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On the influence of water conductivity, pH and climate on bryophyte assemblages in Catalan seminatural springs

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On the influence of water conductivity, pH and climate 1 on bryophyte assemblages in Catalan semi-natural 2 springs 3 Authors: M. Bes¹, J. Corbera¹, F. Sayol², G. Bagaria², M. Jover³, C. Preece^{2,4}, F. 4 Sabater^{1,4,5}, A. Viza^{1,5}, M. Fernández-Martínez^{*1,6} 5 6 Affiliations: 7 ¹ ICHN, Delegació de la Serralada Litoral Central, Mataró, Catalonia, Spain ² CREAF, Cerdanyola del Vallès, 08193 Barcelona, Catalonia, Spain 8 9 ³ Department of Environmental Sciences, Faculty of Sciences, University of Girona, 10 Girona, Catalonia, Spain ⁴ CSIC, Global Ecology Unit, CREAF-CSIC-UAB, Cerdanyola del Vallès, 08193 11

- 12 Barcelona, Catalonia, Spain
- ⁵ Department of Ecology, University of Barcelona, Barcelona, Catalonia, Spain
- ⁶ Centre of Excellence PLECO (Plant and Vegetation Ecology), Department of Biology,
- 15 University of Antwerp, 2610 Wilrijk, Belgium.

16 ***Corresponding author**

- 17 Marcos Fernández-Martínez
- 18 E-mail address: marcos.fernandez-martinez@uantwerpen.be
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22 **Abstract**

Bryophytes are some of the most sensitive biological indicators of environmental change. 23 Springs have a significant presence of bryophytes and hence they are ideal habitats for 24 studying their relationship with the environment. We tested whether bryophyte 25 26 assemblages can be explained with macro-, meso- and micro-ecological variables (i.e., seasonal climate, altitude, water pH and conductivity) sampling bryophytes from 198 27 semi-natural springs distributed along montane regions in the north-eastern Iberian 28 29 Peninsula. We tested the influence of environmental variables on bryophyte 30 assemblages in springs using sparse Partial Least Squares (sPLS). Our results show that variability in bryophyte assemblages is explained by seasonal climate (temperature 31 and precipitation from winter, spring, summer and autumn and temperature and 32 precipitation seasonality), altitude and water conductivity. The results obtained by the 33 present study will be useful for predicting bryophyte diversity in springs using simple and 34 35 easy to obtain variables such as climate, water pH and conductivity.

36 Introduction

Bryophytes are very sensitive to environmental conditions (Kapfer *et al.*, 2012), mainly because they do not have a well-developed cuticle or epidermis, and hence most of them can absorb water and nutrients through their entire surface. This is why bryophytes are a very interesting group to study the effects of environmental variability and pollution on community composition. In fact, bryophytes have previously been used as bioindicators of air pollution (Slack, 1990; Suren & Ormerod, 1998: Suren & Duncan, 1999) and water quality (Arts, 1990; Vanderpoorten & Klein 1999; Ceschin *et al.*, 2012).

44 Despite the fact that bryophytes are generally more widely distributed than other plants because of their dispersal capacity (Patiño et al., 2015; Wang et al., 2017), species 45 distribution is very dependent on environmental conditions. The most studied variables 46 47 include climate (Gignac & Vitt, 1990; Nicholson et al., 1996), water availability (Pentecost & Zhang, 2006), and geology (Pentecost & Zhaohui, 2002; Belland, 2005; Callaghan & 48 Ashton, 2008a) which, through their interaction, determine more local and potentially 49 50 important environmental variables such as water pH (Tyler et al., 2018) and conductivity in springs and streams where bryophytes can establish (Peñuelas & Sabater, 1987; 51 52 Vanderpoorten & Palm, 1998; Sabater et al., 2015).

One of the habitats that can be dominated by bryophytes is the headwaters of rivers and 53 springs. In these environments, climate, altitude, water pH and conductivity have been 54 55 suggested to determine which species can become established (Tomaselli et al., 2011; 56 Ceschin et al., 2012; Tessler et al., 2014a; Vieira et al., 2014). However, other microand mesoscale environmental variables such as substrate or habitat heterogeneity 57 58 (Suren, 1996; Strohbach et al., 2009; Vieira et al., 2014), aspect and slope (Pentecost & Zhang, 2006), or disturbance regime of floods and droughts may also be factors 59 60 explaining variation in bryophyte assemblages in aquatic and semi-aquatic habitats 61 (Suren & Ormerod, 1998; Suren & Duncan, 1999). In similar habitats, such as peatlands,

some studies have suggested that bryophyte assemblages depend on gradients of 62 temperature and precipitation (Gignac, 1994), water pH and conductivity (Gignac, 1992) 63 64 or shade and permafrost (Belland & Vitt, 1995). Other studies carried out in temperate ecosystems of England (Callaghan & Ashton, 2008a), coastal zones of Canada (Belland, 65 2005), and semi-arid zones of Australia (Eldridge & Tozer, 1997) indicate that the 66 different bryophyte assemblages depended on precipitation, soil pH and geology, 67 68 showing a clear differentiation between communities located over calcareous and non-69 calcareous lithologies (Callaghan & Ashton, 2008b; Spitale et al., 2009; Virtanen et al., 70 2009). Additionally, bryophyte communities have also been suggested to vary along altitudinal gradients (Wolf, 1993; Andrew & Rodgerson, 2003; Bruun et al., 2006; Grytnes 71 72 et al., 2006; Ah-Peng et al., 2007; Grau et al., 2007; Spitale, 2016), which can be a proxy for differences in topology and radiation amongst others. 73

Notwithstanding all this research on bryophyte ecology, the study of differences in 74 75 bryophyte assemblages along gradients of altitude, seasonal temperature and precipitation, water pH and conductivity combined, is still very limited in the region of 76 Catalonia and especially in semi-natural or man-made springs (Corbera et al., 2015). In 77 78 our study region (Catalonia, NE Iberian Peninsula) the ecology of bryophytes has been 79 poorly studied (but see Peñuelas, 1983; Peñuelas & Sabater, 1987), especially concerning the interaction between macro-, meso- and microecological variables. 80 Hence, the aim of this study was to test to what extent macro-, meso- and 81 microecological variables such as climate (seasonal temperatures and precipitation and 82 83 their seasonality), altitude, and water pH and conductivity can explain variation in bryophyte species assemblage in springs within a relatively large climate and water 84 chemistry gradient. To achieve our aim, we surveyed springs and their bryophyte 85 communities in montane regions of north-eastern Iberian Peninsula (Catalonia, NE 86 87 Iberian Peninsula). Based on results provided by previous studies, we hypothesised that water pH and conductivity would play a main role determining bryophyte communities of 88

semi-natural springs, due to the direct contact between the bryophytes and the water, and species differences in tolerance ranges for these properties. In addition, because in the Mediterranean climate, summer is the warmest and driest season of the year, we hypothesised that climate from the summer season (dry and warm season) would also significantly explain variability in species composition of spring bryophyte communities. Altitude was also hypothesised to be an important factor for determining spring bryophyte assemblage as it can reflect differences in topography, radiation and climate.

96 Materials and methods

97 Study area and experimental design

98 We surveyed 198 water springs following a 95 km south-to-north and a 50 km east-to-99 west gradient along the north-eastern Iberian Peninsula territory (Figure 1), comprising 100 4 different mountain ranges: the Central Littoral mountain range, Montseny-Guilleries, 101 Lluçanès and Garrotxa. The springs were unevenly distributed across the different 102 mountain ranges: 101 springs occurred in the Montseny-Guilleries, 55 in the Central 103 Littoral mountain range, 32 in Llucanès and 10 in Garrotxa. Climate also differs amongst these mountain ranges. The Central Littoral mountain range has a typical maritime 104 105 Mediterranean climate with mild summers and winters, while the climate is 106 Mediterranean humid and sub-humid in Montseny-Guilleries and Lluçanès and Mediterranean pre-Pyreneeal in Garrotxa (Martín-Vide, 1992), being more continental. 107 108 Lithologically, the springs of the Central Littoral mountain range are located over different 109 types of granite, except for the eastern side which is dominated by metamorphic rocks 110 (Sabater et al., 2015; Fernández-Martínez et al., 2016). In Montseny, springs are located over granite, metamorphic and calcareous rocks while in Guilleries most of them are over 111 granite. Llucanès is completely calcareous and Garrotxa is divided into calcareous and 112 volcanic rocks. This rich lithology produces a large variability in the characteristics of 113

water from springs, affecting dissolved ions and thus conductivity and pH (Sabater *et al.*,2015).

116 Most of the springs we surveyed are small human-made constructions to collect water 117 from the underground and release it through a spout (Figure S1). Some of the springs 118 drained water mines (collecting water from aquafers) while others were built in naturally 119 occurring springs. However, in all of the surveyed springs, flowing water did not receive 120 any sanitary treatment. The morphology of our springs did not vary greatly (for a typical 121 example see Figure S1). Almost all of them contained the same microhabitats such as a 122 spout, wet rock walls, soil ground/walls and a sink where the water makes a little pool. 123 Surveys of the springs were carried out during spring and autumn seasons of the years 2013 and 2015, and although many other springs were surveyed, only those that had 124 125 water flowing at the time of the survey were sampled and included in this study. We also 126 excluded from analysis any springs whose water flow was controlled by a tap. Therefore, 127 the springs we sampled contained bryophytes that were continuously receiving water during most of the year, apart from springs in which drought had temporarily interrupted 128 water flow. Water pH and conductivity of the flowing water was analysed in the field with 129 pH-meter (ORION, Thermo-Scientific, Waltham), previously calibrated with standard 130 131 solutions at pH 7 and 10, while electric conductivity was measured using a conductivitymeter (WTW, Xylem Inc., Weilheim) calibrated at 25°C and with a standard solution of 132 1314 µS cm⁻¹. 133

To determine the bryophyte communities (as presence or absence data) we took a sample of the different bryophyte species present in the spring that were either in direct contact with the water flow, receiving water drops or under water (see yellow line in Figure S1). We did not standardise our bryophyte measurements by the total area sampled because, (1) sampled area was lower than 0.9 m² for most of the springs, for which differences in scale were minimal, (2) species-area relationship was missing for a subset of our dataset (Figure S2) and, (3) surveying a predetermined area would have made us miss some species present in the springs. Species number per spring was generally very low, most of them having between 1 and 4 species (Figure S3). We used identification keys in Smith (1978 & 1990) and Casas *et al.*, (2001 & 2004) and follow the nomenclature established by Hill *et al.*, (2006). Some of the samples could not be identified to species level (3%, 18 out of 538 samples). To study the variation in bryophyte assemblages we used those bryophyte species that appeared in more than 10 springs (13 species in total), to ensure enough replicates per species.

148 Using the geographical coordinates of the springs, obtained in the field with a GPS, we 149 extracted average seasonal (winter, spring, summer and autumn) temperatures and 150 precipitation, as well as mean annual temperature (MAT) and precipitation (MAP), from the Digital Climatic Atlas of Catalonia (Pons, 1996; Ninyerola et al., 2000; available at 151 152 http://www.opengis.uab.cat/acdc/index.htm) with a resolution of 180 m. Seasonal climate 153 was calculated as the average temperature and total precipitation values of December to February for winter, March to May for spring, June to August for summer and 154 September to November for autumn. Using seasonal climate, we also calculated 155 156 temperature and precipitation seasonality as the coefficient of variation of seasonal temperatures and precipitation. Altitude was obtained using a GPS in the field. 157

158 Statistical analyses

159 To test whether bryophyte assemblages depend on environmental conditions we performed a sparse Partial Least Squares (sPLS) analysis (Lê Cao et al., 2008), fitted 160 by the regression mode, where water pH and conductivity, seasonal temperatures and 161 precipitation, temperature and precipitation seasonality, and altitude were the 162 environmental matrix of predictors and the binary variables of bryophyte species 163 presence were the response matrix to be predicted. Because the response matrix 164 165 (species presence) was a binary variable we fitted the model after applying a log-ratio 166 transformation. PLS family methods work by finding the underlying relationships between

two matrices (X and Y, predictors and responses) by extracting latent variables to model 167 the covariance structures of the two spaces. PLS models attempt to estimate axes or 168 169 variates in the X matrix that explains the maximum variance in Y matrix. These methods are particularly useful when the X matrix has more variables than observations, and 170 when high multicollinearity is present among X values. Additionally, sparse PLS methods 171 include a process of selection of the predictor variables to facilitate biological 172 173 interpretation of the results. The sPLS model was fitted using the "spls" function in mixOmics R package (Le Cao et al., 2017). Tuning of the model suggested that 174 extracting more than two variates did not improve the prediction of the response matrix 175 (total Q value saturates at the second variate extracted). To visualise the results of the 176 177 sPLS analysis, we plotted a Clustered Image Map (CIM), using the "cim" function in mixOmics, which shows the correlations between the predictor and the response 178 179 variables while clustering both based on their similarity on the multivariate space.

180 To look for significant differences of the variables selected by the sPLS analysis among species, we performed analyses of variance (ANOVA) in which we related the 181 environmental variables selected in the sPLS as a function of the species. This analysis 182 allows us to estimate ranges of tolerance for the species and to test whether different 183 184 species share their ecological niche or not. Differences amongst species were tested using the post-hoc Tukey HSD test. These analyses were done with the 82 springs in 185 186 which at least one of the most frequent species was found. All ANOVAs accomplished the required statistical assumptions of normality, homoscedasticity and independence of 187 188 the residuals. All the statistical analysis was performed using R statistical software (R Core Team, 2015). 189

190 **Results**

191 Water chemistry and climate of the studied springs

192 We found a relatively large gradient in water chemistry and climatological features of the 193 surveyed springs. Mean annual temperature (MAT) ranged from 7.3 to 15.7°C, mean 194 annual precipitation (MAP) from 555 to 1180 mm year⁻¹, pH from 5.7 to 8.8, conductivity 195 from 19 to 2160 µS cm⁻¹ and altitude from 100 to 1570 m a.s.l. The springs of the Littoral 196 mountain range are located at lower altitudes, being on average, 328 m above sea level (a.s.l.) and therefore, experienced the highest temperatures (13.9°C on average) (Table 197 198 1). Springs from Lluçanès, Montseny-Guilleries and Garrotxa are found at intermediate 199 altitudes, between 550 and 850 m a.s.l., and recorded temperatures between 11 and 200 12.5°C, although some of Montseny springs experienced relatively high temperatures 201 (up to 15.7°C). Annual precipitation in La Garrotxa is higher than in any other of the 202 studied regions (1049 mm), followed by Montseny-Guilleries, whereas Llucanès and the 203 Littoral mountain range are drier (Table 1).

In general, pH of the springs surveyed was slightly alkaline. However, pH in Garrotxa's springs is higher than those from the other regions studied (Table 1). Conductivity, though, is much more variable than pH. Lluçanès' springs are, on average, those with highest conductivity (1059 μ S·cm⁻¹), whereas springs from the Littoral mountain range and Garrotxa are found in the mid-range (around 700 μ S·cm⁻¹), and those from Montseny-Guilleries have the lowest conductivity values of all (286 μ S·cm⁻¹) (Table 1).

210 Relationships between species presence and environmental variables

The sPLS model included conductivity, altitude, spring, summer and autumn temperature, winter, spring, summer and autumn precipitation, and temperature and precipitation seasonality as important predictors of bryophyte species assemblages in springs (Figure 2). The first variate explained 51.1% of the variability in the predictors and 12.9% of the variability of the bryophyte assemblages. As shown by Figure S4, no single species or spring was driving the results of our sPLS model. This variate was positively related to seasonal precipitation, altitude, and temperature seasonality and negatively related to conductivity, seasonal temperatures and precipitation seasonality
(Figure S5). The second variate explained 28.1% of the variability in the predictors and
11.0% of the variability of the bryophyte assemblages. This variate was positively related
mainly to summer precipitation and altitude and negatively related mainly to seasonal
temperatures and precipitation seasonality.

223 Species were clustered into two big clusters (1 and 2) with two smaller clusters each (a 224 and b, Figure 2). The first cluster of species (1a), formed by Eucladium verticillatum, 225 Pohlia melanodon, Didymodon tophaceus and Pellia endiviifolia, was found to be related 226 to warm and especially dry climate and high water conductivity. Their presence was also 227 negatively related to altitude of the springs and temperature seasonality and, less strongly, positively related to precipitation seasonality. Pallustriella commutata and 228 229 Cratoneuron filicinum, which formed cluster 1b, were found to be associated with dry and cold climate with low temperature and precipitation seasonality, but with rainy summers 230 231 and located at high altitude. Pallustriella commutata was also positively related to high water conductivity, while no relationship was found with *C. filicinum*. 232

233 Kindbergia praelonga, Oxyrrhynchium speciosum and Conocephalum conicum formed 234 cluster 2a and were more likely to be found in springs that have a warm and relatively dry climate in spring, and especially in summer, and high water conductivity (except C. 235 conicum). The species of this cluster were also found in springs of low altitude and low 236 seasonal variability of temperature and precipitation (low continentality). The last cluster 237 238 of species (2b) was formed by Plagiomnium undulatum, Brachythecium rivulare, Bryum pseudotriquetrum, and Platyhypnidium riparioides. These species were the group of 239 mosses related to the springs with the coldest and wettest climate, highest temperature 240 seasonality but lowest precipitation seasonality, and highest altitude and having also low 241 242 water conductivity. Plagiomnium undulatum, however, was not so strongly related to cold 243 climate.

Of all studied species, *P. endiviifolia and C. conicum* were the species with the lower correlations (Figure 2). Table 3 shows the average values per species for the environmental variables included in the sPLS. Significant differences appear amongst species for all of the variables, which further corroborates results from the sPLS. This indicates that ecological niche separation occurs within the environmental variables considered.

250 **Discussion**

251 Our results show that macroclimate, altitude and water conductivity can explain variability 252 in bryophyte assemblages in semi-natural and man-made springs. The distribution of some of the most abundant bryophyte species in our springs is clearly separated into 253 254 different ecological niches, defined by the environmental variables we considered. In 255 particular, our findings suggest that models predicting species distributions could be improved by including micro- and mesoscale environmental variables such as water 256 257 conductivity and altitude in addition to macroscale variables such as climate. However, other environmental variables considered in previous studies (Longton et al., 1983; 258 259 Cattaneo & Fortin, 2000; Tessler et al., 2014; Corbera et al., 2015), such as main ions (Ca²⁺, Na⁺, Mg²⁺, K⁺, SO₄²⁻, Cl⁻, PO₄³⁻), pollutants such as nitrate (Vanderpoorten & Klein, 260 1999), or type of microhabitat and its heterogeneity (Suren, 1996) could help to 261 differentiate the ecological niche of bryophyte species even further. Despite that, the 262 263 present study clearly shows several general trends in bryophyte assemblages in springs.

264 Determinants of bryophyte assemblages

The role of precipitation on bryophyte assemblages is evident in previous studies (Eldridge & Tozer, 1997; Gignac, 2001; Callaghan & Ashton, 2008a) although some other papers highlight that the number of rainy days per year is sometimes more important than the total amount of precipitation (Ratcliffe, 1968; Callaghan & Ashton, 2008b) because the time that the bryophytes are dry is more determinant than the total 270 amount of water they receive (Pentecost & Zhang, 2006; Proctor et al., 2007). However, 271 this fact may not be so relevant in our study, because most of the sampled springs 272 provide water almost constantly throughout the year. We found that B. rivulare, P. riparioides, B. pseudotriquetrum, which are species associated with streams and 273 274 believed not to tolerate desiccation, are those with the highest affinity for high precipitation (Figure 2, Table 2). However, other species with supposedly equally high 275 276 affinity for humidity like P. endiviifolia, P. commutata, C. filicinum and E. verticillatum 277 (Table 2), do not show such a strong positive association with precipitation, but even 278 show the opposite (Figure 2).

279 Kapfer et al. (2012) suggested that the length of time water resides in the aquifers of zones with lower precipitation is longer than in zones with higher precipitation (). A longer 280 281 time in contact with the rock causes the water to dissolve more salts and increase its 282 conductivity. This could explain the presence of known calcicolous species, associated 283 with high conductivity water, in springs with low precipitation (Corbera et al., 2015) (Figure 2). Despite the fact that bryophytes are able to tolerate wide ranges of 284 285 temperature, we found different average seasonal temperatures and temperature seasonality (which relates to continentality) per species. This effect of temperature on 286 287 bryophyte assemblages may also be mediated through its effect on the water availability of the spring. Almost all of our springs had forested areas nearby and which typically 288 consume large quantities of subsoil water, especially under high temperatures 289 (Fernández-Martínez et al., 2014), thus reducing water runoff. Also, continentality 290 291 increases temperature extremes throughout the year, which can certainly be an 292 important factor favouring certain species versus others, depending on their temperature 293 tolerance ranges.

Some studies have shown a great polarization in the distribution of bryophytes in relation
to calcareous and non-calcareous lithology (Callaghan & Ashton, 2008a; Virtanen *et al.*,
2009) because of big differences in the cellular exchange capacity of Ca²⁺ (Bates, 1982).

297 In our study, species with affinity for calcareous lithology, like D. tophaceus and E. 298 verticillatum (Table 2), have been found in springs with high water conductivity (on 299 average, 972 and 886 μ S cm⁻¹ respectively) while calcifugous species like *B. rivulare* (Longton et al., 1983) have been found in springs with low water conductivity (292 µS·cm⁻ 300 301 ¹). However, in contrast to other studies carried out with bryophyte communities from peat lands and in the headwaters of rivers (Gignac, 1992; Tessler et al., 2014), we did 302 303 not find any significant relationship between water pH and bryophyte assemblages 304 (Figure 2). This fact may be the result of having analysed only the 13 most frequent bryophyte species that were found in the springs. As these species are more frequent, 305 306 they are also supposed to be more likely to be generalists and to tolerate wider ranges 307 of water pH. Further work, based on more intensive sampling and focused on less 308 frequent species, will help elucidate how water pH, conductivity and other environmental 309 variables (e.g., climate, topography and ionic composition of water) affect bryophyte assemblages in Mediterranean springs. However, our results indicate that water 310 311 conductivity could be a very suitable variable to include in species distribution models to improve their predictions on humid or sub-humid bryophytes, given the fact that it is 312 relatively easy to obtain (it could be inferred from lithology) and significantly separates 313 bryophyte species. 314

315 **Conclusions**

Our results clearly demonstrate that bryophyte assemblages in semi-natural springs can be explained using widely available variables such as mean seasonal temperatures and precipitation, temperature and precipitation seasonality (intra-annual variation), altitude, and water conductivity of the springs. Our results also point out the need to include microand mesoscale variables, in addition to macroscale variables to help improve the prediction of species distribution models.

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328

329 Taxonomic Additions and Changes: Nil.

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506 Figure captions

Figure 1. Map showing the location of the surveyed springs (white dots).

510	Figure 2. Clustered image map of the sPLS model showing the correlations between
511	environmental predictors and presence of bryophyte species. Species and predictors
512	have been clustered according to their similarity forming groups that we split using black
513	lines. Abbreviations: T indicates temperature, P indicates precipitation, and seasons
514	were indicated in lowercase letters as winter (w), spring (sp), summer (sm) and autumn
515	(a). PS and TS indicate seasonality of precipitation and temperature respectively.

Figure 1.



520 Figure 2



Table 1. Average values for altitude, climate (mean ± standard error of the mean [SE]), water pH
and conductivity and bryophyte species richness of the springs and average values per region.
Lluçanès is a calcareous region, Montseny and the Central Littoral mountain range have granitic
and metamorphic lithology and Garrotxa springs are located over volcanic and calcareous rocks.
Acronyms are MAT for mean annual temperature and MAP for mean annual precipitation. Units
are degrees Celsius for MAT, mm·year⁻¹ for MAP, µS·cm⁻¹ for conductivity and metres for altitude.

	Total	Lluçanès	Montseny - Guilleries	Central Littoral	Garrotxa
Springs	198	32	101	55	10
МАТ	12.01 ± 0.13	11.38 ± 0.12	11.18 ± 0.30	13.90 ± 0.16	12.11 ± 0.52
МАР	863 ± 11	766 ± 22	947 ± 19	728 ± 14	1049 ± 31
Altitude	660 ± 24	703 ± 29	828 ± 53	328 ± 25	640 ± 109
рН	7.17 ± 0.04	7.10 ± 0.09	7.18 ± 0.11	7.09 ± 0.10	7.65 ± 0.13
Conductivity	546 ± 32	1059 ± 100	286 ± 50	701 ± 74	688 ± 186
Sp. richness	2.71 ± 0.12	3.16 ± 0.44	2.75 ± 0.23	2.05 ± 0.21	4.27 ± 1.31

Table 2. List of bryophyte species found in ten or more springs and their habitat preferences according to (Casas, 1958 & 1959; Casas et al., 2001 & 2004;
 Atherton et al., 2010) and their ecological characterization following Dierßen (2001). Column M/L indicates whether species are mosses (M) or liverworts (L).
 Column N indicates the number of springs in which each species has been found. Substrate: h.a., highly acidophilous; c.a., considerably acidophilous; m.a.,
 moderately acidophilous; s.n., subneutrophylous; b., basophilous. Humidity: h.h., highly hygrophilic; c.h., considerably hygrophilic; m.h., moderately hygrophilic;
 m., mesophilous; m.x., moderately xerophilous; c.x., considerably xerophilous; h.x., highly xerophilous.

	M/L	Ν	Preferred habitat	h.a.	c.a.	m.a.	s.n.	b.	h.h.	c.h.	m.h.	m m	.x. (c.x.	h.x.
Oxyrrhynchium speciosum (Brid.) Warnst.	М	64	Wet forests, peat lands and springs												
Pellia endiviifolia (Dicks.) Dum.	L	57	Edge of streams, shaded forests, peatlands and wet rocks												
Eucladium verticillatum (With.) Bruch & Schimp.	М	50	Edge of streams, wet and shaded locations. Calcareous water				_	11							
Cratoneuron filicinum (Hedw.) Spruce	М	44	Mixed habitats: edge of streams, rocks and barks												
Plagiomnium undulatum (Hedw.) T.J.Kop.	М	36	Wet rocks, forests, grasslands. Occasionally in streams												
Platyhypnidium riparioides (Hedw.) Dixon	М	24	Rocks and roots underwater or semi-underwater in streams												
Brachythecium rivulare Schimp.	М	20	Aquatic environment. Soils and rocks in streams												
Didymodon tophaceus (Brid.) Lisa	М	20	Wet rocks, calcareous soils, muddy edges of bogs and streams												
Palustriella commutata (Hedw.) Ochyra	М	19	Wet habitats like springs. Calcareous substrates.												
Conocephalum conicum (L.) Underw.	L	13	Wet rocks and soils, usually in contact with water.												
Bryum pseudotriquetrum (Hedw.) P.Gaertn. et al.	М	12	Edge of rivers, streams or lakes												
Pohlia melanodon (Brid.) A.J.Shaw	М	12	Wet and muddy soils. Usually on the edge of streams												
Kindbergia praelonga (Hedw.) Ochyra	М	11	Grasslands, forests, bark and branches and edges of water streams						1						

Substrate

Humidity

535 Table 3. Species average (± standard error of the mean) values for environmental variables that 536 were selected in the sPLS for explaining variability in species distribution. Letters (a, b, c, d, e, f) 537 indicate groups according to the post-hoc Tukey HSD test for multiple comparisons. All ANOVA 538 tables were statistically significant at the <0.001 level. Units were metres for altitude, mm year¹ for precipitation variables and Celsius degrees for temperature variables and (μ S·cm⁻¹) for 539 540 conductivity. Precipitation seasonality (PS) and temperature seasonality (TS) were unit less. 541 Abbreviations: T indicates temperature, P indicates precipitation, and seasons were indicated in 542 lowercase letters as winter (w), spring (sp), summer (sm) and autumn (a).

Species	Tsp		Tsm		Та	
Bryum pseudotriquetrum	6.98 ± 0.58	а	16.1 ± 0.6	а	10.2 ± 0.5	а
Brachythecium rivulare	7.50 ± 0.45	а	16.6 ± 0.5	а	10.6 ± 0.4	ab
Platyhypnidium riparioides	7.72 ± 0.41	ab	16.7 ± 0.4	ab	10.7 ± 0.4	abc
Plagiomnium undulatum	9.45 ± 0.32	bc	18.5 ± 0.3	bc	12.2 ± 0.3	cd
Cratoneuron filicinum	9.58 ± 0.30	cd	18.8 ± 0.3	С	12.3 ± 0.3	d
Oxyrrhynchium speciosum	10.15 ± 0.25	cdc	19.2 ± 0.3	cd	13.0 ± 0.2	def
Palustriella commutata	9.88 ± 0.46	cdc	19.4 ± 0.5	cd	12.5 ± 0.4	bcde
Pellia endiviifolia	10.45 ± 0.27	cdc	19.6 ± 0.3	cd	13.2 ± 0.2	def
Conocephalum conicum	11.03 ± 0.56	cdc	20.0 ± 0.6	cd	13.6 ± 0.50	def
Pohlia melanodon	10.88 ± 0.58	cdc	20.1 ± 0.6	cd	13.6 ± 0.5	def
Eucladium verticillatum	11.06 ± 0.28	с	20.2 ± 0.3	d	13.7 ± 0.3	ef
Didymodon tophaceus	10.91 ± 0.45	cdc	20.3 ± 0.5	cd	13.5 ± 0.4	def
Kindbergia praelonga	11.84 ± 0.61	dc	20.5 ± 0.6	cd	14.8 ± 0.6	f
	Pw		Psm		Ра	
Kindbergia praelonga	Pw 159.2 ± 11.8	abc	Psm 150.9 ± 15.7	a	Pa 252.5 ± 10.7	abcd
Kindbergia praelonga Oxyrrhynchium speciosum	Pw 159.2 ± 11.8 182.1 ± 4.9	abc bcd	Psm 150.9 ± 15.7 175.0 ± 6.5	a ab	Pa 252.5 ± 10.7 263.1 ± 4.4	abcd bc
Kindbergia praelonga Oxyrrhynchium speciosum Pohlia melanodon	Pw 159.2 ± 11.8 182.1 ± 4.9 150.8 ± 11.3	abc bcd ab	Psm 150.9 ± 15.7 175.0 ± 6.5 183.2 ± 15.0	a ab abc	Pa 252.5 ± 10.7 263.1 ± 4.4 233.6 ± 10.2	abcd bc ab
Kindbergia praelonga Oxyrrhynchium speciosum Pohlia melanodon Conocephalum conicum	Pw 159.2 ± 11.8 182.1 ± 4.9 150.8 ± 11.3 196.5 ± 10.9	abc bcd ab bcde	Psm 150.9 ± 15.7 175.0 ± 6.5 183.2 ± 15.0 188.5 ± 14.4	a ab abc abcd	Pa 252.5 ± 10.7 263.1 ± 4.4 233.6 ± 10.2 267.8 ± 9.8	abcd bc ab abcde
Kindbergia praelonga Oxyrrhynchium speciosum Pohlia melanodon Conocephalum conicum Plagiomnium undulatum	Pw 159.2 ± 11.8 182.1 ± 4.9 150.8 ± 11.3 196.5 ± 10.9 201.4 ± 6.3	abc bcd ab bcde cde	Psm 150.9 ± 15.7 175.0 ± 6.5 183.2 ± 15.0 188.5 ± 14.4 194.3 ± 8.3	a ab abc abcd abc	Pa 252.5 ± 10.7 263.1 ± 4.4 233.6 ± 10.2 267.8 ± 9.8 278.0 ± 5.7	abcd bc ab abcde cde
Kindbergia praelonga Oxyrrhynchium speciosum Pohlia melanodon Conocephalum conicum Plagiomnium undulatum Brachythecium rivulare	$\begin{array}{c} \textbf{Pw} \\ 159.2 \pm 11.8 \\ 182.1 \pm 4.9 \\ 150.8 \pm 11.3 \\ 196.5 \pm 10.9 \\ 201.4 \pm 6.3 \\ 220.0 \pm 8.8 \end{array}$	abc bcd ab bcde cde e	Psm 150.9 ± 15.7 175.0 ± 6.5 183.2 ± 15.0 188.5 ± 14.4 194.3 ± 8.3 196.7 ± 11.6	a ab abc abcd abc abc	Pa 252.5 ± 10.7 263.1 ± 4.4 233.6 ± 10.2 267.8 ± 9.8 278.0 ± 5.7 293.7 ± 7.9	abcd bc ab abcde cde de
Kindbergia praelonga Oxyrrhynchium speciosum Pohlia melanodon Conocephalum conicum Plagiomnium undulatum Brachythecium rivulare Eucladium verticillatum	$\begin{array}{c} \textbf{Pw} \\ 159.2 \pm 11.8 \\ 182.1 \pm 4.9 \\ 150.8 \pm 11.3 \\ 196.5 \pm 10.9 \\ 201.4 \pm 6.3 \\ 220.0 \pm 8.8 \\ 146.3 \pm 5.5 \end{array}$	abc bcd ab bcde cde e a	Psm 150.9 ± 15.7 175.0 ± 6.5 183.2 ± 15.0 188.5 ± 14.4 194.3 ± 8.3 196.7 ± 11.6 199.2 ± 7.4	a ab abc abcd abc abcd abcd abc	Pa 252.5 ± 10.7 263.1 ± 4.4 233.6 ± 10.2 267.8 ± 9.8 278.0 ± 5.7 293.7 ± 7.9 234.9 ± 5.0	abcd bc ab abcde cde de a
Kindbergia praelonga Oxyrrhynchium speciosum Pohlia melanodon Conocephalum conicum Plagiomnium undulatum Brachythecium rivulare Eucladium verticillatum Pellia endiviifolia	$\begin{array}{c} \textbf{Pw} \\ 159.2 \pm 11.8 \\ 182.1 \pm 4.9 \\ 150.8 \pm 11.3 \\ 196.5 \pm 10.9 \\ 201.4 \pm 6.3 \\ 220.0 \pm 8.8 \\ 146.3 \pm 5.5 \\ 177.5 \pm 5.2 \end{array}$	abc bcd ab bcde cde e a bc	Psm 150.9 ± 15.7 175.0 ± 6.5 183.2 ± 15.0 188.5 ± 14.4 194.3 ± 8.3 196.7 ± 11.6 199.2 ± 7.4 200.6 ± 6.9	a ab abc abcd abc abcd abc abc	Pa 252.5 ± 10.7 263.1 ± 4.4 233.6 ± 10.2 267.8 ± 9.8 278.0 ± 5.7 293.7 ± 7.9 234.9 ± 5.0 260.4 ± 4.7	abcd bc ab abcde cde de a bc
Kindbergia praelonga Oxyrrhynchium speciosum Pohlia melanodon Conocephalum conicum Plagiomnium undulatum Brachythecium rivulare Eucladium verticillatum Pellia endiviifolia Didymodon tophaceus	$\begin{array}{c} \textbf{Pw} \\ 159.2 \pm 11.8 \\ 182.1 \pm 4.9 \\ 150.8 \pm 11.3 \\ 196.5 \pm 10.9 \\ 201.4 \pm 6.3 \\ 220.0 \pm 8.8 \\ 146.3 \pm 5.5 \\ 177.5 \pm 5.2 \\ 138.6 \pm 8.8 \end{array}$	abc bcd ab bcde cde e a bc a	$\begin{array}{c} \textbf{Psm} \\ 150.9 \pm 15.7 \\ 175.0 \pm 6.5 \\ 183.2 \pm 15.0 \\ 188.5 \pm 14.4 \\ 194.3 \pm 8.3 \\ 196.7 \pm 11.6 \\ 199.2 \pm 7.4 \\ 200.6 \pm 6.9 \\ 210.0 \pm 11.6 \end{array}$	a ab abc abcd abc abcd abc abc abc abc	Pa 252.5 ± 10.7 263.1 ± 4.4 233.6 ± 10.2 267.8 ± 9.8 278.0 ± 5.7 293.7 ± 7.9 234.9 ± 5.0 260.4 ± 4.7 226.6 ± 7.9	abcd bc ab abcde cde de a bc a
Kindbergia praelonga Oxyrrhynchium speciosum Pohlia melanodon Conocephalum conicum Plagiomnium undulatum Brachythecium rivulare Eucladium verticillatum Pellia endiviifolia Didymodon tophaceus Platyhypnidium riparioides	$\begin{array}{c} \textbf{Pw} \\ 159.2 \pm 11.8 \\ 182.1 \pm 4.9 \\ 150.8 \pm 11.3 \\ 196.5 \pm 10.9 \\ 201.4 \pm 6.3 \\ 220.0 \pm 8.8 \\ 146.3 \pm 5.5 \\ 177.5 \pm 5.2 \\ 138.6 \pm 8.8 \\ 219.2 \pm 8.0 \end{array}$	abc bcd ab bcde cde e a bc a e	$\begin{array}{r} \textbf{Psm} \\ 150.9 \pm 15.7 \\ 175.0 \pm 6.5 \\ 183.2 \pm 15.0 \\ 188.5 \pm 14.4 \\ 194.3 \pm 8.3 \\ 196.7 \pm 11.6 \\ 199.2 \pm 7.4 \\ 200.6 \pm 6.9 \\ 210.0 \pm 11.6 \\ 216.4 \pm 10.6 \end{array}$	a ab abc abcd abc abc abc abc bcd	Pa 252.5 ± 10.7 263.1 ± 4.4 233.6 ± 10.2 267.8 ± 9.8 278.0 ± 5.7 293.7 ± 7.9 234.9 ± 5.0 260.4 ± 4.7 226.6 ± 7.9 295.6 ± 7.2	abcd bc ab abcde cde de a bc a e
Kindbergia praelonga Oxyrrhynchium speciosum Pohlia melanodon Conocephalum conicum Plagiomnium undulatum Brachythecium rivulare Eucladium verticillatum Pellia endiviifolia Didymodon tophaceus Platyhypnidium riparioides Bryum pseudotriquetrum	$\begin{array}{c} \textbf{Pw} \\ 159.2 \pm 11.8 \\ 182.1 \pm 4.9 \\ 150.8 \pm 11.3 \\ 196.5 \pm 10.9 \\ 201.4 \pm 6.3 \\ 220.0 \pm 8.8 \\ 146.3 \pm 5.5 \\ 177.5 \pm 5.2 \\ 138.6 \pm 8.8 \\ 219.2 \pm 8.0 \\ 221.1 \pm 11.3 \end{array}$	abc bcd ab bcde cde e a bc a bc a e de	$\begin{array}{c} \textbf{Psm} \\ 150.9 \pm 15.7 \\ 175.0 \pm 6.5 \\ 183.2 \pm 15.0 \\ 188.5 \pm 14.4 \\ 194.3 \pm 8.3 \\ 196.7 \pm 11.6 \\ 199.2 \pm 7.4 \\ 200.6 \pm 6.9 \\ 210.0 \pm 11.6 \\ 216.4 \pm 10.6 \\ 218.9 \pm 15.0 \end{array}$	a ab abc abcd abc abc abc abc bcd abcd	Pa 252.5 ± 10.7 263.1 ± 4.4 233.6 ± 10.2 267.8 ± 9.8 278.0 ± 5.7 293.7 ± 7.9 234.9 ± 5.0 260.4 ± 4.7 226.6 ± 7.9 295.6 ± 7.2 303.3 ± 10.2	abcd bc ab abcde cde de a bc a e e
Kindbergia praelonga Oxyrrhynchium speciosum Pohlia melanodon Conocephalum conicum Plagiomnium undulatum Brachythecium rivulare Eucladium verticillatum Pellia endiviifolia Didymodon tophaceus Platyhypnidium riparioides Bryum pseudotriquetrum Cratoneuron filicinum	$\begin{array}{c} \textbf{Pw} \\ 159.2 \pm 11.8 \\ 182.1 \pm 4.9 \\ 150.8 \pm 11.3 \\ 196.5 \pm 10.9 \\ 201.4 \pm 6.3 \\ 220.0 \pm 8.8 \\ 146.3 \pm 5.5 \\ 177.5 \pm 5.2 \\ 138.6 \pm 8.8 \\ 219.2 \pm 8.0 \\ 221.1 \pm 11.3 \\ 191.3 \pm 5.9 \end{array}$	abc bcd ab bcde cde e a bc a bc a e de bcde	$\begin{array}{r} \textbf{Psm} \\ 150.9 \pm 15.7 \\ 175.0 \pm 6.5 \\ 183.2 \pm 15.0 \\ 188.5 \pm 14.4 \\ 194.3 \pm 8.3 \\ 196.7 \pm 11.6 \\ 199.2 \pm 7.4 \\ 200.6 \pm 6.9 \\ 210.0 \pm 11.6 \\ 216.4 \pm 10.6 \\ 218.9 \pm 15.0 \\ 221.7 \pm 7.8 \end{array}$	a ab abc abcd abc abcd abc abcd bcd abcd cd	Pa 252.5 ± 10.7 263.1 ± 4.4 233.6 ± 10.2 267.8 ± 9.8 278.0 ± 5.7 293.7 ± 7.9 234.9 ± 5.0 260.4 ± 4.7 226.6 ± 7.9 295.6 ± 7.2 303.3 ± 10.2 269.5 ± 5.3	abcd bc ab abcde cde de a bc a e e e bcde

Table 3. Continuation.

Species	Psp		PS		TS	
Kindbergia praelonga	18.10 ± 1.01	а	0.25 ± 0.012	cd	0.40 ± 0.03	а
Pohlia melanodon	19.88 ± 0.96	abc	0.24 ± 0.011	abcd	0.48 ± 0.02	abc
Eucladium verticillatum	20.03 ± 0.47	ab	0.25 ± 0.006	d	0.48 ± 0.01	ab
Didymodon tophaceus	20.03 ± 0.75	ab	0.25 ± 0.009	d	0.50 ± 0.02	abc
Oxyrrhynchium speciosum	21.24 ± 0.42	abc	0.23 ± 0.005	bcd	0.50 ± 0.01	bc
Pellia endiviifolia	22.06 ± 0.44	bcd	0.22 ± 0.005	abcd	0.50 ± 0.01	bc
Conocephalum conicum	22.45 ± 0.93	abcdc	0.21 ± 0.011	abcd	0.48 ± 0.02	abc
Palustriella commutata	22.63 ± 0.77	bcdc	0.24 ± 0.009	abcd	0.54 ± 0.02	bcde
Plagiomnium undulatum	23.41 ± 0.53	cdc	0.21 ± 0.006	abc	0.53 ± 0.01	bcd
Cratoneuron filicinum	23.96 ± 0.50	dc	0.20 ± 0.006	а	0.54 ± 0.01	cd
Brachythecium rivulare	24.89 ± 0.75	dc	0.20 ± 0.009	ab	0.61 ± 0.02	de
Platyhypnidium riparioides	25.36 ± 0.68	с	0.20 ± 0.008	ab	0.60 ± 0.02	de
Bryum pseudotriquetrum	25.84 ± 0.96	с	0.20 ± 0.011	abc	0.64 ± 0.02	е

Species	Conductivity		Altitude	
Platyhypnidium riparioides	227.9 ± 82.4	а	893.5 ± 54.3	cd
Brachythecium rivulare	291.9 ± 90.3	ab	890.9 ± 59.4	cd
Plagiomnium undulatum	300.0 ± 64.7	а	670.8 ± 42.6	abc
Bryum pseudotriquetrum	300.7 ± 116.6	ab	1077.6 ± 76.7	d
Conocephalum conicum	429.5 ± 112.0	ab	448.7 ± 3.7	а
Oxyrrhynchium speciosum	505.1 ± 50.5	ab	551.7 ± 33.2	а
Cratoneuron filicinum	560.3 ± 60.9	ab	755.5 ± 40.1	bc
Kindbergia praelonga	582.3 ± 121.8	abc	380.5 ± 80.1	а
Pellia endiviifolia	611.9 ± 53.5	b	589.6 ± 35.2	ab
Palustriella commutata	717.9 ± 92.7	bc	669.2 ± 61.0	abc
Eucladium verticillatum	885.5 ± 57.1	С	516.7 ± 37.6	а
Didymodon tophaceus	971.9 ± 90.3	С	555.2 ± 59.4	ab
Pohlia melanodon	1122.3 ± 116.6	с	558.6 ± 76.7	ab

548 Supplementary information

- **Figure S1:** Picture of a spring surveyed. The yellow line indicates the sampling zone, coinciding
- 550 with the area under the influence of water.



Figure S2: Graph showing species richness as a function of the area sampled in a subset of 41
sampled springs. Units of sample area are m⁻². No significant correlation between species number
and area was found.





Figure S3. Histogram showing species richness of the studied springs.

- **Figure S4:** Site scores of the sPLS relating environmental variables (predictors) and bryophyte
- 560 presence (species). None of the variates were driven by single sites or species.



- 562 **Figure S5:** Correlation circle plot showing the results of the sPLS model relating environmental
- 563 variables (predictors in blue) and bryophyte presence (species in green). Prec: precipitation,

564 Temp: temperature.

