., 2016). Finally, the mangrove physiological
65 adaptations which allow them to maintain water and ion balance in saline environments can help
66 mitigate the toxic effects of metals.

67 Studies conducted on marine ecosystems by Arumugam et al (2018) show that the concentrations
68 of toxic metals in the sediments of mangrove forests were 3 to 5 times higher than in the
69 surrounding water, and that the sediments of mangrove forests in tropical and subtropical regions

70 have a high potential to store toxic metals (Shi et al., 2019). This shows the importance of such
71 ecosystem in maintaining and possibly restoring the environmental quality of the coastline
72 (Baharvand et al., 2022). For these reasons, monitoring the concentrations of metals in coastal and
73 mangrove forests and evaluating their environmental quality can be considered an essential
74 management tool to protect these ecosystems (Zhang et al., 2022).

75 Due to their richness in biodiversity, strategic location at the threshold of the ecological range of
76 environmental conditions, and sensitivity to pollution, Iranian mangrove forests are highly
77 environmentally relevant study habitats (Meena, 2018). Among them, the mangrove forest of the
78 Oman Sea, located between latitude 25° 11' N and 27° 52' N, is considered a special ecosystem
79 due to its a rich diversity of plant and animal life. Unfortunately, in the last few decades, it has
80 been exposed to several anthropogenic stressors, including oil extraction and refinery and urban
81 sewage discharge, which threaten its biodiversity (Ghayoumi et al., 2019). Because of this, this
82 area has been the object of a few studies monitoring the metal contamination, but the quality and
83 quantitative assessment of its ecological status has been so far poorly researched (Einollahipeer et
84 al., 2012; Pakzadtoochaei et al., 2013).

85 This study aimed at investigating the presence and concentrations of trace metals, including
86 copper, zinc, nickel, and lead, in several mangrove plant tissues and relative surface sediments
87 collected from this Iranian area. The ability of mangroves to uptake metals from the sediments and
88 transfer them to different plant tissues was also explored. Finally, various ecological risk
89 assessment indicators were used to investigate the current environmental quality of the study area.

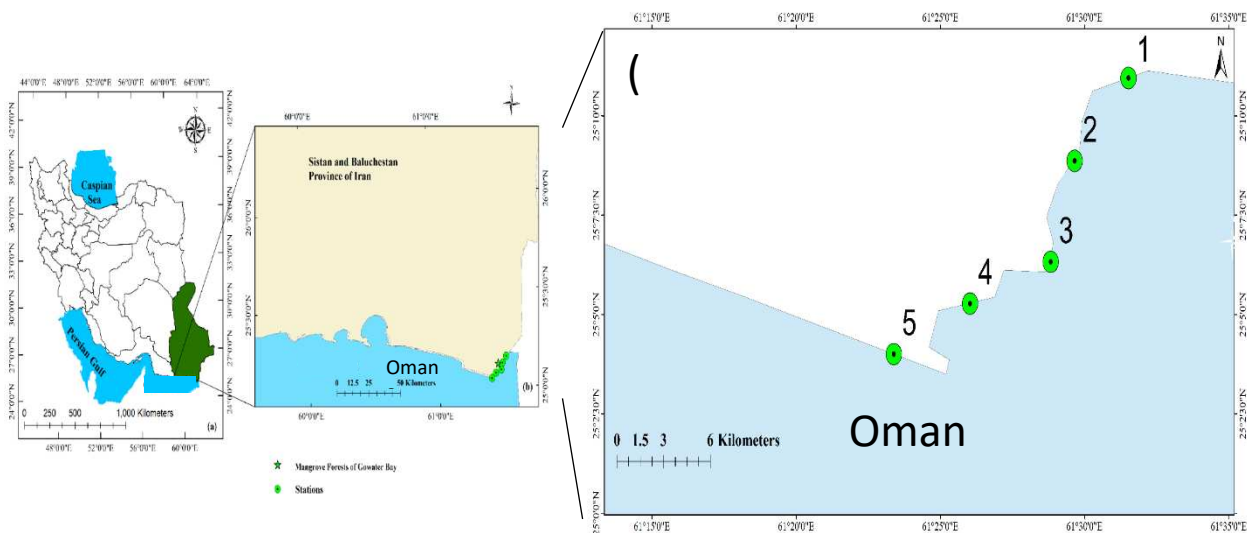
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91 **2. Materials and methods**

92 **2.1 Sampling area**

93 Gowater protected area is located on the northern shores of the Oman Sea, in the Sistan and
94 Baluchestan province, Iran (Zahed et al., 2010). The sampling area included five sites in the
95 mangrove forests between the port city of Chabahar and the protected area of Gowater, which has
96 the highest density of mangrove trees and is representative of all mangrove forests in this area (Fig.
97 1). This area is characterized by a dry and desertic climate, with average annual rainfall below 150
98 mm, an average monthly temperature of 27.5 °C (which fluctuates from a minimum of 21.2 °C in
99 January to a maximum of 32.6°C in June), a monthly average relative humidity of 70%, and a

100 prevalent wind direction from the southwest. The mangrove forests in the southeast of Iran extend
101 from the Sarbaz River and continue to the coasts of the Oman Sea (Dahmardeh bBhrooz, 2022).
102 Sampling points were chosen to represent the entire region and different habitat conditions,
103 spanning from inland to coastal ecosystems.



104
105 Fig 1. Location of sampling stations in the mangrove forests of Gowater Bay in the Oman Sea,
106 Iran

108 2.2 Sample collection

109 Only "true mangroves" were considered for inclusion, defined by their adaptation to intertidal
110 environments, salt regulation mechanisms, and taxonomic separation from terrestrial elements
111 (Tomlinson, 1986). Samples of sediment (n=15), roots (n=15), stems (n=15) and leaves (n=300)
112 of *Avicennia marina* were collected in triplicate from the above-mentioned five sites in spring
113 2022. At each sampling site, the surface sediment layer (about 500 g) was collected with a plastic
114 shovel at 10 cm depth. For root tissue sampling according to the method of Lawton et al. (1981),
115 sampling was done from the nutritious roots of the mangrove plant, and the harvesting of larger
116 respiratory roots was avoided (Lawton et al., 1981). Following the methods of Lindsey et al. (2005)
117 and MacFarlane et al. (2003), 20 leaves were collected from each plant and with three replicates
118 at each site, carefully separating them from the petiole by horticultural scissors (Lindsey et al.,
119 2005; MacFarlane et al., 2003). These samples were selected from 5 to 10 trees taller than 3 meters
120 and in such a way to cover the whole tree crown. Finally, the stem samples were obtained from the

121 transverse and thin sections of the stem tissue with a diameter of 4 millimeters by pruning shears
122 (Davari et al., 2010).

123 All plant tissues were collected from trees having a healthy appearance, with no signs of disease
124 or pest activity on the leaves. Sediment and roots were collected in the vicinity of the same marked
125 tree. After collection, all samples were placed in zip-lock polyethylene plastic bags, transported to
126 the laboratory, and kept at -20°C.

127

128 **2.3 Sample preparation and analysis**

129 Once at the laboratory, sediment samples were dried in an oven at 80 °C for 3 days, pulverized,
130 sieved with a 63 µm mesh steel sieve to separate waste materials and coarser particles, transferred
131 in pre-coded zip-lock bags and stored in the dark at room temperature until analysis (Hashim &
132 Nazli, 2010). Plant tissues, including root, stem, and leaf samples, were carefully rinsed with
133 distilled water, dried in an oven at 60 °C for 24 hours until the weight of the samples reached a
134 constant value, pulverized, and finally stored in the dark at room temperature until analysis.

135 Sediment samples (1 g) were extracted by acid digestion through addition of a mixture of 65%
136 nitric acid and concentrated perchloric acid (4:1, v/v), and kept at a temperature of 140 °C for 2
137 hours and then repeated for 3 hours. After digestion, the samples were filtered by Whatman 42 µm
138 filter paper and finally diluted with double distilled water to a volume of 50 mL (Abdul-Wahab
139 and Jupp, 2009). Plant tissues (1 g) were digested with 10 mL of 65% nitric acid and hydrogen
140 peroxide (4:1, v/v) at 90 °C for 2 hours on a hot plate. After cooling at laboratory temperature, the
141 digested samples were filtered with 0.45 µ filter paper and diluted with double distilled water to a
142 volume of 20 mL (MacFarlane et al., 2007).

143 The concentrations of copper, nickel, lead and zinc in sediment and plant samples were measured
144 using a Konic atomic absorption device model NOVAA 300 and expressed in µg/g dry weight
145 (dw).

146 The sediment physicochemical properties of the study sites are presented in Table 1. The texture
147 of the sediment samples among different sites was sandy clay loam (sites 2, 4 and 5), clay loam
148 (site 1), and loam (site 3), with pH varying between 2.45 and 3.25 in all sites. Sediment acidity
149 may also have resulted from decomposition of mangrove litter. All sediment samples had medium
150 cation exchange capacity (CEC). The lowest and highest average content of organic matter in the

151 samples was 4.76 and 11.10%, respectively. The values of organic carbon ranged from 1.07 to
 152 6.43% with the lowest and the highest values recorded at sediments of study area.

153

154 Table 1: Physiochemical characteristics in mangrove sediments at the surface soil.

Sites	pH	OM (%)	OC (%)	CEC	% Soil composition			Soil Texture
				(cmol/kg)	Sand	Silt	Clay	
S1	3.15	9.20	5.38	24.15	34.08	36.50	29.57	clay loam
S2	3.25	8.12	4.35	9.19	58.05	15.96	26.05	sandy clay loam
S3	2.97	9.37	5.96	20.27	43.58	27.80	28.61	loam
S4	2.45	11.10	6.43	18.76	52.90	17.40	29.65	sandy clay loam
S5	2.97	4.76	1.07	9.69	64.95	8.35	26.82	sandy clay loam

155 OM: organic matter; OC: organic carbon

156

157 Three measures of metal uptake were used for interspecific and intraspecific comparisons: root
 158 bioconcentration factor (BCF), leaf BCF, and transfer factor (TF). However, when combining data
 159 from different studies, validity issues arise. Metal uptake may vary due to sediment conditions
 160 such as anoxic sediments with high sulfur and organic content, which can reduce metal
 161 bioavailability (Harbison, 1986). Metal availability is influenced by sediment factors like cation
 162 exchange capacity, pH, redox status, metal speciation, nutrient availability, and salinity. Limited
 163 data availability for highly contaminated areas can skew observed patterns (Greger, 2004).
 164 Varying sample sizes within and across studies can also impact interpretation by potentially
 165 including anomalous data.

166

167 2.4 Quality Control

168 Instrument calibration was performed with a NIST-traceable std solution (AccuTrace Single
 169 Element Standard; AccuStandard Inc., New Haven, CT, USA). The precision and accuracy of the
 170 applied analytical method were determined by means of seven replicate analyses of standard
 171 reference materials SRM 1633b (constituent elements in coal fly ash), SRM 2709 (San Joaquin
 172 soil baseline trace element concentrations), and SRM 2711 (Montana II soil). Blank samples were
 173 prepared as the samples but without matrix and average blank levels per batch were subtracted
 174 from the sample results, and a value equal to 3 times the standard deviation of the blank
 175 measurement was used as the limit of quantification (LOQ). For compounds absent in the blanks,
 176 LOQs were based on a signal/noise ratio of 10 (S/N = 10). LOQs were 0.09, 0.06, 0.05, and 0.10
 177 $\mu\text{g/g dw}$ in Cu, Ni, Pb, and Zn respectively. In each sample batch, procedural blanks and SRMs

178 were included. The certified values for the reference materials amounted to Zn = 86 ± 2.5, Pb =
179 0.23 ± 0.3, Cu = 21.8 ± 5, and Ni = 1.6 ± 0.12, and the certified values for the used material
180 amounted to Zn = 89 ± 60, Pb = 0.24 ± 0.4, Cd = 22.58 ± 3, and Ni = 1.7 ± 0.13 (6 replicates with
181 recoveries between 88 and 105% and a relative standard deviation (RSD) of 6%).

182

183 **2.5 Statistical Analysis**

184 In this study, SPSS version 19 software was used for statistical analysis. At first, the normality of
185 the data was checked using the Kolmogorov Smirnov test, and after confirmation, one way
186 ANOVA and Tukey statistical tests were used to compare the mean and differences between
187 selected metal concentrations.

188

189 **2.6 Indices**

190 The following indices were used to assess the environmental quality and metal pollution status of
191 the investigated ecosystem.

192 The *bioconcentration factor* (BCF) is the ratio between the concentration of a toxic metal in a
193 living organism and in a non-living environment (water and sediment). Species with BCF > 1 can
194 be considered as element-stabilizing species (Almahasheer, 2019).

195 The *metal transfer factor* (TF) is used to evaluate the ability of the mangrove plant to transfer
196 metals from the underground tissues (roots and rhizomes) to the upper ones (stems and leaves) via
197 the ratio between metal concentration in aerial tissues and in the roots (Hilmi et al., 2023). The
198 higher the rate of TF, the faster the ecosystem purification process happens and the coastal and
199 marine micro-ecosystems are less exposed to pollution (ELTurk et al., 2018).

200 The *geochemical accumulation index* (Igeo), introduced by (Muller, 1969), is a common method
201 for estimating the intensity of contamination of sediments with toxic metals and is calculated based
202 on equation 1.

$$203 \text{Igeo} = \text{Log}_2 \frac{C_n}{1.5 \times B_n} \quad (1)$$

204 Where C_n is the measured concentration of a toxic metal in the sediments and B_n is the
205 concentration of the same element in the earth's crust (background concentration, or element
206 concentration in shale). The index goes from class 0 (unpolluted) to class 6 (strongly polluted,
207 where the values of the elements are at least 100 times the reference values) (Table S1).

208 The *contamination factor* (CF) provides a description of the pollution related to the investigated
 209 toxic elements and the pollution of the sediment environment (Hakanson, 1980). More specifically,
 210 the CF is derived using equation 2:

$$211 \quad CF = \frac{C_i}{C_n} \quad (2)$$

212 Where C_i is the concentration of the element in the sediments and C_n is the concentration of the
 213 same element in shale sample (Table S2), (Hakanson, 1980).

214 The comprehensive ecological risk assessment of selected toxic metals in sediments is determined
 215 using the *potential ecological risk index* (E_{ir}^i) (Hakanson, 1980) and it is calculated according to
 216 equation 3.

$$217 \quad E_{ir} = T_r \times C_f \quad (3)$$

218 Where C_f is the contamination factor and T_r is the toxicity coefficient, whose values are 1, 5, 5,
 219 and 5 for zinc, lead, copper, and nickel metals, respectively (Table S3).

220 Further, the *risk index* (RI) is determined as the sum of E_{ri} (equation 4) and generally indicates the
 221 sensitivity of living organisms to toxic metals and the environmental risks associated with toxic
 222 metal pollution (Kusin et al., 2018).

$$223 \quad RI = \sum_{i=1}^n E_r \quad (4)$$

224 The sediment quality is also calculated through the *pollution load index* (PLI) based on equation
 225 5 (Tomlinson et al., 2014).

$$226 \quad PLI = [CF_1 \times CF_2 \times CF_3 \times \dots \times CF_n]^{\frac{1}{n}} \quad (5)$$

227 Where n is the number of metals (=4, i.e. lead, nickel, copper and zinc), CF_n is the contamination
 228 factor of a metal (see equation 2). PLI indicates how many times the metal content in sediments
 229 exceeds the natural background concentration of metals and is a cumulative indication of the
 230 overall level of toxicity of a sample. The classification of the PLI index is as follows: $PLI < 1$:
 231 uncontaminated; $PLI \geq 1$: contaminated.

232 Finally, the *mean probable effect level quotient index* (mPELq) is used to measure the biological
 233 effect of toxic metals on mangrove shrubs (Long et al., 2006) (equation 6).

$$234 \quad mPELq = \frac{\sum_{m=1}^n n \left(\frac{c_m}{PEL_m} \right)}{n} \quad (6)$$

235 Where c_m is the metal concentration in the sediments, PEL_m values for zinc, lead, copper and
 236 nickel metals are 270, 110, 110 and 50, respectively, and n is the number of metals (Table S4).

237

238 **3. Results and Discussion**239 **3.1 Concentrations of metals in sediments**

240 The content of trace metals in the sediment samples collected in 5 sites of the Oman Sea mangrove
 241 forests are showed in Table 2, together with a comparison of concentrations measured in this study
 242 and in other studies worldwide.

243

244 **Table 2. Content of trace metals in the sediments from the Oman Sea mangrove forests (($\mu\text{g/g}$) (mean \pm SD)
 245 and in other studies worldwide and in international standards**

Sites	Zn	Pb	Ni	Cu	Ref
S1 (n=3)	47.45 \pm 19.45	52.25 \pm 1.95	60.20 \pm 0.95	49.10 \pm 0.95	
S2 (n=3)	30.80 \pm 3.90	26.30 \pm 3.70	49.15 \pm 0.95	49.90 \pm 3.60	
S3 (n=3)	58.35 \pm 1.95	57.10 \pm 1.70	59.30 \pm 3.95	44.50 \pm 8.75	
S4 (n=3)	38.60 \pm 11.20	54.50 \pm 2.95	54.20 \pm 14.60	36.90 \pm 2.35	
S5 (n=3)	47.80 \pm 7.00	54.20 \pm 2.95	58.30 \pm 0.80	37.60 \pm 0.70	
Total	44.60 \pm 12.65	48.90 \pm 3.85	56.20 \pm 24.65	43.60 \pm 6.25	Prsented Study
Mangalavanam, India	139.15	27.91	41.34	33.70	Puthusseri <i>et al.</i> , 2021
Futian, South China	32.80	40.30	14.90	29.40	Wang <i>et al.</i> , 2013
Gabrik Creek (Jask), Iran	69.63	67.63	86.53	-	Zarezadeh & Rezaee, 2016
Klang, Malaysia	163.60	46.94	18.22	38.24	Yap & Al-Mutairi, 2022
Mahshahr, Iran	75.98	15.02	100.96	25.13	Cheraghi <i>et al.</i> , 2015
Sirik, Azini Creek, Iran	109.05	99.40	132.70	-	Zarezadeh <i>et al.</i> , 2014
American Sediment Quality Guidelines (NOAA)	150	46.70	20.90	34	ERL (Effect Range Low)
	410	218	51.60	270	ERM (Effect Range Medium)
Canadian Sediment Quality	120	16	31	16	LEL (Lowest Effect level)

Guidelines (SQGs)	820	75	250	110	SEL (Severe Effect level)
New York Sediment Quality Guidelines	120	32	16	16	LEL (Lowest Effect level)
	270	110	50	110	SEL (Severe Effect level)

246

247 Among all analyzed samples, sediments were the most contaminated with toxic metals, with the
 248 following pattern: nickel (56.22 µg/g dw), lead (48.86 µg/g dw), zinc (45 µg/g dw), copper (43.62
 249 µg/g dw). The slightly higher concentration of nickel in sediments compared with the other metals
 250 was likely caused by anthropogenic sources represented by traffic of ships, boats and tankers, crude
 251 oil, and urban and industrial wastewater (Vieira et al., 2008), and by runoff from the upstream
 252 rivers of this area. In addition, the presence of nickel in sediments can be directly related to the
 253 type of bed and the prevailing morphological conditions, like the proximity of the estuary and its
 254 shallow depth (Zarezadeh et al., 2017). Similarly, lead contamination in the analyzed sediments
 255 might have derived from its presence in gasoline of ships and boats, and from runoff of inland car
 256 traffic (El Tokhi et al., 2008).

257 The solubility and accumulation of metals in mangrove sediments and tissues are influenced by
 258 various factors, and caution should be exercised when comparing findings to other studies. Oxygen
 259 exuded by roots fixes iron (Fe) and co-precipitates metals as oxyhydroxides in the rhizosphere,
 260 reducing trace metal availability and mobility (de Lacerda et al., 2022). Natural processes, like sea
 261 level rise, erosion, saline intrusion, tidal forcing, sediment remobilization, porewater salinization,
 262 sulfide oxidation, and metal release affect metal dynamics. (Aragon and Miguens, 2001; Lacerda
 263 et al., 1988; Nguyen et al., 2020). Metal-chloride complex formation, sulfate reduction, and
 264 changes in rainfall and environmental stress can increase metal bioavailability and toxicity
 265 (Lacerda et al., 1988). Suspended particles and particulate metals are transported by floods to the
 266 continental shelf, decreasing metal bioavailability in sediments (Nguyen et al., 2020).
 267 Acidification increases the solubility of trace metals by dissolving carbonates. These factors
 268 influence the solubility, availability, and toxic effects of metals in mangrove ecosystems.

269 Previous studies have shown that mud sediments can be good accumulators for both organic and
 270 inorganic pollutants due to the larger ratio between surface and volume of the particles. Because
 271 most sediments of mangrove forests are made of silt and clay, which are very small in size, their
 272 relatively high concentration of metals is justified (Zahed et al., 2010). Still, such concentrations

273 were in the same order of magnitude as measured in India (Puthusseri et al., 2021), lower than in
 274 other Iranian locations (Zarezadeh et al., 2014; Zarezadeh & Rezaee, 2016), and higher than in
 275 China (Wang et al., 2013). In addition, to determine the degree of metal contamination of the
 276 surface sediments of the studied area, their average concentrations were compared with
 277 international sediment quality standards including the American Sediment Quality guidelines
 278 (NOAA), the Canadian Sediment Quality guidelines (SQGs), and the New York Sediment Quality
 279 guidelines, which can be used to classify polluted sediments and predict the possibility of adverse
 280 biological effects in aquatic organisms that are in contact with these sediments (Table 1). The
 281 NOAA categorizes the level of pollution in effect range low (ERL) and effect range medium
 282 (ERM), the Canadian SQGs and the New York sediment quality standard express the pollution
 283 levels as lowest effect level (LEL) and severe effect level (SEL) (Yazdan Panah et al., 2019) (Long
 284 et al., 1995). The average concentration of nickel in the sediments from the Coasts of the Oman
 285 Sea mangroves (56.22 µg/g) resulted slightly above the ERM limit for this element (51.60 µg/g)
 286 and the SEL according to the New York sediment quality standard (50 µg/g), but lower than the
 287 SEL based on the Canadian SGQ (250 µg/g). Also the average concentrations of lead (48.86 µg/g)
 288 and copper (43.62 µg/g) were higher than ERL and LEL levels but lower than SEL. Finally, the
 289 average concentration of zinc (44.60 µg/g) was lower than all standards levels.

290

291 3.2 Concentrations of metals in plant tissues

292 The concentrations of lead, nickel, copper, and zinc in plant tissues (root, stem and leaf) are
 293 presented in Table 3.

294 **Table 3. Content of trace metals in tissues of stem, root and leaf of *A. marina* (µg/g) Mean±SD and in**
 295 **other studies worldwide.**

296

Sites	Root				
	Zn	Pb	Ni	Cu	
S1 (n=3)	26.xx±2.44	10.31±1.16	40.22±0.97	26.37±4.39	
S2 (n=3)	28.xx±3.87	11.28±3.62	38.52±0.94	27.94±3.50	
S3 (n=3)	9.xx±4.54	9.20±0.27	39.00±1.74	24.71±1.39	
S4 (n=3)	11.xx±3.90	10.17±0.74	40.27±3.47	26.37±0.64	
S5 (n=3)	26.00±0.63	11.70±1.28	42.78±0.21	27.53±1.14	
Total	20.xx±7.35	10.53±1.84	40.15±4.30	26.58±2.59	Prsresent Study
	Stem				
S1 (n=3)	43.xx±7.99	1.12±0.36	7.88±0.97	28.20±0.92	
S2 (n=3)	18.xx±1.84	1.66±0.16	8.85±0.97	25.65±0.94	

S3 (n=3)	16.xx±2.21	1.34±0.16	9.91±0.89	24.50±0.85	
S4 (n=3)	23.xx±3.90	0.83±0.11	5.67±0.82	24.25±0.91	
S5 (n=3)	14.xx±2.51	1.38±0.42	3.44±1.36	25.20±1.17	
Total	23.xx±11.58	1.17±0.43	7.15±2.67	25.56±1.74	Prsésent Study
Leaf					
S1 (n=20)	31.xx±4.72	0.99±0.24	2.60±0.68	33.57±7.67	
S2 (n=20)	21.xx±3.97	0.97±0.12	2.69±0.77	39.93±22.91	
S3 (n=20)	24.xx±4.60	0.93±0.12	2.27±0.69	35.94±21.80	
S4 (n=20)	51.xx±9.44	0.91±0.17	1.98±0.94	60.43±6.01	
S5 (n=20)	49.xx±6.49	0.91±0.18	3.17±0.68	24.18±1.33	
Total	35.xx±13.90	0.94±0.17	2.54±0.81	38.81±17.69	Prsésent Study
Root					
Port Harcourt, Nigeria (<i>A.marina</i>)	6.52	3.28	1.76	4.82	Ubong <i>et al.</i> , 2018
Hawks Bay Karachi, Pakistan (<i>A.marina</i>)	23.34	-	-	5.25	Siddiqui & Saher, 2015
Carambolim, India (Macrophyte)	307.8	6.71	0.68	25.68	Vardanyan and Ingole,2004
Stem					
Guanabara Bay, SE Brazil (<i>L.racemosa</i>)	26.70	3.38	-	-	Machado <i>et al.</i> , 2002
Natal, Brazil (<i>R. mangle</i>)	1.36	-	4.43	0.31	Silva <i>et al.</i> , 2006
Hainan Island, China (<i>Rhizophora apiculata</i>)	6.20	-	-	2.90	Qiu <i>et al.</i> , 2011
Guangdong, Province of China (<i>A.marina</i>)	6.48	-	2.17	1.33	Zheng <i>et al.</i> , 1998
Leaf					
Natal, Brazil (<i>R. mangle</i>)	0.46	-	2.04	0.94	Silva <i>et al.</i> , 2006
Peninsular Malaysia (<i>Sonneratia caseolaris</i>)	5.90	35.5	-	26.80	Nazli and Hashim, 2010

Tamil Nadu, India (<i>A. indicum</i>)	107.80	23.21	-	14.78	Agoramoorthy <i>et al.</i> , 2008
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297

298 The trend of accumulation of toxic metals in the plant roots was Ni (40.15 µg/g) > Cu (26.58 µg/g)
 299 > Zn (19.82 µg/g) > Pb (10.53 µg/g), different from the other plant tissues, where the trend Cu
 300 (25.56 and 38.81 µg/g), Zn (22.73 and 34.93 µg/g) > Ni (7.15 and 2.54 µg/g) > Pb (1.17 and 0.94
 301 µg/g) could be observed for stems and leaves, respectively.

302 From this, it appears that essential metals have higher concentrations in aerial tissues, while non-
 303 essential metals have higher concentrations in root tissues. Also, our study results showed that
 304 concentrations of lead and nickel in the roots were significantly higher than in the other plant
 305 tissues, while this was not observed for the other metals. This difference can be due to the fact that
 306 the roots are in direct contact with the sediments and thus can directly accumulate the metals. Due
 307 to the presence of reduction conditions, frequent tide-related floodings, high levels of organic
 308 materials and sulfides, and the fine-grained texture of mangrove sediments wetlands, sediments in
 309 particular are considered sink areas for toxic metals (Alharbi *et al.*, 2019). From here, metals can
 310 be absorbed by the roots and stored in their tissue or be absorbed and then transferred to aerial
 311 tissues. The surface absorption of elements by the root epidermis, the presence of root Casparian
 312 bands and the impenetrability of the wall of the wood vessels in the root may be among the factors
 313 influencing the elements' fate (Baharvand *et al.*, 2022). In addition, differences in metal
 314 concentrations in root and aerial parts of the plants may be due to differences in the physiological
 315 structure of the tissues (Zheng *et al.* (1998)). Roots are perennial and permanent plant organs and
 316 have a longer time to accumulate metals, while leaves are subject to seasonal fall (Zheng *et al.*,
 317 1998) (Kabata-Pendias and Pendias in 2001). For example, the slight increase in average
 318 concentrations of zinc and copper from roots to leaves, as opposed to the higher concentrations of
 319 lead and nickel in the roots than in the stems and leaves, might be because copper and zinc are
 320 essential trace elements, necessary to the correct functioning of the plant. This is in accordance
 321 with the study by Ingole and Vardanyan (2004) who showed that the concentrations of toxic metals
 322 in the tissues of saline plants in Sevan, Armenia and Carambolim, India, were higher in plant root
 323 and stem tissues and that the lowest concentrations of metals corresponded to non-essential
 324 elements (Ingole & Vardanyan, 2004). This was confirmed by other studies on mangrove forests
 325 showing that copper and zinc had the highest concentrations in the root and leaf tissues and that

326 they are both essential elements for plants (Shete et al., 2007) (Victorio et al., 2020) (Wozny and
327 Krzeslowska (1993).

328 Eynollahipeer et al. (2012) studies looked at toxic metal accumulation in mangrove sediments and
329 tissues in Goater Bay of Chabahr city. The general trend in metal accumulation patterns in
330 sediments and plant tissues were somewhat similar in the study of Eynollahipeer et al. (2012) and
331 the present study. In sediments, Ni showed the highest accumulation in both cases. In plant tissues,
332 Cu and Zn levels tended to be higher in leaves and stems, while Ni and Pb were higher in roots.
333 This suggests mangrove roots take up and concentrate certain metals like Ni and Pb from sediments
334 more readily. In the study of Eynollahipeer et al. (2012), Cd accumulated more readily in plant
335 tissues based on higher BCF values. In this study, we found higher transfer factors for Cu and Zn
336 from roots to shoots. Both studies showed low ecological risk from metal pollution based on risk
337 indices. However, in this research, a have higher potential toxicity based on a 21% probability was
338 estimated (Einollahipeer, 2012).

339 A comparison of selected metal concentrations in the roots, stems and leaves from mangrove
340 forests in the present study with results obtained from other similar studies worldwide is showed
341 in Table 2. The levels of copper from this study were generally higher than in Brazil (Silva et al.,
342 2006; Machado et al., 2002), China (Qiu et al., 2011; Zheng et al., 1998), Pakistan (Siddiqui &
343 Saher, 2015), and Nigeria (Ubong et al., 2018), but comparable to the concentrations measured in
344 India (Vardanyan and Ingole, 2004) and Malaysia (Nazli and Hashim, 2010). Concentrations of
345 nickel in the roots from this study were higher than in all other selected locations, but the levels of
346 nickel in the other tissues were comparable with results obtained worldwide. Conversely,
347 concentrations of lead were generally similar to or lower than average levels from other locations.
348 Finally, levels of zinc in the mangrove forests from the Oman Sea were lower than measured in
349 India (Vardanyan and Ingole, 2004; Agoramoorthy et al., 2008), comparable with levels in Brazil
350 (Machado et al., 2002) and Pakistan (Siddiqui & Saher, 2015) and higher than Nigeria (Ubong et
351 al., 2018), Brazil (Silva et al., 2006) and China (Qiu et al., 2011; Zheng et al., 1998).Glasby et al.
352 (2019) examined the impact of bushfires on estuarine wetlands, while the current study focuses on
353 toxic metals in mangrove forests. Both studies investigate how toxic metals or bushfires affect the
354 health, organisms, habitats, and overall ecological dynamics of their respective ecosystems.
355 However, they differ in their specific areas of focus. The present study concentrates on mangrove
356 forests, which are coastal wetland habitats dominated by mangrove trees, while Glasby et al.

357 (2019) examines estuarine wetlands, which are transitional zones between rivers and the sea. The
 358 present research emphasizes toxic metals from various sources, while Glasby et al. (2019)
 359 highlights bushfires caused by natural or human-induced factors (Glasby et al., 2023).

360 TM Glasby, PT Gibson, R Laird, D S Swadling, G West. 2023. Black summer bushfires caused
 361 extensive damage to estuarine wetlands in New South Wales, Australia. *Ecological
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364 3.3 Environmental quality assessment

365 Results of the calculations of the geochemical accumulation (I_{geo}), Contamination factor (CF),
 366 potential ecological risk factor (E^i_r), and the potential ecological risk index (RI) are reported in
 367 Table 4. The analysis of metals in sediment varies across studies due to different digestion methods
 368 and acids used. HF, which extracts metals from the silicate matrix, was not employed in any of the
 369 studies. Various extraction methods were used, including $H_2O_2 + HCl + HNO_3 + HClO_4$ (Silva
 370 et al., 1990), microwave digestion in $HNO_3 + HCl$, and hotplate digestion in $HNO_3 + H_2O_2$, as
 371 well as other acids such as $HNO_3 + HClO_3$, $HNO_3 + HSO_3$, (Che, 1999) and weakly bound
 372 fraction examinations with 0.1 N HCl (Chiu and Chou, 1991) or 0.1 M HCl (Lacerda, 1997).
 373 Recovery rates varied among metals when comparing different extraction methods. The choice of
 374 extraction method can impact estimates of sediment metal loadings, particularly affecting
 375 bioconcentration factors (BCFs) more than transfer factors (TFs). Caution is necessary when
 376 interpreting the data due to potential biases and variations caused by sampling protocols for
 377 vegetation collection and environmental factors like temperature, light, and inundation frequency,
 378 which influence metal uptake patterns (Greger, 2004).

379
 380 **Table 4. Averages of geo-accumulation (I_{geo}), Contamination factor (CF), potential ecological risk factor (E^i_r)**
 381 **and potential ecological risk (RI) indexes of the measured toxic metals *A. marina* sediments.**
 382

(I_{geo}) Index				
Sites	Zn	Pb	Ni	Cu
1	2.77	2.93	3.63	2.89
2	2.01	3.52	2.85	2.96
3	3.20	2.98	3.61	3.07
4	2.86	2.60	3	2.67
5	3.08	2.91	3.4	2.88
(CF) index				
Sites	Zn	Pb	Ni	Cu
1	1.40	2.21	2.98	1.88
2	1.23	2.80	2.78	2.02

3	2.40	2.10	2.95	2.24	
4	1.79	2.40	2.20	1.60	
5	2.30	2.05	2.50	1.81	
(E_rⁱ) index					
Sites	Zn	Pb	Ni	Cu	RI
1	1.28	4.18	2.89	7.27	26.33
2	1.90	3.37	2.76	6.85	30.22
3	1.79	3.83	2.10	6.64	28.35
4	1.30	4.26	2.05	7.57	31.11
5	1.55	3.43	2.91	6.89	27.55
Total	7.82	19.07	12.71	35.22	143.56
(PLI) and (mPELq) indexes					
Sites	PLI		mPELq		
1	0.45		0.87		
2	0.55		0.74		
3	0.37		0.63		
4	0.42		0.45		
5	0.40		0.42		

383

384 According to the calculated geo-accumulation index, sediments from the studied mangrove forest

385 could be classified as moderately to highly polluted. The intensity of contamination with these

386 metals followed the order: nickel (3.2) > lead, copper and zinc (2.98, 2.89 and 2.78, respectively).

387 A similar pattern of metal pollution was obtained based on the contamination factor (CF) for the

388 investigated metals in the region, for which the average values were between 1 and 3, classified as

389 low to moderate pollution status.

390 The results of the evaluation of the potential ecological risk factor (E_rⁱ) for Pb, Zn, Ni, and Cu were

391 low (E_rⁱ < 40) and exhibited a low risk. This was confirmed by the calculated ecological risk index

392 (143.5), representing a low ecological risk. Copper had the greatest influence on the value of the

393 index, and zinc had the least influence. This is generally in accordance with the results of another

394 study on toxic metal contamination in coastal sediments from the South Pars Special Economic

395 Zone where most of the investigated sites were classified in the low to medium risk category in

396 terms of ecological risk due to metal contamination (Haghshenas et al., 2017). Fu et al. (2014)

397 evaluated the concentration and ecological risk of mercury, arsenic, chromium, lead, zinc and

398 copper in the sediments of the Jialu river in China. The analysis of the E_rⁱ showed that, except for

399 cadmium, classified as high-risk, other metals were in low risk status (Fu et al., 2014). Liu et al.

400 (2014), investigated the E_rⁱ of chromium, copper, zinc, cadmium, arsenic, mercury and lead in

401 mangrove ecosystem sediments in south China and found that the sediments were in a relatively

402 severe ecological risk, especially due to the presence of mercury and cadmium (Liu et al. , 2014).

403 The pollution load index (PLI) of the selected metals was below 1 in all analyzed sample sites
 404 (Table 3), suggesting that the region can be classified as non-polluted. This is consistent with the
 405 results of Yu et al. (2011) and Suresh et al. (2012). Islam et al. (2015) investigated the PLI in urban
 406 river sediments in Bangladesh, and found it was > 1 , indicating a reduction of sediment quality
 407 and the contamination of the studied river sediments with toxic metals. This was likely attributed
 408 to the discharge in the river of a urban sewage. This difference with the current study shows that,
 409 although human activities and industrialization are spreading in this area, their effects are still not
 410 critical on sediment quality. However, due to the growing urban population and industrialization
 411 of the region, constant monitoring of the area is highly recommended.

412 Finally, to determine the possible biological effects of toxic metals in sediments, the mean probable
 413 effect level quotient index (mPELq) was calculated between 0.42 and 0.87, classified as low to
 414 moderately polluted with a 21% probability of biological toxicity (Table S4). The same index was
 415 calculated by Aljahdali and Alhassan (2020) to evaluate the possible biological effects of copper,
 416 zinc, cadmium, chromium, lead, nickel and cobalt in the coastal sediments of the Kaduna River,
 417 Nigeria. Their results showed that the studied area was in the high pollution class with 94%
 418 probability of biological toxicity. The elevated mPELq was attributed to the impact of human
 419 intervention in the catchment area caused by industrial activities and atmospheric depositions
 420 (Aljahdali, 2020 & Alhassan). Also Rastegari Mehr et al. (2020) investigated the mPELq index to
 421 evaluate the possible biological effects of mercury, nickel, zinc, copper, lead and chromium on the
 422 coastal sediments of the Musa estuary and found the mPELq index between 0.5-1.51, with a
 423 probability of biological toxicity of 49% (Rastegari Mehr et al., 2020).

424 Finally, the BCF and TF were calculated based on the concentrations of metals in the sediments
 425 and in the plant tissues (Table 5).

427 **Table 5. Bioconcentration Factor (BCF) and Transfer Factor (TF) of toxic metal in *A. marina*.**

Tissue	Zn	Pb	Ni	Cu
BCF in Root	0.88	0.75	0.77	1.05
BCF in Stem	0.95	0.52	0.43	1.03
BCF in Leaf	0.93	0.68	0.65	1.02
TF in Stem	1.15	0.40	0.47	1.04
TF in Leaf	1.50	0.58	0.45	1.20

428

429 To be biological indicators, tissue accumulation of metals should be sediment-dose dependent. Otherwise,
430 BCFs and TFs have very limited utility concluding biological indicator potential.. For zinc, leaf and stem
431 tissues are the best biological indicators, with BCF values higher than in roots. The TF results for
432 lead and nickel were < 1 , indicating that the state of accumulation and accessibility in the plant is
433 average. For zinc and copper, the calculated TF was > 1 , indicating that, after their absorption from
434 the environment by the roots, the metals were transported to the aerial parts of the plant.
435 MacFarlane et al. (2007) calculated the BCF for copper, lead and zinc in the same mangrove plant
436 species (*A. marina*), obtaining values > 1 , and concluded that the root tissue is a suitable
437 bioindicator for these metals. In the same study, the TF was determined for copper and zinc as 1.52
438 and 1.53. They considered the reduction of the metal transfer factor from the root to the plant as a
439 result of the type of metal consumption for the plant (MacFarlane et al., 2007).

440 **Conclusion**

441 This study found that the concentrations of nickel in sediments was slightly higher compared to
442 other metals, and this was likely due to anthropogenic sources such as ship traffic, crude oil, urban
443 and industrial wastewater, and runoff from upstream rivers. Lead contamination in sediments was
444 likely derived from ships, boats, inland car traffic, and gasoline. The accumulation of toxic metals
445 in plant tissues varied, with nickel having the highest concentration in roots, followed by copper,
446 zinc, and lead. Essential metals tended to have higher concentrations in aerial tissues, while non-
447 essential metals had higher concentrations in root tissues. The roots, being in direct contact with
448 sediments, accumulated higher levels of lead and nickel compared to other plant tissues. Mangrove
449 sediments were considered sink areas for toxic metals due to reduction conditions, frequent
450 floodings, high organic material and sulfide levels, and fine-grained texture. The concentrations of
451 nickel in the roots were higher in the studied area compared to other locations, but levels in other
452 tissues were comparable worldwide. The sediments in the studied mangrove forest were classified
453 as moderately to highly polluted based on the geo-accumulation index. The intensity of
454 contamination followed the order: nickel $>$ lead, copper, and zinc. The contamination factor
455 indicated low to moderate pollution status. The potential ecological risk factor and ecological risk
456 index suggested low ecological risk, with copper having the greatest influence on the index value
457 and zinc having the least. Other studies on metal contamination in coastal sediments have also
458 indicated low to medium ecological risk. The pollution load index indicated that the region can be

459 classified as non-polluted. The mean probable effect level quotient index indicated low to moderate
460 pollution with a 21% probability of biological toxicity. Plant tissues, especially roots, were
461 identified as good biological indicators for lead, nickel, and copper accumulation. Zinc
462 accumulation was higher in leaf and stem tissues. The transportation factor indicated that lead and
463 nickel had average accumulation and accessibility in plants, while zinc and copper had higher
464 accumulation and transportation to aerial parts after absorption by the roots. Overall, the study
465 highlights the presence of toxic metals, their sources, and their accumulation patterns in sediments
466 and plant tissues in the studied mangrove forest. The findings provide valuable information for
467 assessing the ecological risk and potential biological effects of these metals in the ecosystem.

468
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470 Software, Visualization, Investigation. Fatemeh Rajaei: Data curation. Reza Dahmardeh Behrooz:
471 Methodology, Writing- Original draft preparation. Giulia Poma: Writing- Reviewing and Editing.
472 All authors have read and agreed to the published version of the manuscript.

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476 **Ethics approval and consent to participate** All procedures performed in this study were in
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479 **Consent for publication** I understand that the text and any pictures published in the article will
480 be freely available on the internet and may be seen by the general public. The pictures, and text
481 may also appear on other websites or in print, may be translated into other languages or used for
482 commercial purposes.

483 **Data availability** Data will be made available on request.

484

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566 **JUNE R. LAWTON, ANN TODD, D. K. NAIDOO** .PRELIMINARY INVESTIGATIONS INTO THE
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